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Review

Exosomes in hepatocellular carcinoma microenvironment and their potential clinical application value

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ABSTRACT

Hepatocellular carcinoma (HCC) has become a challenging disease in the world today. Due to the limitations on the current diagnosis and treatment as well as its high metastatic ability and high recurrence rate, HCC gradually becomes the second deadliest tumor. Exosomes are one of the types of cell-derived vesicles and can carry intracellular materials such as genetic materials, lipids, and proteins. In recent years, it has been verified that exosomes are linked to numerous physiological and pathological processes, including HCC. However, how exosomes affect HCC progression remains largely unknown. In this review, the exosome-mediated cellular material transfer between cells of different types in the HCC microenvironment and their effects on the behaviors and functions of recipient cells are studied. Furthermore, we also addressed the underlying molecular mechanisms. We believe that new light on the diagnosis of this cancer as well as its treatment strategies will be shed after a collation of literature in this area.

1. Introduction

Primary liver cancer (PLC) includes many types, among them, hepatocellular carcinoma (HCC) occupies about 75–85% of PLC, which makes it the most common type [1]. HCC has become very commonplace nowadays, recent research shows that about 840,000 people suffering from HCC on the original basis every year, moreover, there are 780,000 people die due to this deadly disease every year [1]. Till now, the major diagnostic methods of HCC include clinical manifestations, imaging such as ultrasonography, molecular markers such as

Abbreviations: ABC-G2, sub-family G member 2; AFP, α-fetoprotein; AKT, protein kinase B; AMPK, AMP-activated protein kinase; ANGPT2, angiopoietin-2; AUC, area under the curve; B4GALT3, beta-1,4-galactosyltransferase 3; BAK1, BCL2 antagonist/killer 1; bFGF, basic fibroblast growth factor; Breg, Regulatory B; CA, carbonic anhydrase; CAFs, cancer-associated fibroblasts; CAP1, cyclase-associated protein 1; CAV1, caveolin 1; CAV2, caveolin 2; CDK2, cyclin-dependent kinase 2; CLEC3B, C-type lectin domain family 3 member B; CTLA-4, cytotoxic T-lymphocyte associated protein 4; CXCR, C-X-C chemokine receptor; DLL4, delta-like 4 ligand; DOX, doxorubicin; DVT, deep vein thrombosis; ECM, extracellular matrix; ECs, endothelial cells; EMT, epithelial-mesenchymal transition; ENO1, alpha-enolase; eNOs, endothelial nitric oxide synthase; EPCs, endothelial progenitor cells; ER, endoplasmic reticulum; ERG, ETS-related gene; ETS, erythroblast transformationspecific; EVs, vesicles; exoDOX, exosomal DOX; FAK, focal adhesion kinase; GLUT-1, glucose transporter 1; HCC, hepatocellular carcinoma; HIF, hypoxia inducible factor; HIF-1a, hypoxia-inducible factor 1a; HK2, hexokinase II; HSCs, hepatic stellate cells; HSPGs, heparin sulfate proteoglycans; IL-6, interleukin-6; IL6R, interleukin 6 receptor; IL-8, interleukin-8; IL-10, interleukin-10; ILVs, intraluminal vesicles; IME, immune microenvironment; ING4, inhibitor of growth family member 4; LAG3, lymphocyte-activation gene 3; LHX6, LIM homeobox 6; LOXL4, lysyl oxidase-like 4; MAPK, mitogen-activated protein kinase; MMP2, matrix metallopeptidase 2; MMP9, matrix metallopeptidase 9; MVBs, multivesicular bodies; NF-κB, nuclear factor-κappa B; NK, natural killer; p120, p120-catenin; PAX2, paired box 2; PBX3, PBX homeobox 3; PDK1, pyruvate dehydrogenase kinase 1; PD-L1, programmed death-ligand 1; Pgp, P-glycoprotein; PI3K, phosphatidylinositol 3-kinase; PLC, primary liver cancer; PTEN, phosphatase and tensin homolog; ROC, receiver operating characteristic curve; ROS, reactive oxygen species; RTKs, receptor tyrosine kinases; SALL4, sal-like protein 4; SMAD3, SMAD family member 3; SMAD4, SMAD family member 4; SOX2, SRY-box 2; STAT6, signal transducer and activator of transcription 6; TAMs, tumor-associated macrophages; TGF- β , transforming growth factor- β ; TIM-1, T cell immunoglobulin and mucin domain 1; TIM-3, T cell immunoglobulin and mucin domain 3; TILs, tumor-infiltrating lymphocytes; TLR, Toll-like receptor; TME, tumor microenvironment; ucRNA, ultraconserved lncRNA; USP7, ubiquitin-specific protease 7; VASN, vasorin; VE-Cad, VE-Cadherin; VEGF, vascular endothelial growth factor; VHL, von Hippel-Lindau; ZFP36, zinc finger protein 36 homolog; ZO-1, zonula occludens 1.

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Received 17 January 2021; Received in revised form 15 March 2021; Accepted 17 March 2021 Available online 26 March 2021 0753-3322/© 2021 The Author(s). Published by Elsevier Masson SAS. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-ac-ad/4.0/). serum α -fetoprotein (AFP), and diagnostic biopsy, while the treatment options for HCC mainly include surgical resection, ultrasound-guided radiofrequency ablation, liver transplantation, local radiotherapy or chemotherapy and comprehensive treatment [2,3]. So far, although we have been working on the diagnosis and treatment of HCC, certain drawbacks still possess and we have a quite long distance to go. For example, AFP is a commonly used diagnostic marker for this disease, however, the area under the receiver operating characteristic (ROC) curve (AUC) of AFP is not very satisfactory, it only exhibits AUC values of 0.726, and has a sensitivity of 0.779 and a specificity of 0.823 [4]. With the advancement of medical research, we have discovered some potential diagnostic markers one after another, making the shortcomings of AFP more obvious [4]. Besides, surgical resection is suited for the therapy of patients with a solitary tumor, when the number of tumors is high, limitations and disadvantages have also emerged [3]. According to a recently released cancer statistics report, the survival rate of most common cancers, including HCC, have improved to a certain degree, but the prognosis of HCC is still not very satisfactory, due to its high metastatic ability as well as its high recurrence rate, its 5-year survival rate only reached 18%, which makes HCC the second most lethal tumor after pancreatic cancer [5]. Therefore, understanding the underlying molecular mechanisms of this disease is urgently needed, to shed new light on the diagnosis as well as its treatment strategies.

