

Paraniesslia tuberculata gen. et sp. nov., and new records or species of *Clypeosphaeria*, *Leptosphaeria* and *Astrosphaeriella* in Hong Kong freshwater habitats

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Abstract: *Paraniesslia tuberculata* gen. et sp. nov., *Clypeosphaeria uniseptata* sp. nov. and *Leptosphaeria ginimia* sp. nov. are described and illustrated from wood submerged in freshwater habitats in Hong Kong, while *Astrosphaeriella stellata* is reported as a new record from a river in Hong Kong. *Paraniesslia tuberculata* has characteristic features of the Niessliaceae, but is unique in producing greenish-brown ascospores with tuberculate ornamentation.

Key Words: Aquatic fungi, ascomycetes, lignicolous, streams, systematics

INTRODUCTION

Submerged woody substrata are important in regulating stream hydraulics and habitats for stream organisms (Triska and Cromack 1980). The lignicolous fungal communities are diverse in taxonomic groups (Shearer 1993, Goh and Hyde 1996) and important in breaking down woody substrates in freshwater ecosystems because they produce wood-decaying enzymes (Wong et al 1998). Our study of fungal biodiversity on submerged wood in streams in Hong Kong (Goh and Hyde 1999, Tsui et al 2000) yielded four interesting ascomycetes, including three new species reported in this paper.

MATERIALS AND METHODS

Samples of submerged woody substrata were collected from different streams in Hong Kong, taken to the laboratory in plastic bags, and processed following the methods described in Tsui et al (2000). Fungi were isolated using single-spore isolation techniques (Choi et al 1999) and where successful, cultures have been deposited in Hong Kong University Culture Collection (HKUCC).

TAXONOMY

Paraniesslia K. M. Tsui, K. D. Hyde et Hodgkiss, gen. nov.

Perithecia superficialia, pyriformia vel subglobosa, setosa, papillata, ostiolata, periphysata, solitaria vel gregaria, brunnea. Peridium membranaceum pluribus stratis *textura angulari* compositum, extus visum *textura epidermoidea*. Setae acutae, rectae, non ramosae, brunneae vel nigrae. Filamenta interascalica septata. Asci unitunicati, octospori, clavati, pedicellati, tenuitunicati, apice truncato, apparatu apicali praediti. Ascosporae uniseriatae imbricatae vel biseriatae, ellipsoideae, 1-septatae, viridifuscae ad brunneae, verrucosae, tunica gelatinosa praesens vel absens.

Perithecia superficial, pyriform to subglobose, papillate, with setae, ostiolate, periphysate, solitary to gregarious, brown. Peridium membranous, *textura angularis* in longitudinal section, *textura epidermoidea* in surface view. Setae acute, straight, unbranched, septate, brown or black. Interascal filaments septate. Asci unitunicate, 8-spored, clavate, pedicellate, thin-walled, apex truncate, with a nonamyloid discoid refractive apical apparatus. Ascospores overlapping uniseriate to biseriate, ellipsoidal, 1-septate, verrucose, greenish brown to brown, with or without a mucilaginous sheath.

