

1 **Running title:** temperature influences high-altitude tadpoles' growth

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3 **Effects of temperature on growth and development of amphibian larvae across**
4 **an altitudinal gradient in the Tibetan Plateau**

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17

18 **Abstract**

19 Organisms living in extreme environments, such as amphibians inhabiting the Tibetan
20 Plateau, are faced with a magnitude of potentially strong selection pressures. With an
21 average elevation exceeding 4,500 m, the Tibetan Plateau is mainly characterized by
22 low temperatures, but little is known about the influence of this factor on the growth,
23 development, and behavior of amphibian larvae living in this environment. Using a
24 common garden experiment, we studied the influence of temperatures on the early
25 growth and development of tadpoles of the Tibetan brown frog (*Rana kukunoris*)
26 endemic to the eastern Tibetan plateau. We discovered that temperature had a
27 significant influence on early growth and development of the tadpoles from
28 high-temperature treatment, growing and developing faster than their siblings from
29 low-temperature treatment. However, high-altitude individuals grew faster than
30 low-altitude individuals at low temperatures, while the opposite was true at high
31 temperatures. These results support the temperature adaptation hypothesis as tadpoles'
32 growth and developmental rates were maximized at the temperatures experienced in
33 their native environments. These results suggest that variation in ambient temperature,
34 combined with evolutionary adaptation to temperature of local environments, is
35 probably one of the most critical environmental factors shaping altitudinal differences
36 of the growth and development of amphibian larvae at the Tibetan plateau.

37

38 **Key words**

39 Development; growth; *Rana kukunoris*; survival rate; temperature

40 **Introduction**

41 Life history theories predict that environment conditions can pose strong influences
42 on evolution of life history traits of animals, including amphibians (Stearns, 1992;
43 Morrison & Hero, 2003a; Liao et al., 2014; Liao et al., 2015; Zhong et al., 2018;
44 Wang et al., 2019; Yu et al., 2019), and that environmental conditions experienced
45 during early life can have lasting effects on later life characteristics of individuals
46 (Burton & Metcalfe, 2014; Garcia et al., 2017). Although organisms such as
47 amphibians can alleviate the negative fitness consequences of harsh environmental
48 conditions through behavioral and physiological plasticity (Voituron et al., 2005; Muir
49 et al., 2014a), there is growing evidence to suggest that temperature lead to
50 evolutionary adaptations in amphibian life history traits, including the growth and
51 development of their larvae (e.g. Wells, 1977; Morrison & Hero, 2003b). However,
52 relatively little is known about the importance of this stressor, and its possible effects
53 on the evolution of amphibian life histories, especially in harsh high-altitude
54 environments.

55 Previous studies show that temperature has markedly affect amphibian life
56 history traits including embryonic growth and development rates (Berven, 1982;
57 Gollman & Gollman, 1996; Laugen et al., 2003), and body size at metamorphosis
58 (Morrison & Hero, 2003a). The tadpoles from cold environments, between such
59 high-altitude and -latitude localities, generally have longer larval periods because of
60 slow growth rates and finally result in larger body sizes at metamorphosis than those
61 from warmer areas (e.g. Berven, 1982; Howard & Wallace, 1985; Morrison & Hero,

62 2003a; Liao & Lu 2012; Liu et al. 2018). This variation in growth and development
63 rates has been attributed mainly to environmental temperature in that it will control
64 developmental and physiological processes of organisms (Gollman & Gollman, 1996).
65 The study results of a large volume of literature (e.g. Levinton & Monahan, 1983;
66 Hangartner et al., 2011; Miaud & Merilä, 2001, Muir et al., 2014b) show that
67 amphibian life history traits including growth and development rates exhibit high
68 adaptation to local environmental temperature where they live, they are typically
69 maximized at the temperatures most commonly experienced in the native
70 environments (cf. temperature local adaption hypothesis, Yamahira & Conover, 2002).
71 However, there is some evidence that many amphibian species show counter-gradient
72 variation in growth and development rates. Namely, although developing more slowly
73 in the wild, individuals from cold climate localities grow and develop faster when
74 reared in similar temperature with the conspecifics from warmer localities (Licht,
75 1975; Berven et al., 1979; Gollman & Gollman 1996; Laugen et al., 2003; Palo et al.,
76 2003; Morey & Reznick, 2004; Richter-Boix et al., 2006; Lind et al., 2008). These
77 findings have been explained in terms of selection favoring faster growth and
78 development rates of tadpoles from colder regions to compensate for negative effects
79 of temperatures and growth season lengths to growth and development in low
80 temperature populations (Conover & Present, 1990; Conover & Schultz, 1995;
81 Yamahira & Conover, 2002; Laugen et al., 2003).