In the 1960s, Wolf discovered a new substance isolated from human platelets, which he called 'platelet-dust', this is the first detection of extracellular vesicles (EVs) [6]. With the progress in this area, we found that there are two types of EVs, including ectosomes and exosomes, among them, exosomes are smaller in size, they have diameters of 40-160 nm, with an average size of 100 nm [7]. The mechanism of exosome formation is gradually understood, intraluminal vesicles (ILVs) are formed in late endosomes through the endocytic pathway, they can be released outside the cell by the mechanism of multivesicular bodies (MVBs) fuse with the cell plasma membrane, these vesicles that be released outside the cell we call as exosomes [8]. Because of the way it is produced, exosomes can carry a variety of intracellular materials such as genetic materials, lipids, and proteins [7]. However, the physiological functions of exosomes are still not very well known and need further studies. Exosomes were initially thought to be artifacts or fragments of degraded or dead cells and had little functional potential [8]. In recent years, it has been verified that exosomes are linked to numerous physiological and pathological processes, including cancers, this may be related to the transport of genetic materials, proteins, and lipids by exosomes to recipient cells [7,9,10]. For example, Lugini et al. found that exosomes from colorectal cancer cells could transform normal mesenchymal stem cells from human colon into cancer-like cells [9]. In addition, Cossetti et al. found that somatic cell-derived exosomes could transfer genetic material into germ cells, which was different from Mendelian modes of inheritance [10]. Because of these functions, exosomes can be designed to serve as the carrier for multiple therapeutic substances and deliver contents to target cells, which is becoming a research hotspot [7]. In addition, over the past few decades, vesicles with exosomal characteristics were isolated from different body fluids, such as blood [11], urine [12], breast milk [13], ascites fluid [14], and bile [15], prompting the widely distributed of exosome in living organisms. What's more, the levels of exosomes, as well as their contents are substantially different in diverse disease states, and the number of plasmatic exosomes has shown great potential for the screening and follow-up of cancer patients in recent years [16-18]. Osti et al. found that the plasmatic exosome concentration was higher in glioblastoma patients than in healthy people, however, the level of plasmatic exosomes decreased following surgery, intriguingly, they noticed a significant increase in relapsed patients [19]. Additionally, Logozzi et al. discovered that melanoma patients exhibited a high level of plasmatic exosomes compared to healthy individuals, and a reduction in the level of plasmatic exosomes was observed while undergoing chemotherapy [20]. Similar phenomena have also been observed in other cancers, such

as Oral cancer and prostate cancer [21,22]. All these suggest that exosomes may be a potential non-invasive approach for disease diagnosis.

2. The role of exosomes in HCC microenvironment

Cancers develop in an environment which is known as tumor microenvironment (TME), a complicated system, except for tumor cells, it also includes extracellular matrix (ECM) and many other cell species, such as endothelial cells, fibroblasts, and immune cells [23]. Microenvironmental acidity, hypoxia and low nutrient supply are hallmarks of tumors, the abnormal tumor vasculature results in different blood perfusion in TME, hypoxia, and low nutrient supply are generalized phenomenons in tumor tissue that with perfusion deficit; hypoxia leads to anaerobic glycolysis in cells, which increases the production of lactic acid, in addition, tumor cells favor glycolysis even there exists abundant oxygen, this phenomenon is termed "Warburg Effect", then tumor cells translocate H+ into extracellular milieu via a variety of mechanisms, which finally causes a low pH in TME [24]. Among these mechanisms, carbonic anhydrase (CA) comes in the focus for its potent ability in proton transport. Two studies have found that the expression and activity of carbonic anhydrase in tumor cells were increased due to the low pH which made it possible to allow the survival of tumor cells in this harsh environment [25,26]. Recently, the interaction between TME and exosomes has become a focus of research. Exosome secretion is regulated through various mechanisms in TME, such as the levels of phosphorylated molecules and noncoding RNA-mediated passway [27-30]. Moreover, microenvironmental acidity and hypoxia have also been found to regulate the secretion of exosomes. Logozzi et al. demonstrated that microenvironmental acidity was involved in exosome secretion in almost all tumor types, in their research, they discovered that exosome secretion of tumor cells was increased in pH 6.5, while pH 7.4 showed the opposite result [31]. Interestingly, in another study, they found that the intraluminal acidity of plasmatic exosomes was increased as well while tumor cells were exposed to the acidic condition [25]. Furthermore, Wang et al. discovered that the production of exosomes was increased when tumor cells were exposed to hypoxia in advanced breast cancers [32]. The simultaneous presence of these factors in TME regulates the evolution and progression of cancers including immune escape, drug resistant, and metastases [33]. For instance, Federici et al. discovered that acidic microenvironment contributed to the resistance of melanoma cells to cisplatin, the mechanism related to decreased uptake or neutralization of weakly basic drugs by the acidic TME, furthermore, they also found another mechanism in this study, exosome-mediated drug transport from cell interior into the external environment, these two mechanisms formed a very efficient framework of tumor resistance to drugs, inasmuch as acidic microenvironment could increase the exosome secretion of tumor cells [34]. After the study of several cancers including HCC, we find that whether tumor cell-derived exosomes or stromal cell-derived exosomes, they all have an impact on TME and regulate the progress of cancer [35-37]. In our review, we observed that the substances in exosomes fall into two main categories, those targeting stromal cells (Table 1) and those that target HCC cells (Table 2). However, we need further researches to explore the role exosomes play in TME and the mechanisms of how exosomes influence TME. Due to this reason, we will emphasize the effect of exosomes on HCC microenvironment and their mechanisms(Fig. 1). In this way, we try to explore their potential clinical application value.