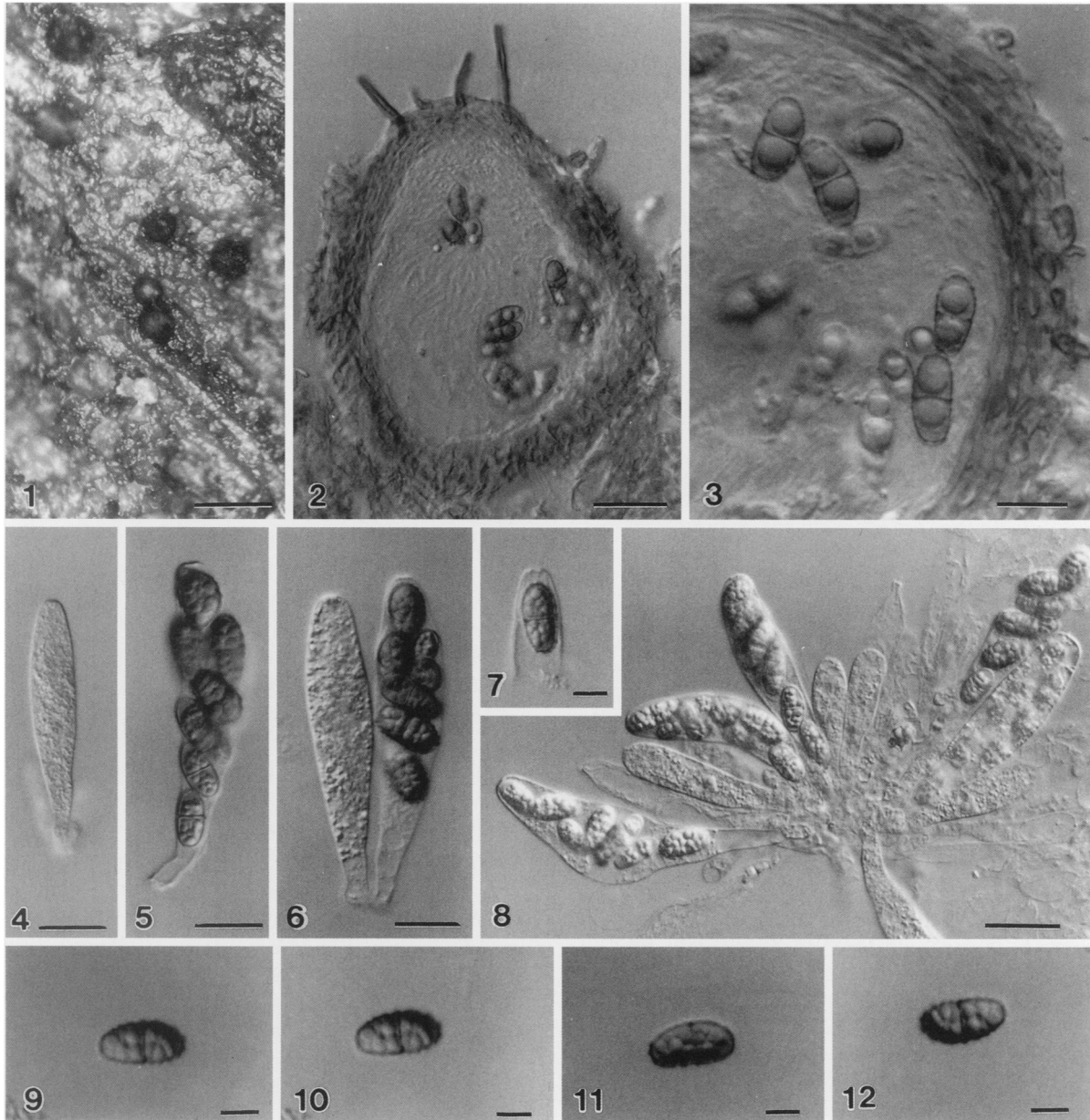
Etymology. From Latin, *Paraniesslia*, refers to the resemblance to *Niesslia*.

Species typica. *Paraniesslia tuberculata* K. M. Tsui, K. D. Hyde et Hodgkiss

Paraniesslia tuberculata K. M. Tsui, K. D. Hyde et Hodgkiss sp. nov. FIGS. 1–12

Ascomata 100–130 μm alta, 100–120 μm diam, partim superficialia vel superficialia, pyriformia vel subglobosa, setosa, ostiolata, solitaria, pallide brunnea vel brunnea. Peridium 14–20 μm crassum, pluribus stratis *textura angulari* compositum, extus visum *textura epidermoidea*, membranaceum. Setae acutae, non ramosae, brunneae. Filamenta interascalica septata, tenuitunicata, deliquescentia. Asci 50–60 \times 8–14 μm , octospori, clavati, pedicellati, tenuitunicati, apice truncato, apparatu apicali ca 1 μm alto \times 2.5 μm diam praediti. Ascosporae 11–14 \times 4–6 μm , uniseriatae imbricatae, 1-septatae, oblonge ellipsoideae, ad apices rotundatae, tuberculatae, viridifuscae, immaturae hyalinae.