82 High-altitude environments are characterized by low temperatures, low oxygen
83 levels, and strong UV-B radiation (Blumthaler et al., 1997; Bickler & Buck, 2007;

84 Scheinfeldt & Tishkoff, 2010). This is true also in the case of the Tibetan plateau,
85 which has an average elevation exceeding 4,500 m (Han et al., 2018). However, the
86 varying geomorphological and climatic conditions of the Tibetan plateau may impact
87 upon amphibian life history traits by selecting for divergence in growth and
88 development rates among localities. Yet, the influence of temperature on the life
89 history traits of amphibians' larvae are poorly understood (Morrison & Hero, 2003b;
90 Ma et al., 2009), especially when it comes to the Tibetan plateau.

91 *Rana kukunoris* is endemic to the eastern Tibetan plateau (Xie et al., 2000), and
92 the species is an explosive breeder with a pure capital breeding strategy (Lu et al.,
93 2008; Chen & Lu, 2011; Chen et al., 2011; Chen et al., 2013a; Chen et al., 2018). The
94 species is sexually size dimorphic with females being the larger sex (Chen et al., 2011;
95 Feng et al., 2015). Female fecundity is positively related to body size (Lu et al., 2008)
96 and males prefer to mate with larger females (Chen & Lu, 2011). The next spring after
97 winter the species will begin breeding, so females invest some energy into gonads
98 before winter, and their gonad weight decreases with increasing altitude due to
99 relatively lower breeding energy investment at high altitudes (Chen et al., 2013b). the
100 species has a wide altitudinal range with 2200 m-4400 m (Xie et al., 2000) and
101 temperatures experienced naturally by the species range from 4° C to 32° C (Chen's
102 personal observation). But so far, little is known about the effect of temperature on the
103 growth, development and activity of their tadpoles.

104 The aim of this study was to investigate the impacts of temperature on the
105 growth, development and activity of the tadpoles of *Rana kukunoris* to discover the

106 growth and developmental strategies of high-altitude tadpoles. Specifically, we
107 investigated how temperature influence 1) development, 2) behavior, 3) size at
108 metamorphosis, and 4) survival rate of *R. kukunoris* tadpoles from different altitudes
109 until metamorphosis using controlled laboratory experiments.

110

111 **Material and Methods**

112 *Collection and husbandry of animals*

113 We collected freshly egg clutches of *R. kukunoris* from small ponds near Galitai
114 (32°58'N; 103°20'E, 3800 m) and Hongxin (34°05'N; 102°45'E, 3200 m) in the
115 Zoige county in Sichuan province and from a pond near Gahai (34°17'N; 102°17'E,
116 3500 m) in Gansu province in March 2018. In addition, clutches were also collected
117 from a pond near Gaoqiang village (32°31'N; 96°25'E, 4200 m) in Yushu in Qinghai
118 province in May 2018. Egg clutches were transferred to laboratory of Mianyang
119 Normal University (31°27'N; 104°35'E, 467m) and maintained in white plastic
120 containers (35 ×25 × 20 cm) supplied with aerated dechlorinated water until they
121 reached Gosner stage 18 (Gosner, 1960). Room temperature was set to 24°C, a
122 common temperature the species experiences naturally during development. To
123 investigate the influences of temperature we carried out the following experiment
124 using the tadpoles from egg clutches from difference altitudes.

125

126 *Experimental treatments*

127 To examine the effect of temperature on growth and development of the tadpoles, we

128 use a factorial design with three temperature conditions (16°C, 24°C, and 32°C) ×
129 four altitude sites (3200 m, 3500 m, 3800 m and 4200 m). All experiments were
130 repeated 10 times. Temperatures were selected on the basis of the temperatures
131 recorded in the field conditions, and they were maintained using automatic air
132 conditioning. Eggs from different altitudes were housed separately, ten embryos
133 randomly selected at stage 18 (Gosner, 1960) from the same clutch were transferred to
134 1 L plastic containers filled with 750 mL dechlorinated water. After hatched (stage
135 25), tadpoles were fed with goldfish feed (Foshan T and F Pet Food Co., Ltd.) twice a
136 week water changes. All the water was changed out each time. The photoperiod was
137 set to 12 h: 12 h (L: D).