2.1. Exosomes regulate the progression of hepatocellular carcinoma via targeting endothelial cells

Tumor cells can regulate endothelial cells (ECs) behaviors via the release of contents in exosomes, in turn, increased vascular permeability, as well as active angiogenic process, lead to tumor metastasis and growth. Fang et al. observed that miR-103 was obviously increased in HCC cell-derived exosomes and could be delivered into ECs, which

Table 1

| Exosomal cargos | Cellular origin | Expression level | Recipient cells | Effects on cell behaviors and functions | Mechanism | Reference |
|--|----------------------------------|---------------------|--|--|---|-----------|
| miR-103 | HCC cells | High | ECs | Attenuate endothelial junction integrity and enhance vascular permeability | Down-regulate VE-Cad, p120, and ZO-1 | [38] |
| miR-210 | HCC cells | High | ECs | Enhance angiogenic capabilities | Down-regulate SMAD4 and STAT6 | [39] |
| ephrin-B2 | HCC cells | High | EPCs | Enhance angiogenic capabilities | Not mentioned | [41] |
| ANGPT2 | HCC cells | High | ECs | Enhance angiogenic capabilities | Up-regulate AKT/eNOs and AKT/ β-catenin pathways | [43] |
| circ-100338 | High- metastatic HCC cells | High | ECs | Promote proliferation, angiogenesis, and increase vascular permeability | Not mentioned | [45] |
| VASN | HCC cells | High | ECs | Promote migration | Not mentioned | [46] |
| miR-21 | HCC cells | High | EPCs | Suppress proliferative, migratory, and invasive capabilities | Down-regulate IL6R | [49] |
| CLEC3B | HCC cells | Low | ECs | Enhance the expression of VEGF | Down-regulate AMPK signaling pathway | [51] |
| miR-200b-3p | HCC cells | Low | ECs | Enhance angiogenic capabilities | Upregulate ERG | [52] |
| miR-1247–3p | High- metastatic HCC cells | High | Normal fibroblasts | Transform into CAFs | Down-regulate B4GALT3 to up-regulate β 1-integrin/NF- κ B signaling pathway | [55] |
| miR-21 | HCC cells | High | HSCs | Transform into CAFs | Down-regulate PTEN to up-regulate PDK1/AKT signaling pathway | [56] |
| TUC339 | HCC cells | High | Macrophages | Suppress phagocytic activity | Down-regulate TLR signaling pathway, FcyR-mediated phagocytosis pathway, and the actin cytoskeleton pathway | [59,60] |
| miR-150 | HCC cells | High | Macrophages | Enhance the secretion of VEGF | Down-regulate ING4 to up-regulate HIF Down-regulate OS9 | [61] |
| miR-23a-3p | HCC cells | High | Macrophages | Increase T-cell apoptosis and suppress T-cell functions | Up-regulate PTEN/AKT signaling pathway to up-regulate PD-L1 | [63] |
| miR-146a-5p | HCC cells | High | Macrophages | Promote macrophages toward M2-polarized TAMs and suppress T-cell functions | Not mentioned | [64] |
| RTKs | HCC cells | High | Monocytes | Block apoptosis | Up-regulate MAPK signaling pathway to inhibit caspase cleavage | [65] |
| circ-UHRF1 | HCC cells | High | NK cells | Suppress the anti-tumor functions | Down-regulate miR-449c-5p to up- regulate TIM-3 | [67] |
| miR-92b | HCC cells | High | NK cells | Suppress cytotoxicity | Down-regulate CD69 | [68] |
| 14–3–3ζ | HCC cells | High | TILs | Suppress the anti-tumor functions | Not mentioned | [71] |
| HMGB1 | HCC cells | High | TIM-1+Breg cells | Promote TIM-1+Breg cells expansion and suppress CD8+ T cells functions | Up-regulate TLR2/4-MAPK signaling pathway | [72] |
| MET proto- oncogene, S100A4, CAV1, and CAV2 | Metastatic HCC cell | High | Normal hepatocytes | Enhance the migratory and invasive capabilities | Up-regulate PI3K/AKT and MAPK signaling pathways to up-regulate active MMP-2 and MMP-9 | [73] |
| circ-MMP2 | High- metastatic | High | Low-metastatic HCC cells and normal | Enhance migratory and invasive capabilities | Down-regulate miR-136-5p to up- regulate MMP2 expression | [75] |
| Cdr1as | HCC cells | High | Normal hepatocytes | Enhance proliferative and | Not mentioned | [76] |
| linc-ROR | HCC cells | High | Normal hepatocytes | Endow stem cell properties | Not mentioned | [77] |

resulted in the destruction of endothelial integrity as well as an increase in vascular permeability, eventually promoting HCC metastasis, the mechanism related to the inhibition of the expression of membraneassociated adhesion molecules such as VE-Cadherin (VE-Cad), p120catenin (p120) and zonula occludens 1 (ZO-1) by miR-103 [38]. In HCC cell-derived exosomes, six substances were upregulated, including miR-210, ephrin-B2, Delta-like 4 ligand (DLL4), angiopoietin-2 (ANGPT2), circ-100338, and Vasorin (VASN), which had been verified to have links with active endothelium-dependent angiogenesis, however, the mechanisms were not identical [39,41,43,45,46]. MiR-210 could be delivered into ECs through exosomes, then miR-210 directly targeted SMAD family member 4 (SMAD4) and signal transducer and activator of transcription 6 (STAT6) and down-regulated their expressions, which promoted the tubulogenesis of ECs, this finally promoted tumor angiogenesis [39]. Endothelial progenitor cells (EPCs) had a close relationship with ECs, the latter evolved from EPCs [40]. One study observed that ephrin-B2 and DLL4 could be delivered into EPCs through exosomes and promoted tumor angiogenesis, previous studies