Ascomata 100–130 μm high, 100–120 μm in diam, partly superficial to superficial, pyriform or globose to subglobose, short papillate with black setae, ostiolate, solitary, light brown to brown (FIGS. 1–2). Peridium 14–20 μm wide, *textura epidermoidea* in sur-



FIGS. 1–12. Light (1) and differential interference contrast (2–12) micrographs of *Paraniesslia tuberculata* (from holotype). 1. Appearance of ascomata on wood. 2. Section of the ascoma. 3. Section of the peridium. 4–7. Asci. Note the discoid apical refractive rings in 7. 8. Asci with paraphyses. 9–12. Ascospores with rough walls. Bars: 1 = 200 μm , 2 = 20 μm , 3 = 8 μm , 4–6 = 12 μm , 7 = 5 μm , 8 = 15, 9–12 = 6 μm .

face view, and *textura angularis* in longitudinal section; composed of 3–4 layers of compressed polygonal cells, membranous (FIG. 3). Setae acute, septate, brown (FIG. 2). Interascal filaments septate, thin-walled, deliquescent. Asci 50–75 \times 7–14 μm (\bar{x} = 60 \times 9 μm , n = 15), 8-spored, clavate, short pedicellate, thin-walled, apex truncate, with a discoid refractive apical apparatus, ca 1 μm high \times 2.5 μm diam (FIGS. 4–8). Ascospores 11–14 \times 4–6.5 μm (\bar{x} = 13 \times 5 μm , n = 30), overlapping uniseriate, fusoid, ellipsoidal with rounded ends, 1-septate, slightly constricted at

septum, ornamented, tuberculate, greenish-brown, hyaline when immature, without a mucilaginous sheath (FIGS. 8–12). No cultures obtained.

Etymology. From *tuberculata*, in reference to its tuberculate ascospore ornamentation.

Specimens examined. CHINA. HONG KONG: Tai Po, Lam Tsuen River, on submerged wood, Sept 1996, K. M. Tsui, KM 83 (HOLOTYPE, HKU(M) 4647), *ibid*, Lam Tsuen River, on submerged wood, May 1998, K. M. Tsui, KM 83 (HKU(M) 12152).

Notes. *Paraniesslia* is placed in the Niessliaceae,

which accommodates taxa having small, collabent, setose, perithecioid ascomata, unitunicate asci with a nonamyloid, discoid apical apparatus, and uniseptate ascospores (Samuels and Barr 1997). The genera in the Hypocreales have phialidic anamorphs and apical paraphyses, which may appear as deliquescing strands in mature ascomata when they develop from the upper meristematic tissues downwards to the base of the fruiting bodies (Rossman et al. 1999). Although the anamorphs and apical paraphyses were not clearly observed in *P. tuberculata*, the presence of deliquescing interascal filaments and the absence of true paraphyses warrant its disposition in the Hypocreales.

The identification of this taxon has been problematic. Using the key of Samuels and Barr (1997), this taxon should belong in either *Cryptoniesslia* Scheuer or *Niesslia* Auersw. *Paraniesslia tuberculata* resembles *Cryptoniesslia setulosa* Scheuer, in the Niessliaceae, in having inconspicuous, deliquescing interascal filaments found in the centrum. *Cryptoniesslia setulosa* also differs in having immersed, black ascomata and hyaline, 1-septate, long fusiform ascospores (Scheuer 1993). *Niesslia* species are characterized by having hyaline, 1-septate ascospores (Barr 1990, 1993), while the ascospores in *Paraniesslia tuberculata* are verrucose and greenish brown. *Paraniesslia* is therefore different from the described genera of Niessliaceae (sensu Samuels and Barr 1997).

Another similar genus is *Phaeotrichosphaeria* Sivan., which produces setose ascomata, unitunicate asci with refractive apical apparatus and brown, 1-septate ascospores (Sivanesan 1983). *Phaeotrichosphaeria* however is regarded as a relative of *Lasiosphaeria* Ces. & De Not. in the Lasiosphaeriaceae, and has thick-walled, noncollabent ascomata, narrow, true paraphyses, and *Endophragmiella* B. Sutton anamorphs (Sivanesan 1983, Barr 1990). The inclusion of *Paraniesslia tuberculata* in *Savoryella* R. A. Eaton & E. B. G. Jones may be considered. Their shared character is asci that are clavate and possess a discoid apical apparatus. *Paraniesslia tuberculata* is easily distinguished in having brown, setose ascomata without a neck. The ascospores are greenish brown and ellipsoidal, while *Savoryella* has black ascomata and brown, 4-septate ascospores with hyaline end cells. Also *Savoryella* is not closely related to the Niessliaceae and may belong in the Halosphaeriales (Ranghoo pers comm).