138

139 *Trials to quantify behavior*

140 Body weight of metamorphs (tail completely absorbed; Gosner stage 46) was
141 also measured to the nearest 0.001 g with an electronic balance, body length and the
142 maximum jumping performance of metamorphs were measured with a plastic ruler,
143 the nearest 0.5 mm. Frogs were placed on the plastic plate under the room temperature
144 of 24°C and allowed to rest for 5s before inducing them to jump by pinching the
145 urostyle with a pair of fine forceps (Wilson et al., 2000). This elicited escape
146 responses from the individuals causing them to jump away from the observer. Each
147 metamorph was stimulated to jump at least five times, with the jump that produced
148 the greatest distance for given individual used as a measure of maximum jumping
149 performance (Wilson et al., 2000). The test frogs were maintained at 24°C for at least

150 1 hour prior to experimental procedures.

151

152 *Statistical analyses*

153 We used Linear Mixed Models (LMM) to investigate the effect of temperatures on the
154 development and behaviour of tadpoles and metamorphs. Development rate (the time
155 (day) from stage 18 to stage 46)/ body length of metamorph frogs was used as the
156 response variable. Specifically, we considered the temperature and altitude as fixed
157 explanatory variables and container identity as a random factor. Altitude \times
158 temperature interaction was also considered to test if the effects of temperature
159 treatments were similar in populations from different altitudes. Embryonic mortality,
160 treated as a binomial response variable, was analyzed with a type III generalized
161 linear model with logit-link function and binomial error structure (Räsänen et al.,
162 2003). Post-hoc tests (using SNK test) were performed to investigate significant
163 factors in the linear mixed models.

164 Before the analyses, all the variables including body length and jumping distance
165 and development rate were log-transformed to meet the assumptions of normality and
166 homoscedasticity. Normality of residuals and homogeneity of variances were
167 validated through visual inspection of graphical model evaluation plots. To account
168 for the complex covariance structure in our mixed models, we used REML estimation
169 for random effects, and the Satterthwaite procedure to approximate degrees of
170 freedom. Analyses were conducted using R studio and the “lmerTest” package. Data
171 are presented as means \pm standard deviation (SD).

172

173 **Results**

174 *The influence of temperature treatments*

175 Temperature had positive influence on the early developmental rate of the tadpoles
176 (estimate = -35.437 ± 8.128 ; $df = 116$; $t = -4.360$; $P < 0.001$; Table 1; Fig. 1), and
177 developmental rate also differed between individuals from different altitudes (estimate
178 = -12.588 ± 3.122 ; $df = 116$; $t = -4.032$; $P < 0.001$). However, a significant interaction
179 effect between altitude and temperature treatment (estimate = 4.075 ± 0.991 ; $df = 116$;
180 $t = 4.113$; $P < 0.001$) suggested that the developmental rate of tadpoles from different
181 altitudes differed depending on the temperature.

182 The results of post-hoc tests (SNK tests) showed that high-altitude tadpoles
183 developed faster in coldest temperature treatment than low-altitude tadpoles, with an
184 increasing development rate of 32.12% -35.77% between temperatures. Generally, in
185 the higher temperatures low-altitude tadpoles developed faster compared to
186 high-altitude tadpoles with a decreasing development rate of 34.09%-54.34% (Table 1;
187 Fig. 1). The development pattern of the tadpoles was complex in higher temperatures,
188 the development rate of tadpoles from 3200-m population and 3500-m population
189 showed no significant difference, the tadpoles from 3800-m population showed faster
190 development than those from 4200-m population (Fig. 1). Nonetheless, the total trend
191 was the tadpoles from low-altitude populations (3200m and 3500m) developed faster
192 than those from high- altitude populations (3800 m and 4200m; Table 1)

193 Metamorphs' jumping ability differed among temperature treatments (estimate =

194 15.161 ± 5.117 ; $df = 107.013$; $t = 2.963$; $P = 0.004$; Fig. 2), as well as among
195 individuals from different altitudes (estimate = 5.776 ± 1.965 ; $df = 107.013$; $t = 2.939$;
196 $P = 0.004$). However, a significant temperature treatment \times altitude interaction
197 (estimate = -1.809 ± 0.624 ; $df = 107.013$; $t = -2.901$; $P = 0.005$) revealed that the
198 effect of temperature treatment depended on altitude of origin. Tadpoles from low
199 altitude population showed greater improvement of jumping ability (39.52%) with
200 increasing temperature, high altitude population showed relatively small increasing
201 jumping ability (9%). This was not a case after controlling for the effect of body
202 length (estimate = 0.628 ± 0.320 ; $df = 115$; $t = 1.960$; $P = 0.052$; temperature:
203 estimate = 12.640 ± 5.241 , $df = 115$; $t = 2.412$; $p = 0.017$; altitude: estimate = $4.671 \pm$
204 2.031 , $df = 115$; $t = 2.299$; $P = 0.023$; interaction: estimate = -1.483 ± 0.641 , $df = 115$;
205 $t = -2.314$; $P = 0.022$), suggesting body size was an important factor influencing
206 Metamorphs' jumping ability.