uncovered that high expression levels of ephrin-B2 and DLL4 played important roles in arterial differentiation of ECs, in this study they found that DLL4 functions via the upregulation of DLL4/Notch signaling pathway, regrettably, ephrin-B2 mechanism of action remained unknown [41,42]. On the surface of exosomes which were secreted by HCC cells, a high ANGPT2 level was detected and it could be delivered into ECs, which could promote angiogenesis of ECs via the upregulation of protein kinase B (AKT)/endothelial nitric oxide synthase (eNOs) as well as AKT/ β -catenin signaling pathways [43]. This was distinct from the ANGPT2/Tie2 pathway we knew before, the latter was the way free ANGPT2 works [44]. In addition, a high circ-100338 level was detected in exosomes which were secreted by high-metastatic HCC cells and it could be delivered into ECs, unlike the abovementioned substances, circ-100338 not only promoted ECs proliferation and angiogenesis but also increased vascular permeability, however, the mechanisms need to be further understood [45]. In another study, a high VASN level was detected in exosomes which were secreted by HCC cells and it could be delivered into ECs through heparin sulfate proteoglycans (HSPGs)

Table 2

The effect of exosomes that target HCC cells and their mechanisms.

| miR-92a-3p | High-metastatic HCC cells | High | Low-metastatic HCC cells | Promote EMT progression | Down-regulate PTEN and up-regulate Akt/Snail signaling pathway | [78] |
|--------------------|--------------------------------------|------|--|--|---|-------|
| LOXL4 | High-metastatic HCC cells | High | Low-metastatic HCC cells | Enhance the migratory and invasive capabilities | Up-regulate FAK/Src signaling pathway | [79] |
| CAP1 | High-metastatic HCC cells | High | Low-metastatic HCC cells | Enhance the migratory and invasive capabilities | Not mentioned | [80] |
| circ-PTGR1 | High-metastatic HCC cells | High | Low-metastatic HCC cells | Enhance the migratory and invasive capabilities | Down-regulate miR-449a to up-regulate MET | [81] |
| CXCR4 | High-metastatic HCC cells | High | Low-metastatic HCC cells | Enhance the migratory and invasive capabilities | Up-regulate MMP-9, MMP-2, and VEGF-C | [82] |
| ENO1 | High-metastatic HCC cells | High | Low-metastatic HCC cells | Promote proliferation, migration, and invasion | up-regulate integrin α6β4 to up-regulate FAK/Src-p38MAPK signaling pathway | [86] |
| linc-VLDLR | Resistant HCC cells | High | Non-resistant HCC cells | Enhance chemoresistance | Upregulate ABCG2 | [89] |
| linc-ROR | Resistant HCC cells | High | Non-resistant HCC cells | Enhance chemoresistance | Not mentioned | [91] |
| miR-32–5p | Resistant HCC cells | High | Non-resistant HCC cells | Enhance multidrug resistance | Down-regulate PTEN to up-regulate the PI3K/Akt signaling pathway | [92] |
| miR-744 | Resistant HCC cells | Low | Non-resistant HCC cells | Promote proliferation and suppress the chemosensitivity | Up-regulate PAX2 | [93] |
| miR-21/ miR-10b | Acidic microenvironment HCC cells | High | Non-acidic microenvironment HCC cells | Promote proliferation, migration, and invasion | Up-regulate Vimentin and Snail while down-regulate PTEN and E-cadherin | [95] |
| miR-1273f | Hypoxic HCC cells | High | Normoxic HCC cells | Induce proliferation and malignant transformation | Down-regulate LHX6 to up-regulating Wnt/β-catenin signaling pathway | [96] |
| SMAD3 | Attached HCC cells | High | Detached HCC cells | Enhance cell adhesion | Up-regulate SMAD3 signaling pathway to up-regulate ROS | [97] |
| circ-CCT3 | CAFs | High | HCC cells | Enhance glucose metabolism | Upregulate HK2 | [102] |
| miR-320a | CAFs | Low | HCC cells | promote proliferation and migration | up-regulate MAPK signaling pathway | [103] |
| miR-150-3p | CAFs | Low | HCC cells | Promote migration and invasion | Not mentioned | [104] |
| miR-335-5p | HSCs | High | HCC cells | Suppress proliferation and invasion | Not mentioned | [105] |
| CD11b/ CD18 | TAMs | High | HCC cells | Boost migratory | Up-regulate MMP-9 signaling pathway | [106] |
| miR-125a/b | TAMs | Low | HCC cells | Promote proliferative capabilities and stem cell properties | Up-regulate CD90 | [107] |
| circ- 0051443 | Normal hepatocytes | High | HCC cells | Promote cell apoptosis and suppress the cell cycle | Down-regulate miR-331-3p to up-regulate BAK1 | [108] |
| SENP3- EIF4A1 | Normal hepatocytes | High | HCC cells | Suppress proliferation and migration | Down-regulate miR-9-5p to up-regulate ZFP36 | [109] |
| circ-DB | Adipocytes | High | HCC cells | Promote proliferation and decrease DNA damage | Down-regulate miR-34a to up-regulate the USP7/Cyclin A2 signaling pathway | [110] |
| miR-23a/b | Adipocytes | High | HCC cells | Promote chemoresistance, growth, | Down-regulate VHL to up-regulate HIF- | [111] |

mediated endocytosis, which promoted migration of ECs, and finally promotes angiogenesis, however, the mechanisms need to be further understood [46].