Clypeosphaeria uniseptata K. M. Tsui, K. D. Hyde et Hodgkiss, sp. nov. FIGS. 13–21

Ascomata 150–200 µm alta, 400–550 µm diam, subglobosa vel applanata, sub clypeo immersa ad partem erumpentia, ostiolata, coriacea, nigra. Papilla conica, periphysata. Peridium ca 20 µm crassum, textura angulari. Paraphy-

ses ca 5 µm crass, filamentos, septati, hyalinae. Asci 120–160 × 6–7 µm, unitunicati, cylindrici, pedicellati, apparatu subapicali iodo caerulescente (J+) praediti. Ascospores 14–20 × 5–7 µm, ellipsoideae, uniseriatae imbricatae, bicellulares, constrictae ad septum, laeves, crassitunicatae, pallide brunneae.

Ascomata 150–200 high, 400–550 diam, subglobose or applanate, erumpent to partly immersed beneath a clypeus, ostiolate, coriaceous, black (FIGS. 13, 14). Papilla conical, periphysate (FIG. 14). Peridium ca 20 µm wide, *textura angularis*, composed of several layers of brown, compressed angular cells (FIG. 15). Paraphyses ca 5 µm wide, mostly longer than asci, filamentous, septate, hardly found. Asci 120–160 × 6–8 µm (\bar{x} = 126 × 6.5 µm, n = 25), unitunicate, long cylindrical, pedicellate, with a J+, discoid, sub-apical ring (FIGS. 16, 17). Ascospores 14–19 × 5–7 µm (\bar{x} = 16 × 6 µm, n = 40), overlapping uniseriate, ellipsoidal, 1-septate, slightly constricted at septum, smooth, thick-walled, pale brown (FIGS. 18–21).

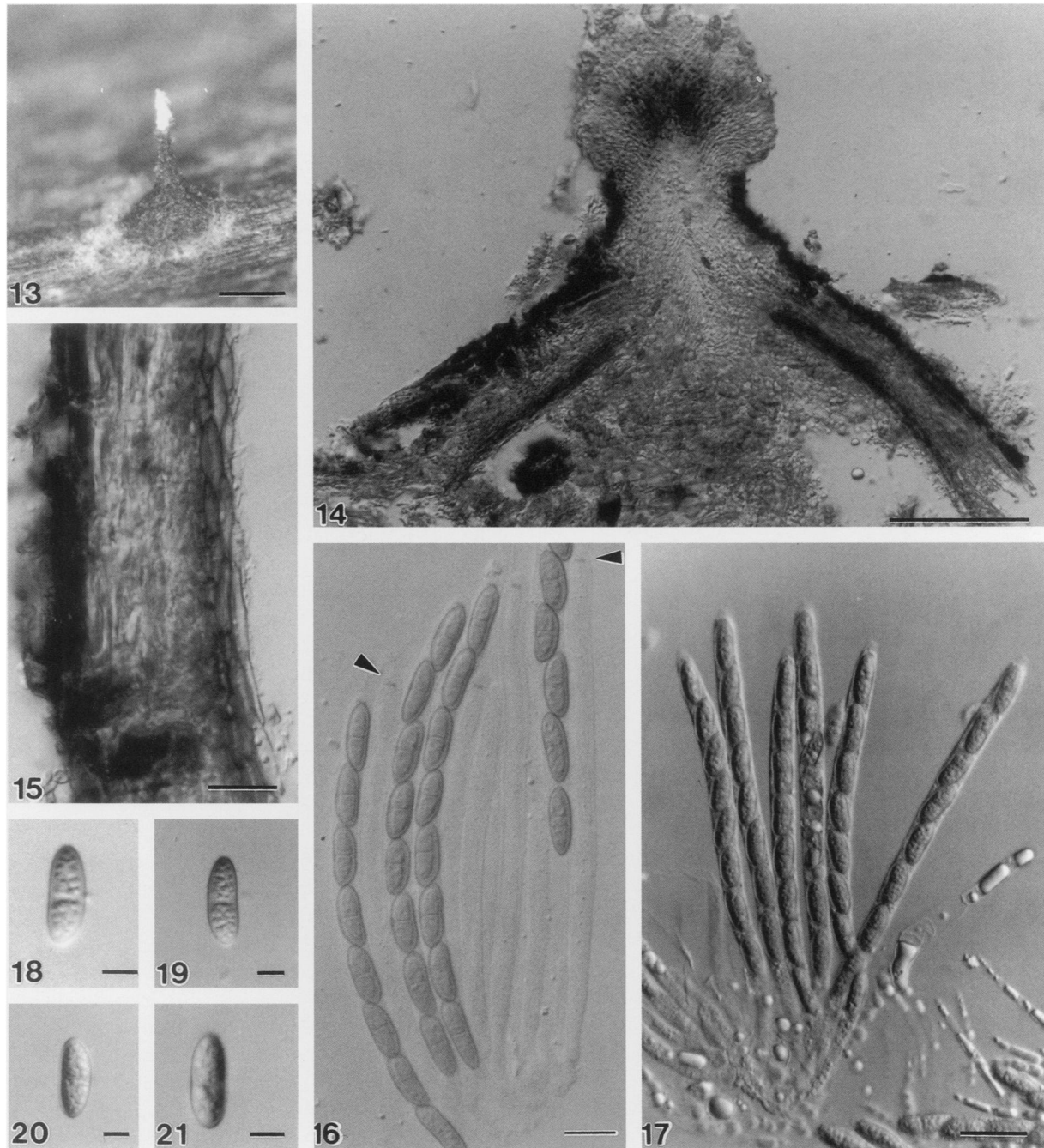
Colonies on potato dextrose agar fast growing, with superficial white hyphae, cottony and flat with crenate edges, white reverse. No conidia or conidiophores produced.