207 There was significant effect of temperature treatment (estimate = 4.016 ± 1.450 ;
208 $df = 107.012$; $t = 2.769$; $P = 0.007$) and altitude of origin (estimate = 1.761 ± 0.557 ; df
209 = 107.012 ; $t = 3.161$; $P = 0.002$) on body length of metamorphs. The temperature \times
210 altitude interaction was significant (estimate = -0.519 ± 0.177 , $df = 107.012$; $t =$
211 -2.937 ; $P = 0.004$, Fig. 3), showing that effect of temperature treatment differed
212 depending on the altitude of origin. In general, low temperatures resulted in larger
213 size in metamorphosis (9.4%-19.72%; Fig.3), but tadpoles at metamorphose from
214 high- and low- altitude populations showed no significant difference in body size in
215 high temperature treatment (Table1; Fig.3).

216 Mortality of tadpoles over the experimental period was independent of
217 temperature treatments (estimate = -112.81 ± 84.88 ; $z = -1.329$; $P = 0.184$), altitude
218 (estimate = -46.29 ± 33.00 ; $z = -1.403$; $P = 0.161$) and their combination (altitude \times
219 temperature interaction: estimate = -13.76 ± 10.38 ; $z = 1.326$; $P = 0.185$; Table 1).

220

221 **Discussion**

222 In this study, we found that temperature had a critical influence on the growth and
223 development rate of Tibetan brown frog tadpoles. Specifically, our results showed that
224 tadpoles from different altitudes responded differently to different temperature
225 treatments: high-altitude individuals grew faster than low-altitude individuals at low
226 temperatures, while at high temperatures the results were less clear. This suggests that
227 high-altitude populations have adapted to life in cold temperatures as their growth rate
228 was maximized at the low temperatures they experience in their native environment
229 (cf. Levinton & Monahan, 1983; Jin et al., 2016; Mai et al., 2017).

230 Low temperatures act to reduce growth and developmental rates, and
231 consequently, prolonged growth and developmental periods typically lead to
232 metamorphosis at larger body size (Berven, 1982; Howard & Wallace, 1985;
233 Morrison & Hero, 2003a, b; Kuparinen et al., 2010; Liao & Lu, 2010). Our results
234 conform to this pattern since we also found that the body length of metamorphs was
235 also influenced by temperature treatments: Tibetan brown frog tadpoles from low
236 temperature treatments showed slower development rate and larger body size at
237 metamorphosis.

238 Temperature is a critical environmental factor that has shaped most life history
239 traits of amphibians (Stearns, 1992; Morrison & Hero, 2003a; Kuparinen et al., 2010;
240 Liao et al., 2010; Liao et al., 2016). Our results showed that tadpoles from all
241 populations developed fastest in the high temperature treatment. This is typical for
242 amphibians (Abrams et al., 1996; Morrison & Hero, 2003b) and can be explained by
243 the fact that high temperature increases metabolic enzyme activity, speed up the
244 process of metabolism, increase food conversion efficiency and eventually contribute
245 to the fast growth and development of tadpoles (Marian & Pandian, 1985). Similar
246 pattern applies also to plants (Way & Oren, 2010), insects (Wu et al., 2015), fishes
247 (Jerry & Thépot, 2015), and reptiles (Ewert, 1985). In our study, we also found that in
248 cold temperature condition, tadpoles from high-altitude populations showed faster
249 developmental rate compared to these from low-altitude populations, supporting the
250 temperature adaptation hypothesis (Yamahira & Conover, 2002).

251 To sum up, we investigated the impacts of temperature on the growth,
252 development and behaviour of Tibetan brown frog tadpoles, and found that
253 temperature appears to play a critical role in their growth, development, but not
254 jumping behavior. Evidence was also found for adaptation to cold temperature in high
255 altitude populations, which exhibited faster growth and developmental rates in cold
256 temperatures than those from lower altitudes. Finally, it should be noted that our study
257 design was not optimal because only small clutches from each of the study
258 populations were used to carry out the experiments. Hence, there was no among
259 family level replication in our study, and the inferences are based on the assumption

260 that the randomly selected family from each of the populations is representative of the
261 given population.

262

263 **Acknowledgments**

264 This study was funded by the Natural Sciences Foundation for Distinguished Young
265 Scholar of Sichuan (grant number 2016JQ0038), Key Foundation of Sichuan
266 Provincial Department of Education (grant number 18ZA0255), the National Sciences
267 Foundation of China (grant number 31670392 and 31872216) and the Finnish
268 Cultural Foundation (grant to PM) and the Academy of Finland (grants 129662,
269 134728, and 218343 to JM). All field and laboratory work were conducted under
270 permission from Mianyang Normal University.