Deep vein thrombosis (DVT) was very common in cancer patients, those who suffered from DVT were associated with a higher death risk [47,48]. The high miR-21 level was detected in exosomes which were secreted by HCC cells and it could be transported into EPCs, which suppress proliferative, migratory, and invasive capabilities of EPCs via down-regulating interleukin 6 receptor (IL6R), previous study verified that EPCs were closely related to human thrombus resolution, as a result, high-level miR-21 expression increased the mortality of HCC patients [49,50]. Contrary to these up-regulated substances, there are also some substances that are downregulated in exosomes secreted by HCC cells. Dai et al. discovered that C-type lectin domain family 3 member B (CLEC3B) was detected to have a low level in HCC-derived exosomes and could be delivered into ECs, which might result in the down-regulation of AMP-activated protein kinase (AMPK) signaling pathway, this related to an increase of vascular endothelial growth factor (VEGF) level in ECs, high level of VEGF enhanced angiogenesis and this finally promoted the tumor progression [51]. Another study found that there was a low expression of miR-200b-3p in exosomes which were secreted by HCC cells and it could be delivered into ECs, low level of miR-200b-3p enhanced angiogenic capabilities of ECs via upregulating erythroblast transformation-specific (ETS)-related gene (ERG) [52].

2.2. Exosomes regulate the progression of hepatocellular carcinoma via targeting fibroblasts and hepatic stellate cells

Cancer-associated fibroblasts (CAFs) occupy a key position in HCC stroma, which are transformed from activated fibroblasts and are closely related to tumor progression and metastasis [53,54]. A study showed that high miR-1247-3p level was detected in exosomes which were secreted by high-metastatic HCC cells and it possessed the ability to transform normal fibroblasts into CAFs via acting directly on beta-1, 4-galactosyltransferase 3 (B4GALT3) and up-regulating β1-integrin/NF-κB signaling pathway, this ultimately promoted lung metastasis of tumor cells, further study showed that CAFs could secrete pro-inflammatory cytokines, including interleukin-6 (IL-6) and interleukin-8 (IL-8) and this resulted in stemness. epithelial-mesenchymal transition (EMT), chemoresistance as well as tumorigenicity of HCC cells [55]. Moreover, miR-21 was aberrantly high expressed in HCC-derived exosomes and could be imported into hepatic stellate cells (HSCs), which would transform HSCs into CAFs, further study showed that this might be related to the down-regulation of phosphatase and tensin homolog (PTEN) and the upregulation of pyruvate dehydrogenase kinase 1 (PDK1)/AKT signaling pathway, in the past, PTEN was considered a tumor-suppressor gene, further studies showed that CAFs could secrete angiogenic cytokines including transforming growth factor- β (TGF- β), matrix metallopeptidase 2 (MMP2), matrix metallopeptidase 9 (MMP9), basic fibroblast growth factor (bFGF) and VEGF, which enhanced angiogenic capabilities of ECs, this finally promoted tumor progression [56,57].



Fig. 1. The effect of exosomes on HCC microenvironment and their mechanisms.

2.3. Exosomes regulate the progression of hepatocellular carcinoma via targeting immune cells

2.3.1. Macrophages and monocytes

Tumor-associated macrophages (TAMs) occupy a prominent position in TME and they play an important role in HCC development. During HCC progression, macrophages can be converted from M1 type to M2 type, the former inhibits tumor progression, in contrast, the latter promotes tumor progression [58]. Ultraconserved lncRNA (ucRNA) TUC339 is overexpressed in the HCC-derived exosomes, which is connected with tumor cell growth and adhesion; two pieces of research discovered that TUC339 was a key mechanism regulating macrophages activation by regulating the M1/M2 polarization, they found the mechanism may be TUC339 down-regulate FcyR-mediated phagocytosis pathway, Toll-like receptor (TLR) signaling pathway and the actin cytoskeleton pathway to decrease the ability of macrophages to phagocytize tumor cells; they also discovered that TUC339 was associated with another two signaling pathways, C-X-C chemokine receptor (CXCR) chemokine receptor binding pathways and cytokine receptor signaling pathways, however, the research on mechanisms remained only at an early stage [59,60]. Liu et al. demonstrated TAMs secreted VEGF to pro-angiogenesis in HCC progression, in this research high miR-150 level was detected in both human HCC plasma and exosomes which were secreted by HCC cells, high miR-150 level were related to high VEGF level and TAMs were the target cells of miR-150, later

intensive studies showed that miR-150 could be targeted to inhibitor of growth family member 4 (ING4), the latter was previously considered a tumor suppressor and could regulate hypoxia inducible factor (HIF) activity, ultimately increasing the level of VEGF; they also predicted another target, OS9, which could be down-regulated by miR-150, the knockdown of OS9 increase the level of VEGF [61,62]. Unlike the behavioral changes mentioned above, two studies had found another novel function of macrophages in HCC microenvironment. Among them, Liu et al. demonstrated that the release of exosomes that contained high-level miR-23a-3p could be promoted by endoplasmic reticulum (ER) stress in HCC cells, and these exosomes are then transported into macrophages, which could increase programmed death-ligand 1 (PD-L1) expression through the up-regulation of PTEN/AKT signaling pathway, this resulted in a negative impact on CD8+ T cells, including a suppression on functions as well as increased apoptosis [63]. In addition, Yin et al. found that the transcription factor Sal-like protein 4 (SALL4) could up-regulate the expression of miR-146a-5p in HCC cells through binding to the promoter, this enhanced miR-146a-5p level in exosomes and could be transferred into macrophages via endocytosis in HCC microenvironment, leading to macrophages toward M2-polarized TAMs and suppression of T-cell functions [64]. The precursors of TAMs are monocytes that undergo apoptosis in about two days, however, the inflammatory tumor microenvironment allows monocytes to survive longer in order to produce more TAMs, regrettably, the mechanisms remain obscure [65]. A recent study identified HepG2 cells-derived

exosomes participates in this process, which transmitted phosphorylated receptor tyrosine kinases (RTKs) to monocytes, this process could up-regulated mitogen-activated protein kinase (MAPK) signaling pathway, Ras-Raf-MEK-ERK, finally led to an inhibition of caspase cleavage as well as a cell cycle block [65].