Etymology. From *uniseptata*, in reference to the 1-septate ascospores.

Specimens examined. CHINA. HONG KONG: Tai Po, Lam Tsuen River, on submerged wood, Sep 1997, K. M. Tsui, KM 247 (HOLOTYPE, HKU(M) 8095), cultures from holotype (HKUCC 6579).

Notes. Species in *Clypeosphaeria* Fuckel have clypeate, conspicuously papillate ascomata, long cylindrical asci with subapical, amyloid rings and uniseriate, ellipsoidal, brown ascospores. This genus was discussed by Barr (1989) and three species, *C. mamillana* (Fr.) Lamb., *C. perfidiosa* (De Not.) Barr and *C. americana* Barr & Samuels were included. The genus is now considered to be monotypic because *C. perfidiosa* and *C. americana* were transferred to *Stereosphaeria* Kirschst. (Hyde et al 1998). *Clypeosphaeria mamillana* has a wedge-shaped subapical ring as compared to discoid in *C. uniseptata* and thin-walled ascospores that have one septum near the base (Barr 1989), or have 5 pseudosepta (Hyde et al 1998), as compared to 1 septum in *C. uniseptata*.

Species in *Amphisphaeria* Ces. & De Not. also have clypeate ascomata, long cylindrical asci with amyloid, subapical discoid rings, and ellipsoidal, 1-septate, brown ascospores (Barr 1990), and are characterized by having *Pestalotia*-like anamorphs (Kang et al 1999). We found no evidence of *Pestalotia*-like anamorphs in cultures of *C. uniseptata*, and its characteristic ascomatal morphology warrant the placement of this taxon in *Clypeosphaeria*. *Clypeosphaeria uniseptata* resembles *Amphisphaeria pakistane* E. Müll. &