271

272 **References**

- 273 Abrams, P.A., Leimar, O., Nylin, S. & Wiklund, C. (1996). The effect of flexible
274 growth rates on optimal sizes and development times in a seasonal
275 environment. *Am. Nat.*, 147, 381–395.
- 276 Berven, K. (1982). The genetic basis of altitudinal variation in the wood frog *Rana*
277 *sylvatica*. I. An experimental analysis of life history traits. *Evolution*, 36,
278 962–983.
- 279 Berven, K.A., Gill, D.E. & Smith-Gill, S.J. (1979). Countergradient selection in the
280 green frog, *Rana clamitans*. *Evolution*, 33, 609–623.
- 281 Bickler, P.E. & Buck, L.T. (2007). Hypoxia tolerance in reptiles, amphibians, and

282 fishes: life with variable oxygen availability. *Annu. Rev. Physiol.*, 69, 145–170.

283 Blumthaler, M., Ambach, W. & Ellinger, R. (1997). Increase in solar UV radiation
284 with altitude. *J. Photochem. Photobiol. B-Biol.*, 39, 130–134.

285 Burton, T. & Metcalfe, N.B. (2014). Can environmental conditions experience in early
286 life influence future generations? *Proc. Biol. Sci.* 281, 20140311.

287 Chen, W. & Lu, X. (2011). Sex recognition and mate choice in male *Rana kukunoris*.
288 *Herpetol. J.*, 21, 141–144.

289 Chen, W., Peng, L.Q., Jiang, L.C., Pike, D.A., Friesen, C.R. & Brown, G. (2018).
290 High altitude frogs (*Rana kukunoris*) adopt a diversified bet-hedging strategy
291 in the face of environmental unpredictability. *Asian Herpetol. Res.*, 9, 43–49.

292 Chen, W., Tang, Z.H., Fan, X.G., Wang, X.Y. & Pike, D.A. (2013b). Maternal
293 investment increases with altitude in a frog on the Tibetan Plateau. *J. Evol.*
294 *Biol.*, 26, 2710–2715.

295 Chen, W., Wang, X.Y. & Fan, X.G. (2013a). Do anurans living in higher altitudes have
296 higher prehibernation energy storage? Investigations from a high-altitude frog.
297 *Herpetol. J.*, 23, 45–49.

298 Chen, W., Yu, T.L. & Lu, X. (2011). Age and body size of *Rana kukunoris*, a
299 high-elevation frog native to the Tibetan plateau. *Herpetol. J.*, 21, 149–151.

300 Conover, D.O. & Present, T.M.C. (1990). Countergradient variation in growth rate:
301 compensation for length of the growing season among Atlantic silversides
302 from different latitudes. *Oecologia*, 83, 316–324.

303 Conover, D.O. & Schultz, E.F. (1995). Phenotypic similarity and the evolutionary

304 significance of countergradient variation. *Trends Ecol. Evol.*, 10, 248–252.

305 Ewert, M.A. (1985). Embryology of turtles. Pp. 75–267 in C. Gans, F. Billett, and
306 P.F.A. Madereson, eds. *Biology of the Reptilia*. Wiley, New York.

307 Feng, X.Y., Chen, W., Hu, J.H. & Jiang, J.P. (2015). Variation and sexual dimorphism
308 of body size in the plateau brown frog along an altitudinal gradient. *Asian*
309 *Herpetol. Res.*, 6, 291–297.

310 Garcia, T.S., Urbina, J.C., Bredeweg, E.M., Ferrari, M.C.O. (2017). Embryonic
311 learning and developmental carry-over effects in an invasive anuran.
312 *Oecologia*, 184, 623–631.

313 Gollman, B. & Gollman, G. (1996). Geographic variation of larval traits in the
314 Australian frog *Geocrinia victoriana*. *Herpetologica*, 52, 181–187.

315 Gosner, K.L. (1960). A simplified table for staging anuran embryos and larvae with
316 notes on identification. *Herpetologica*, 16, 183–190.

317 Han, X.Z., Zhang, Q., Fan, L.Q., Yang, L. & Liu, Z.S. (2018). The Influence of
318 oxygen on the development of *Nanoranaparkeri* tadpoles. *Asian Herpetol.*
319 *Res.*, 9, 50–55.

320 Hangartner, S.B., Laurila, A. & Räsänen, K. (2011). Adaptive divergence of the moor
321 frog (*Rana arvalis*) along an acidification gradient. *BMC Evol. Biol.*, 11,
322 366–378.