2.3.2. Natural killer cells

Natural killer (NK) cells belong to a special class of cytotoxic lymphocytes and can mediate anti-tumor response through lysing cells charged with 'non-self' antigens thus it can be seen, they are strongly related to tumor immune microenvironment (IME), their activities are largely related to activating and inhibitory receptor signaling and the reduction of their activities results in increased susceptibility to cancers [66]. The dysfunction of NK cells had been verified in HCC, in one study, a high level of circ-UHRF1 was detected in exosomes which were secreted by HCC cells, it could be transferred into NK cells via endocytosis, which inhibited the anti-tumor functions of receptor cells via sponging miR-331-3p and up-regulating T cell immunoglobulin and mucin domain 3 (TIM-3), it previously showed that TIM-3 could inhibit antitumoral immunity in HCC, they also observed this might be associated with anti-PD1 therapy [67]. Another research discovered that there existed a high miR-92b level in HCC-derived exosomes and could be transferred into NK cells, which down-regulated cytotoxicity of NK cells by suppressing its target mRNAs, in this study, they mainly focused on CD69 which was a key activation marker on NK cells [68].

2.3.3. T cells and B cells

Tumor-infiltrating lymphocytes were another main component of tumor IME, the anti-tumor effects of T cells were largely suppressed in HCC [69,70]. In a study by Wang et al. high 14-3-3 clevel was detected in HCC cells, further study showed that 14-3-35 could be transferred into tumor-infiltrating T lymphocytes (TILs) through exosomes, in this study TILs with high 14-3-3 ζ level presented a decrease in inflammatory cytokines levels but an increase in anti-inflammatory cytokines levels and there was a high percentage of CD8+ T cells which carried exhaustion markers such as TIM-3, lymphocyte-activation gene 3 (LAG3) and cytotoxic T-lymphocyte associated protein 4 (CTLA-4), they found that the differentiation of naive T cells deviated at the same time, additionally, they also discovered that the activity of CD3+ T cells was inhibited in the high-level 14-3-3 ζ group, all these alterations mentioned above finally inhibited the anti-tumor functions of TILs, however, the mechanisms remained to be investigated [71]. Regulatory B (Breg) cell was a subtype of B cell, it was closely related to tumor progression as a component of humoral immunity, Ye et al. discovered a new type of Breg cell, TIM-1⁺Breg cell, different from traditional peripheral Breg cells, it exhibited a CD5^{high}CD24⁻CD27^{-/+}CD38^{+/high} phenotype, the further study discovered that T cell immunoglobulin and mucin domain 1 (TIM-1)⁺Breg cells were transformed from B cells, the mechanism might relate to the activation TLR2/4-MAPK pathway by high mobility group box 1 (HMGB1) which presented a high level in HCC-derived exosomes and could be transferred into B cells via endocytosis, this mechanism also promoted TIM-1⁺Breg cells expansion, then the receptor cells promoted the progress of HCC by secreting interleukin-10 (IL-10) and suppressing CD8⁺ T cells functions [72].

2.4. Exosomes regulate the progression of hepatocellular carcinoma via targeting normal hepatocytes

As is well known, HCC cells develop from normal hepatocytes and their interactions affect HCC progression, in recent years we found a novel mode of action of HCC cells on normal hepatocytes. He et al. found that metastatic HCC cell could affect the biological behaviors of normal hepatocytes through exosome-based intracellular materials delivery, in this research, they observed two HCC cell lines MHCC97L and HKCI-8 could deliver MET proto-oncogene, S100A4, caveolin 1 (CAV1), and caveolin 2 (CAV2) to immortalized hepatocyte MIHA through exosomes,

then up-regulated phosphatidylinositol 3-kinase (PI3K)/AKT and MAPK signaling pathways to increase secretion of active MMP-2 and MMP-9, previous studies demonstrated that these two proteins were related to metastasis, this finally enhanced the migratory capability of MIHA as well as its invasive ability [73,74]. Moreover, high circ-MMP2 level was detected in exosomes which were secreted by high-metastatic HCC cells and it could be delivered into normal hepatocytes, which promoted HCC metastasis, interestingly, they observed the same result in another type of recipient cells, low-metastatic HCC cells, the mechanism was circ-MMP2 up-regulated MMP2 expression via binding to miR-136-5p [75]. In addition, high circRNA Cdr1as level was detected in exosomes which were secreted by HCC cells and could induce cellular responses after captured by normal hepatocytes, then enhanced proliferative and migratory capabilities of recipient cells, however, the mechanisms were not mentioned [76]. In another study, a high Linc-ROR level was detected in exosomes which were secreted by HCC cells and it could be delivered into normal hepatocytes, which endowed stem cell properties to normal hepatocytes through increasing stem cells related proteins such as CD133, NANOG, SRY-box 2 (SOX2), P53 and OCT4, however, the mechanisms remained to be investigated [77].

2.5. Exosomes regulate the progression of hepatocellular carcinoma via targeting HCC cells

2.5.1. Exosome-mediated delivery of substances between different types of HCC cells

HCC cells can be divided into several types according to their biological characteristics such as migratory capability and drug resistance. The level of substances varies greatly among the exosomes secreted by different types of HCC cells. Importantly, the exosome-mediated communication between these HCC cells changes the biological characteristics of recipient cells to some extent.

$2.5.1.1. \ Between \ high-metastatic \ HCC \ cells \ and \ low-metastatic \ HCC \ cells.$