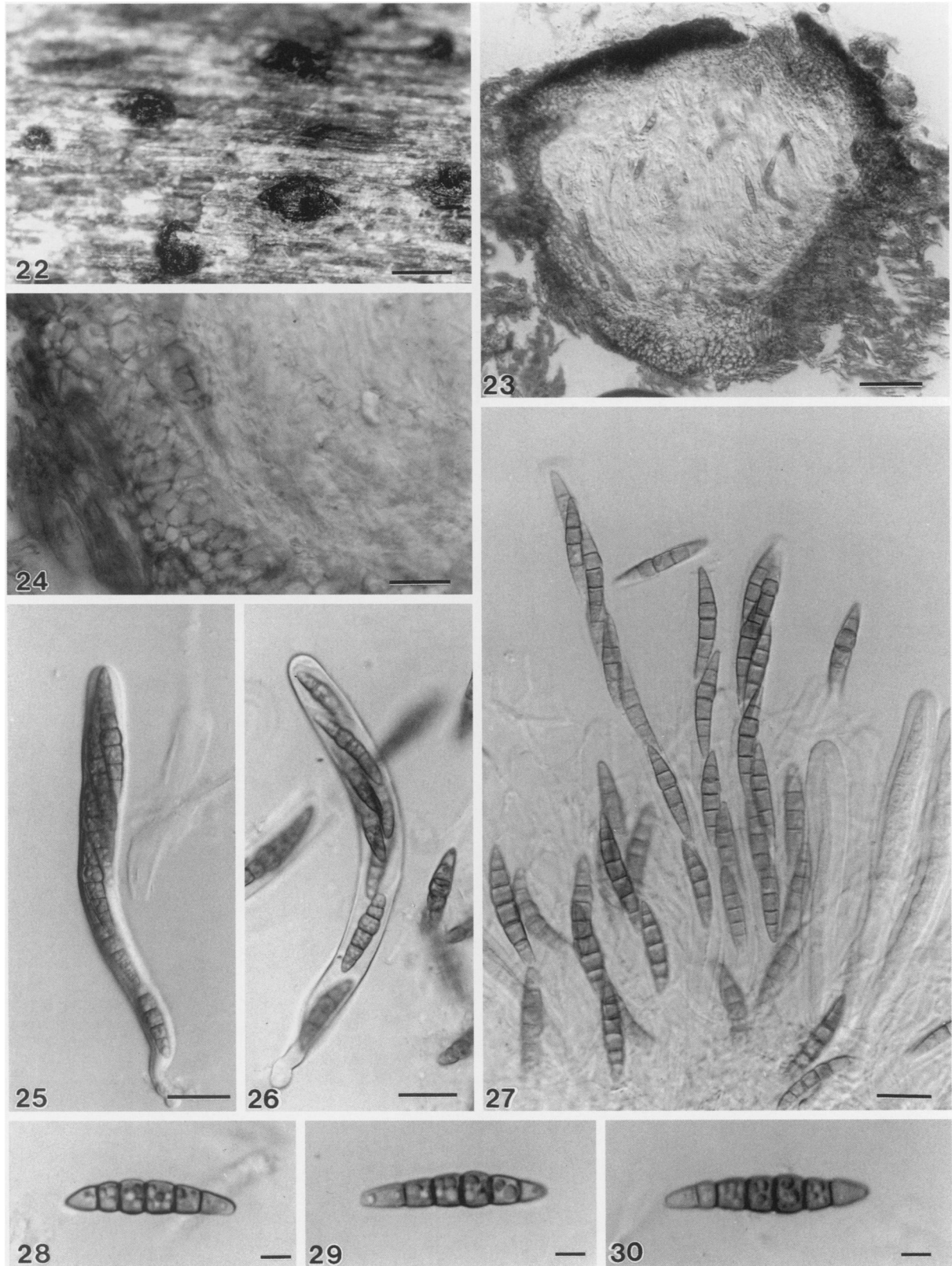
FIGS. 13–21. Light (13) and differential interference contrast (14–21) micrographs of *Clypeosphaeria uniseptata* (from holotype). 13. Appearance of ascomata on wood. 14. Section of the ascoma. 15. Section of the peridium. 16–17. Asci with inconspicuous interascal filaments. Note the amyloid discoid subapical apparatus (stained). 18–21. Ascospores. Scale bars: 13 = 300 μm , 14 = 100 μm , 15 = 15 μm , 16–17 = 10 μm , 18–21 = 5 μm .

Ahmad [ZT 9032, holotype] in producing ellipsoidal, brown, 1-septate ascospores, but is unique in having thick-walled ascospores.