323 Howard, J.H. & Wallace, R.L. (1985). Life history characteristics of populations of
324 the long-toed salamander (*Ambystoma macrodactylum*) from different
325 altitudes. *Am. Midl. Nat.*, 113, 361–372.

326 Jerry, D.R. & Thépot, V. (2015). The effect of temperature on the embryonic
327 development of barramundi, the Australian strain of *Lates calcarifer* (Bloch)
328 using current hatchery practices. *Aquaculture Rep.*, 12, 132–138.

329 Jin, L., Yang, S.N., Liao, W.B. & Lüpold, S. (2016). Altitude underlies variation in the
330 mating system, somatic condition and investment in reproductive traits in male
331 Asian grass frogs (*Fejervarya limnocharis*). *Behav. Ecol. Sociobiol.*, 70:
332 1197-1208.

333 Kuparinen, A., Laugen, A.T., Laurila, A. & Merilä, J. (2010). Developmental
334 threshold model challenged by temperature. *Evol. Ecol. Res.*, 12, 821–829.

335 Laugen, A.T., Laurila, A., Räsänen, K. & Merilä, J. (2003). Latitudinal countergradient
336 variation in the common frog (*Rana temporaria*) development rates – evidence
337 for local adaptation. *J. Evol. Biol.*, 16, 996–1005

338 Levinton, J.S. & Monahan, R. K. (1983). The latitudinal compensation hypothesis:
339 Growth data and a model of latitudinal growth differentiation based upon
340 energy budgets. II. Intraspecific comparisons between subspecies of
341 *Ophryotrocha puerilis* (Polychaeta: Dorvilleidae). *Biol. Bull.*, 165, 699–707.

342 Liao, W.B. & Lu, X. (2010). Age structure and body size of the Chuanxi Tree Frog
343 *Hyla annectans chuanxiensis* from two different elevations in Sichuan (China).
344 *Zool. Anz.*, 248, 255-263.

345 Liao, W.B. & Lu, X. (2012). Adult body size = f (initial size + growth rate × age):
346 explaining the proximate cause of Bergman’s cline in a toad along altitudinal
347 gradients. *Evol. Ecol.*, 26, 579-590.

- 348 Liao, W.B., Liu, W.C. & Merilä, J. (2015). Andrew meets Rensch: Sexual size
349 dimorphism and the inverse of Rensch's rule in Andrew's toad (*Bufo*
350 *andrewsi*). *Oecologia*, 177, 389-399.
- 351 Liao, W.B., Lu, X. & Jehle R. (2014). Altitudinal variation in reproductive investment
352 and trade-off between egg size and clutch size in the Andrew's Toad (*Bufo*
353 *andrewsi*). *J. Zool.*, 293, 84-91.
- 354 Liao, W.B., Luo, Y., Lou, S.L., Lu, D. & Jehle, R. (2016). Geographic variation in
355 life-history traits: growth season affects age structure, egg size and clutch size
356 in Andrew's toad (*Bufoandrewsi*). *Front. Zool.*, 13, 6.
- 357 Liao, W.B., Zhou, C.Q., Yang, Z.S., Hu, J.C. & Lu, X. (2010). Age, size and growth in
358 two populations of the dark-spotted frog *Rana nigromaculata* at different
359 altitudes in southwestern China. *Herpetol. J.*, 20, 77–82.
- 360 Licht, L.E. (1975). Comparative life history features of the western spotted frog, *Rana*
361 *pretiosa*, from low- and high- elevation populations. *Can. J. Zool.*, 53,
362 1254–1258.
- 363 Lind, M.I., Persbo, F. & Johansson, F. (2008). Pool desiccation and developmental
364 thresholds in the common frog, *Rana temporaria*. *P. Roy. Soc. Lond. B. Bio.*,
365 275, 1073–1080.
- 366 Liu, Q.,Feng, H.,Jin, L.,Mi, Z.P., Zhou Z. M. & Liao W.B.(2018). Latitudinal
367 variation in body size in *Fejervarya limnocharis* supports the inverse of
368 Bergmann's rule.68, 113–128.
- 369 Lu, X., Zeng, X.H., Du, B. & Nie, C. (2008). Reproductive ecology of *Rana*

370 *kukunoris* Nikolskii, 1918, a high-elevation frog native to the Tibetan Plateau.
371 *Herpetozoa*, 21, 67–77.

372 Ma, X.Y., Lu, X. & Merilä, J. (2009). Altitudinal decline of body size in a Tibetan
373 frog. *J. Zool.*, 279, 364–371.