There was growing evidence that the migratory and invasive capacities of low-metastatic HCC cells could be increased by high-metastatic HCC cells via the exosome-mediated transfer of substances, studies on the underlying mechanisms contribute to a better understanding of HCC progression. The high miR-92a-3p level was detected in exosomes which were secreted by high-metastatic HCC cells and it could be delivered into low-metastatic HCC cells, which promote EMT progression of recipient cells, further studies of the mechanism found that this might be related to the down-regulation of PTEN and the up-regulation of Akt/Snail signaling pathway [78]. In addition to the EMT process, Li et al. found another important process of tumor cell invasion, cell-matrix adhesion, they discovered that lysyl oxidase-like 4 (LOXL4) could be transported from highly metastatic HCC cells into the low metastatic ones through exosomes, following this, the migratory capabilities of recipient cells, as well as their invasive abilities, were enhanced, further studies of the mechanism found that this might be related to the upregulation of focal adhesion kinase (FAK)/Src pathway, which promoted cell-matrix adhesion [79]. Moreover, a high level of adenylyl cyclase-associated protein 1 (CAP1) was detected in exosomes which were secreted by high-metastatic HCC cells, previous studies found that CAP1 had a tight relationship with HCC metastasis, in this study, they discovered that CAP1 could be delivered into low-metastatic HCC cells through exosomes, this finally enhanced the migratory capabilities of recipient cells as well as their invasive abilities, however, the mechanisms remained to be investigated [80]. Similar findings had also been identified in another two studies. Among them, Wang et al. found that circ-PTGR1 could be transported from highly metastatic HCC cells into the low metastatic ones through exosomes, and circ-PTGR1 could enhance the migratory capabilities of low-metastatic HCC cells as well as their invasive abilities by binding to miR-449a and up-regulating MET [81]. Another study obtained the same results, in this study they mainly focused on CXCR4

and the mechanism was CXCR4 up-regulated the expression of MMP-2, MMP-9, and VEGF-C [82]. With the rapid development of this emerging field, many metabolism-related enzymes have been found to regulate exosome release [83,84]. Moreover, exosomes are also found to regulate cell metabolism by delivering metabolism-related enzymes. Alpha-enolase (ENO1) is considered a multifunctional protein that mainly participates in glycolytic reactions and acts as a plasminogen receptor [85]. A recent study by Jiang et al. found that high-metastatic HCC cells could deliver ENO1 into low-metastatic HCC cells, which eventually promoted proliferative, migratory, and invasive capabilities of recipient cells, mechanistically, ENO1 could up-regulate integrin $\alpha 6\beta 4$ which was strongly related to the activation of the FAK/Src-p38MAPK signaling pathway [86].

2.5.1.2. Between resistant HCC cells and non-resistant HCC cells. Sorafenib can delay the progress of tumors as well as angiogenesis via blocking a wide variety of growth factor pathways [87]. For advanced HCC patients, sorafenib remains the only possible cure which is also the only FDA-approved treatment [88]. However, the chemoresistance to drugs contributes to high HCC-related deaths. One research discovered that while HCC cells were exposed to sorafenib, the linc-VLDLR level increased, which promoted chemoresistance via upregulating sub-family G member 2 (ABC-G2), the previous study had shown that ABC-G2 could export active drug out of the cells and this contributed to drug resistance; in addition, this research found that resistant HCC cells could transport linc-VLDLR to non-resistant HCC cells through exosomes and thus promote chemoresistance of non-resistant HCC cells [89,90]. This team had also found a similar result in another lncRNA, linc-ROR [91]. In addition, a marked rise in the expression level of miR-32–5p contributed to promoting multidrug resistance of HCC cells, further study had shown that resistant HCC cell-secreted exosomal miR-210 may be transferred into non-resistant HCC cells, leading to enhanced cancer multidrug resistance through the down-regulation of PTEN and the up-regulation of PI3K/Akt signaling pathway [92]. In contrast to the above up-regulated substances, miR-744 presented a low level in exosomes which were secreted by resistant HCC cells and these exosomes could be endocytosed by non-resistant HCC cells, the low expression of miR-744 up-regulated the expression of paired box 2 (PAX2), this finally promoted proliferation and inhibit the chemosensitivity of non-resistant HCC cells [93].

2.5.1.3. Between other types of HCC cells. It had been previously reported that the pH gradient in and around tumor cells was dysregulated, with intracellular pH (pHi) higher than 7.4 and extracellular pH (pHe) between 6.6 and 7.2, creating a good environment for metastasis [94]. Tian et al. discovered that Low pH conditions would significantly enhance the release of exosomes from HCC cells, these acidic pH-derived exosomes could transfer miR-21 and miR-10b into non-acidic microenvironment HCC cells, these two miRNAs could up-regulate Vimentin and Snail expression while down-regulate PTEN and E-cadherin levels in the recipient cells, which led to enhanced non-acidic microenvironment HCC cells proliferation, migration, and invasion [95]. In addition, Yu et al. discovered that hypoxia played a pivotal role in HCC, which could increase exosomal production in HCC microenvironment, leading to the tumor progress, in their study, they observed that miR-1273f was significantly increased in exosomes which were derived from hypoxic HCC cells and could down-regulate the expression level of tumor suppressor gene LIM homeobox 6 (LHX6) in normoxic HCC cells, which would cause the up-regulation of Wnt/ β -catenin signaling pathway, as a result, promoting malignant progression in normoxic HCC cells [96]. Moreover, Fu et al. found that attached HCC cell-secreted exosomal SMAD family member 3 (SMAD3) might be transferred into detached HCC cells and thereby promoted cell adhesion by up-regulating SMAD3 signal pathway and increased the level of reactive oxygen species (ROS) [97]. In a previous study, ROS was found to play an important role in cell

adhesion, this finally promoted tumor metastases [98].

2.5.2. Exosome-mediated delivery of substances from stromal cells to HCC cells

While stromal cells uptake HCC cell-derived exosomes, they also secrete their own exosomes and can be transferred into HCC cells via endocytosis, in this way, biological behaviors of HCC cells are modified.