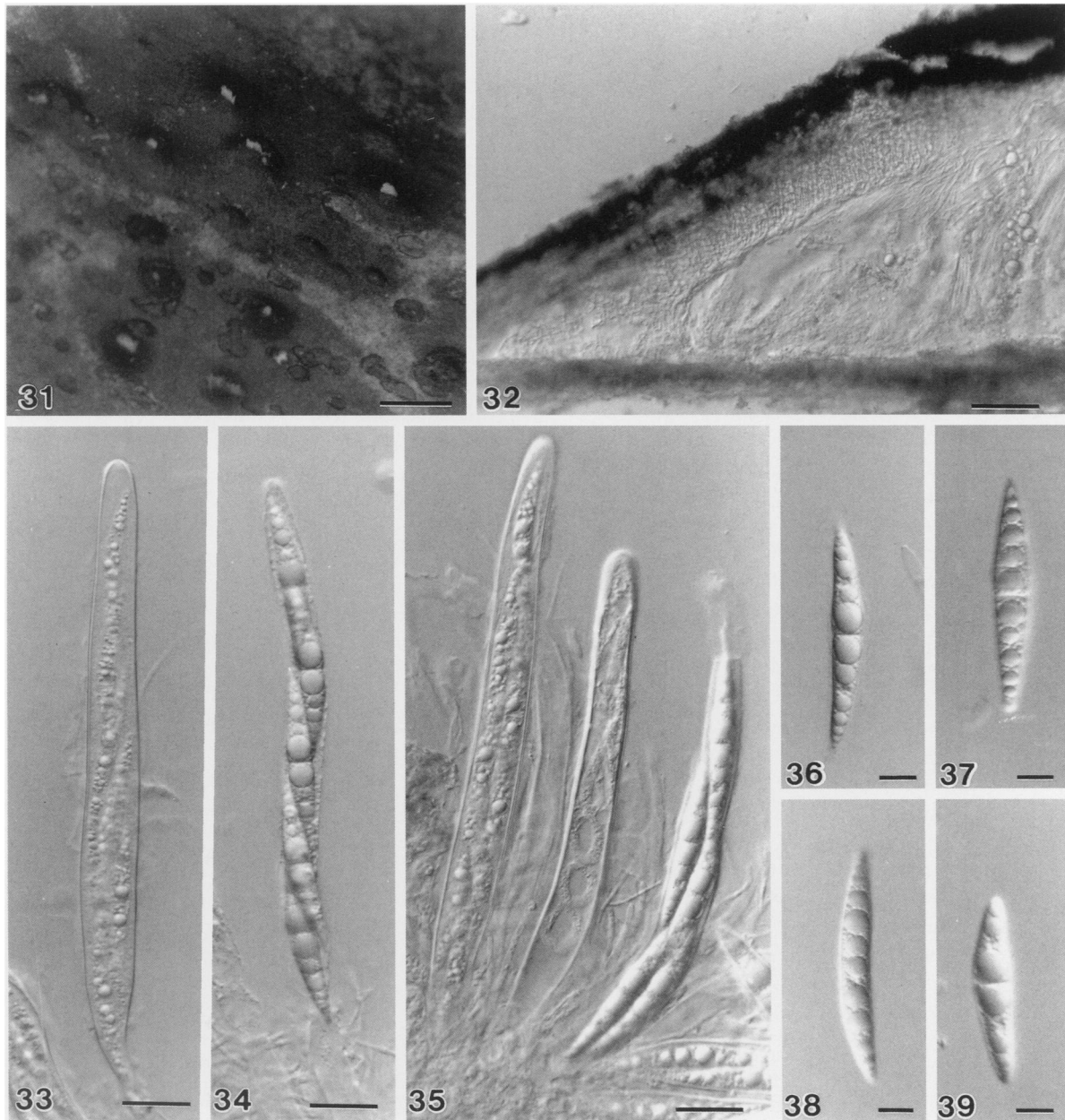
Leptosphaeria ginimia K. M. Tsui, K. D. Hyde et Hodgkiss, sp. nov. FIGS. 22–30

Ascomata 240–300 μm alta, 200–300 μm diam, erumpentia, superficialia vel partim immersa, subglobosa vel globosa,

gregaria, ostiolata, atrobrunnea vel nigra. Paries ascomatis bistratus, 20–30 μm crassus, stratum internum e textura angulari; stratum externum cellulis scleroparenchymaticis, 6–8 \times 3–6 μm , atrobrunneis, compressis compositum. Pseudoparaphyses numerosae, ca 2 μm crassae, septatae, filamentosae. Asci 100–130 \times 10–15 μm , 8-sporei, cylindrici, breve pedicellati, bitunicati, fissitunicati. Ascosporae 27.5–35 \times 6–7.5 μm , biseriatae, 5-(6)-septatae, fusiformes, ad



FIGS. 22-30. Light (22) and differential interference contrast (23-30) micrographs of *Leptosphaeria ginimia* (from holotype). 22. Appearance of ascomata on wood. 23. Section of the ascoma. 24. Section of the peridium. 25-26. Asci. 27. Asci with pseudoparaphyses. 28-30. Ascospores. Scale bars: 22 = 250 μm , 23 = 50 μm , 24 = 10 μm , 25-27 = 20 μm , 28-30 = 5 μm .