374 Mai, C.L., Liu, Y.H., Jin, L., Mi, Z.P. & Liao, W.B. 2017. Altitudinal variation in
375 somatic condition and investment in reproductive traits in male Yunnan pond
376 frog (*Pelophylax pleuraden*). *Zool. Anz.*, 266, 189-195.

377 Marian, M.P. & Pandian, T.J. (1985). Effect of temperature on development, growth
378 and bioenergetics of the bullfrog tadpole *Rana tigrine*. *J. Therm. Biol.*, 10,
379 157–161.

380 Miaud, C. & Merilä, J. (2001). Local adaptation or environmental induction? Causes
381 of population differentiation in alpine amphibians. *Biota*, 2, 31–50.

382 Morey, S.R. & Reznick, D.N. (2004). The relationship between habitat permanence
383 and larval development in California spadefoot toads: field and laboratory
384 comparisons of developmental plasticity. *Oikos*, 104, 172–190.

385 Morrison, C. & Hero, J.M. (2003a). Geographic variation in life-history
386 characteristics of amphibians: a review. *J. Anim. Ecol.*, 72, 270–279.

387 Morrison, C. & Hero, J.M. (2003b). Altitudinal variation in growth and development
388 rates of tadpoles of *Litoria chloris* and *Litoria pearsoniana* in Southeast
389 Queensland, Australia. *J. Herpetol.*, 37, 59–64.

390 Muir, A.P., Biek, R. & Mable, B.K. (2014a). Behavioural and physiological
391 adaptations to low-temperature environments in the common frog, *Rana*

392 *temporaria*. *BMC Evol. Biol.*, 14, 110. doi:10.1186/1471-2148-14-110

393 Muir, A.P., Biek, R., Thomas, R. & Mable, B.K. (2014b). Local adaptation with high
394 gene flow: temperature parameters drive adaptation to altitude in the common
395 frog (*Rana temporaria*). *Mol. Ecol.*, 23, 561–574.

396 Palo, J.U., O’Hara, R.B., Laugen, A.T., Laurila, A., Primmer, C.R. & Merilä, J. (2003).
397 Latitudinal divergence of common frog (*Rana temporaria*) life history traits
398 by natural selection: evidence from a comparison of molecular and
399 quantitative genetic data. *Mol. Ecol.*, 12, 1963–1978.

400 Räsänen, K., Pahkala, M., Laurila, A. & Merilä, J. (2003). Does jelly envelope protect
401 the common frog *Rana temporaria* embryos from UV-B radiation?
402 *Herpetologica*, 59, 293–300.

403 Richter-Boix, A., Llorente, G.A. & Montori, A. (2006). A comparative analysis of the
404 adaptive developmental plasticity hypothesis in six Mediterranean anuran
405 species along a pond permanency gradient. *Evol. Ecol. Res.*, 8, 1139–1154.

406 Scheinfeldt, L.B. & Tishkoff, S.A. (2010). Living the high life: high-altitude
407 adaptation. *Genome Biol.*, 11, 133.

408 Stearns, S.C. (1992). The evolution of life histories. Oxford: Oxford University Press.

409 Voituron, Y., Joly, P., Eugène, M. & Barré, H. (2005). Freezing tolerance of the
410 European water frogs: the good, the bad, and the ugly. *Am. J. Physiol. Regul.*
411 *Integr. Comp. Physiol.*, 288, 1563–1570.

412 Wang, X., Huang, Y., Zhong, M., Yang, S., Yang, X., Jiang, J. & Hu, J. (2019).
413 Environmental stress shapes life-history variation in the swelled-vented frog

414 (*Feirana quadranus*). *Evol. Ecol.*, 33, 435–448.

415 Way, D.A. & Oren, R. (2010). Differential responses to changes in growth
416 temperature between trees from different functional groups and biomes: a
417 review and synthesis of data. *Tree Physiol.*, 30, 669–688.

418 Wells, K.D. (1977). The social behaviour of anuran amphibians. *Anim. Behav.*, 25,
419 666–693.

420 Wu, T.H., Shiao, S.F. & Okuyama, T. (2015). Development of insects under
421 fluctuating temperature: a review and case study. *J. Appl. Entomol.*, 139,
422 592–599.

423 Xie, F., Fei, L. & Ye, C.Y. (2000). On taxonomic status and relationships of *Rana*
424 *japonica* group, in China (Amphibia: Anura: Radidae). *Cultum. Herpetol.*
425 *Sinica.*, 8, 74–80.

426 Yamahira, K. & Conover, D.O. (2002). Intra- vs. interspecific latitudinal variation in
427 growth: adaptation to temperature or seasonality? *Ecology*, 83, 1252–1262.