2.5.2.1. CAFs and HSCs. Glucose metabolism disorder was a characteristic feature of tumor cells, despite oxygen sufficiency, tumor cells were more likely to perform glycolysis [99]. Previous studies found that hexokinase II (HK2) was the rate-limiting enzyme for glycolysis and the high HK2 level was closely related to a poorer prognosis in HCC patients, however, how HK2 was up-regulated remained largely unknown [100, 101]. One research identified circ-CCT3 as a candidate molecule for this process, they found that circ-CCT3 was highly expressed in CAF-derived exosomes and could be transferred into HCC cells, high levels of circ-CCT3 upregulated HK2 and thus promoted glucose metabolism of HCC cells [102]. In addition, two other studies discovered that the expression levels of two molecules, including miR-320a and miR-150-3p, were significantly decreased in CAF-derived exosomes and could be delivered into HCC cells [103,104]. Among them, miR-320a was an antitumor miRNA and could bind to PBX homeobox 3 (PBX3) in HCC cells, which down-regulated MAPK pathway and then suppressed MMP2 and cyclin-dependent kinase 2 (CDK2) expressions, in contrast, low levels of miR-320a induced the opposite effects and eventually promoted proliferation and migration of HCC cells [103]. Low miR-150-3p expression levels were associated with high incidences of migration and invasiveness of HCC cells and could promote the tumor progress, regrettably, the mechanism had not been mentioned in detail in this study [104]. In addition to CAFs, similar results were found in HSCs. Wang et al. discovered that high levels of miR-335-5p could be transported from HSCs to HCC cells through exosomes, which result in the inhibition of HCC cell proliferation and invasion abilities in vitro and the volume reduction of HCC tumor in vivo, however, the mechanisms remained elusive [105].

2.5.2.2. TAMs. TAM-derived exosomes are also closely related to HCC progress. Wu et al. found that integrin, $\alpha M \beta 2$ (CD11b/CD18), was highly expressed in M2 macrophage-derived exosomes and could be transferred into HCC cells, which could boost migratory of HCC cells by up-regulating MMP-9 signaling pathway [106]. Another study had reported that TAMs potentially promote HCC progress by exosome-mediated transfer of low-expressing miR-125a and miR-125b to HCC cells, the main mechanism was the low miR-125a and miR-125b levels could up-regulate CD90, which promoted cell proliferation and stem cell properties [107].

2.5.2.3. Normal hepatocytes. Two molecules including circ-0051443 and SENP3-EIF4A1 were down-regulated in HCC cells but exhibited high expressions in normal hepatocyte-derived exosomes [108,109]. Low levels of circ-0051443 in HCC cells could block apoptosis and promote cell cycle progression, however, normal hepatocytes could transfer high-level circ-0051443 into HCC cells via exosomes, then it up-regulated BCL2 antagonist/killer 1 (BAK1) by sponging miR-331-3p, this eventually suppressed the malignant biological behaviors [108]. In addition, Wang et al. found that low SENP3-EIF4A1 level in HCC cells promoted tumor progress, however, normal hepatocytes could transfer SENP3-EIF4A1 into HCC cells through exosomes, which could suppress proliferation and migration abilities of HCC cells by binding to miR-9-5p and up-regulating zinc finger protein 36 homolog (ZFP36) [109].

2.5.2.4. Adipocytes. Two recent studies reported new advancements about adipocyte-derived exosomes in HCC microenvironment. Zhang et al. found that circ-DB was highly expressed in adipocytes and could be

transported into HCC cells through exosomes, further research showed that circ-DB could function as a sponge by adsorbing miR-34a to upregulate the ubiquitin-specific protease 7 (USP7)/Cyclin A2 pathway, this eventually promoted the proliferation in parallel with a decreased DNA damage [110]. Another study found that miR-23a/b was highly expressed in adipocyte-derived exosomes and could be transferred into HCC cells, which promoted chemoresistance, growth, and migration of HCC cells, mechanistically, they found that von Hippel-Lindau (VHL) was down-regulated by miR-23a/b and thereby the expressions of hypoxia-inducible factor 1 α (HIF-1 α), glucose transporter 1 (GLUT-1) and VEGF were up-regulated [111].

3. The potential diagnostic and therapeutic values of exosomes in HCC

With the in-depth study in this emergent field, the potential clinical application value of exosomes is gradually explored. As discussed in the preceding sections, more than 50 substances have been reported to have dysregulated expression in HCC and the number is still increasing. In addition, numerous studies have discovered that the types of molecules inside exosomes, as well as their levels exist differences as the disease progresses [112–114]. For instance, Xue et al. found that miR-93 was highly expressed in HCC cell-derived exosomes and they discovered an intriguing phenomenon, the higher the miR-91 level, the later the tumor stage [112]. Lee et al. also found a similar result on another molecule, IncRNA-ATB [113]. Therefore, a deep analysis of the substances in these exosomes such as components and their levels may help achieve an accurate diagnosis of HCC [114]. Treatment of HCC remains a challenge, particularly in an advanced stage. One study discovered that doxorubicin (DOX) could be loaded into exosomes, the effectiveness of exosomal DOX (exoDOX) was almost the same as doxorubicin treatment alone, however, the cardiotoxicity produced by the former was lower [115]. Moreover, Kim et al. found that exosomes could carry drugs and transport them inside tumor cells, which could avoid the loss of active drug out of the cells via P-glycoprotein (Pgp), a drug efflux transporter [116]. These findings entail the promise of novel therapeutic approaches for HCC patients to an extent.

4. Conclusions and future perspectives

Exosomes as one of the types of cell-derived vesicles play a crucial role in HCC progression via mediating cell-cell communications. As a result, it has become a hot research topic in recent years and considerable research progress has been obtained in this area. The present review first introduced the exosomal cargos secreted by HCC cells in HCC microenvironment and their effects on recipient cells' behaviors and functions through regulating signaling pathways and downstream molecule expression after being delivered into stromal cells. Then we discussed that stromal cells could in turn influence HCC cells by a similar pathway. Moreover, this phenomenon was also present between different types of HCC cells. However, the mechanisms of some of the exosomal cargos mentioned above still remain unknown. Although exosomes have shown their potential clinical application value for HCC, this area of research is still at the preclinical stage and there still exists many challenges, for example, we still lack technologies to efficiently assay exosomes and the artificial synthetic exosomes are still difficult to obtain, additionally, the technology in the delivery of exosomes that contain drugs to the target cells is quite immature. Therefore, more research efforts should be devoted to these problems. We believe that indepth explorations of this field will shed new light on the diagnosis of this cancer as well as its treatment strategies.

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