428 Yu, T.L., Wang, D.L., Busam, M. & Deng, Y.H. (2019). Altitudinal variation in body
429 size in *Bufo minshanicus* supports Bergmann's rule. *Evol. Ecol.*, 33, 449–460.

430 Zhong, M.J. & Liao, W.B. (2018). A review for life-history traits variation in frogs
431 especially for anurans in China. *Asian Herpetol. Res.*, 9, 165–174.

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433

434 **Tables**

435 **Table 1.** Mean (Standard deviation SD) developmental time (in days) of Tibetan
436 brown frog tadpoles from four different populations in three temperature treatments.
437 The developmental time refers to days in between Gosner (1960) stages 18 and 45.
438 Means with different letters (a, b, c, d) are significantly different at $P < 0.05$ (SNK
439 tests).
440

441

442

443 **Figure legends**

444

445 **Figure 1** Effects of temperature on early development of Tibetan brown frog tadpoles
446 from four different populations. Tadpoles were exposed to different temperature
447 treatments from Gosner stage 18 onwards, and the developmental time is expressed as
448 the days from stage 18 to stage 45. Standard deviation is presented by the bars. The
449 legend indicates different altitudes.

450

451 **Figure 2** Effects of temperature on jumping ability (distance) of Tibetan brown frog
452 metamorphs from four different populations. Standard deviation is presented by the
453 bars. The legend indicates different altitudes.

454

455 **Figure 3** Effects of temperature on body length of Tibetan brown frog metamorphs
456 from four different populations. Standard deviation is presented by the bars. The
457 legend indicates different altitudes.

458

459

460

Table 1

		<i>N</i>	3200 m	3500 m	3800 m	4200 m
Parameter	Treatments					
Development time	16°C	10	137.70 ± 0.67 ^a	111.00 ± 0.47 ^b	97.60 ± 0.70 ^c	91.60 ± 1.17 ^d
	24°C	10	27.60 ± 0.97 ^a	23.60 ± 0.70 ^b	46.70 ± 0.67 ^c	29.10 ± 0.74 ^d
	32°C	10	23.80 ± 1.23 ^a	21.90 ± 1.29 ^a	45.00 ± 0.47 ^b	27.90 ± 0.74 ^c
Body length	16°C	10	1.28 ± 0.07 ^a	1.38 ± 0.06 ^b	1.36 ± 0.03 ^b	1.43 ± 0.08 ^b
	24°C	10	1.20 ± 0.03	1.18 ± 0.05	1.18 ± 0.04	1.22 ± 0.12
	32°C	10	1.16 ± 0.06	1.13 ± 0.05	1.18 ± 0.06	1.15 ± 0.10
Jumping distance	16°C	10	4.58 ± 0.62 ^a	3.71 ± 0.53 ^b	5.07 ± 0.64 ^c	5.43 ± 0.42 ^c
	24°C	10	6.36 ± 1.17 ^a	4.98 ± 0.56 ^b	4.86 ± 0.92 ^b	5.77 ± 1.41 ^c
	32°C	10	6.39 ± 1.10	5.43 ± 0.97	5.68 ± 0.98	5.92 ± 1.50
Survival rate	16°C	10	95.00 ± 10.80	95.00 ± 10.80	98.00 ± 6.32	100.00 ± 0.00
	24°C	10	97.00 ± 6.75	97.00 ± 6.75	99.00 ± 3.16	99.00 ± 3.16
	32°C	10	99.00 ± 3.16	98.00 ± 6.32	100.00 ± 0.00	97.00 ± 6.75

Figure 1

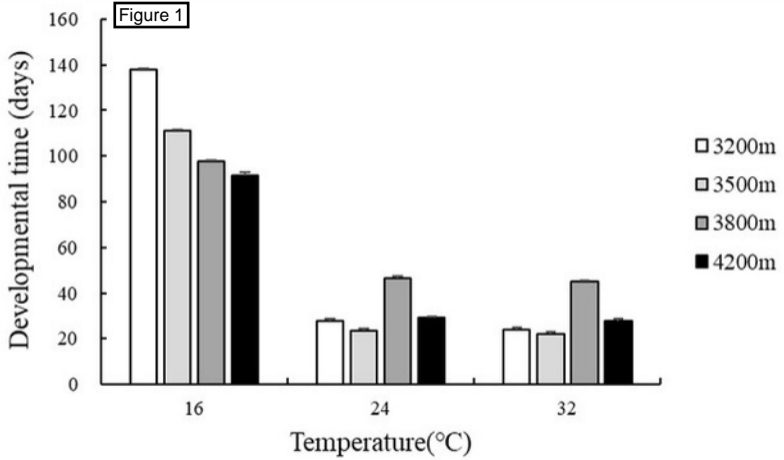


Figure 3