FIGS. 31–39. Light (31) and differential interference contrast (32–39) micrographs of *Astrosphaeriella stellata* [from HKU(M) 16118]. 31. Appearance of ascomata on wood. 32. Section of the ascoma. 33–34. Asci. 35. Asci with pseudoparaphyses. 36–39. Ascospores. Scale bars: 31 = 900 μm , 32 = 50 μm , 33 = 15 μm , 34–35 = 20 μm , 36–39 = 8 μm .

septa constrictae, guttulae, fulvae vel brunneae, tunica gelatinosa praeditae.

Ascomata 240–300 μm high, 200–300 μm in diam, erumpent, superficial or partly immersed, subglobose to globose, gregarious, ostiolate, dark brown to black (FIG. 22). Peridium 20–30 μm wide, consisting of two layers: cells of outer layer interspersed with host cells, comprising 3–4 layers of brown polygonal 6–8 \times 3–6 μm scleroparenchymatic cells; inner layer *textura angularis* (FIGS. 23, 24). Pseudoparaphyses cellular, numerous, ca 2 μm wide, septate, filamentous,

and embedded in a transparent gelatinous matrix (FIG. 27). Asci 95–140 \times 9–13 μm (\bar{x} = 121 \times 11 μm , n = 25), 8-spored, cylindrical, short pedicellate, bitunicate, fissitunicate, ectoascus rupturing at the apex (FIGS. 25–27). Ascospores 27.5–35 \times 5.5–7 μm (\bar{x} = 32 \times 6 μm , n = 35), overlapping biserial, 5–(6)-septate, fusiform, swollen at third cell, constricted at the septa, guttulate, yellow to pale brown, surrounded by a mucilaginous sheath (FIGS. 28–30).

Colonies on potato dextrose agar slow growing, mostly immersed, brown to gray reverse, with super-

ficial hyphal growth, fluffy, effuse, with dentate edges, producing yellow pigments. No anamorphs produced.

Etymology. The Latin *ginimia*, derived from Ginimi Chan, for her appreciation and support of first author's work in mycology.

Specimens examined. CHINA. HONG KONG: Tai Po, Lam Tsuen River, on submerged wood, 31 May 2000, *K. M. Tsui*, KM31 (HOLOTYPE, HKU(M) 16115), culture of holotype (HKUCC 6465), Sep 1996, *K. M. Tsui*, KM31 (HKU(M) 4613), (HKU(M) 4627), (HKU(M) 5380), Sep 1997, *K. M. Tsui*, KM31 (HKU(M) 8056), Dec 1998, *K. M. Tsui*, KM31 (HKU(M) 12193).

Notes. *Leptosphaeria* Ces. & De Not. is a heterogeneous genus, and highly resembles *Phaeosphaeria* Miyake which also produces fusiform to cylindrical, bitunicate asci, and yellow, guttulate ascospores with more than 3 septa (Shoemaker and Babcock 1989). This taxon is better placed in *Leptosphaeria* (sensu Holm 1957, Shearer et al 1990, Ahn and Shearer 1997) because it possesses a thick ascomatal wall comprising scleroplectenchymatic cells and was collected from woody substrata.

Leptosphaeria tetonensis (Ellis & Everhart) Rehm resembles *L. ginimia* in producing 5 to 6-septate, brown ascospores with enlarged third cells from apex (Shoemaker 1984), but differs in lacking guttules and a mucilaginous sheath. The ascospores in *L. ginimia* are also yellow to brown while those in *L. tetonensis* are reddish brown (Shoemaker 1984). This taxon is unique among other species in *Leptosphaeria* (Müller 1950, Crivelli 1983, Shoemaker 1984). The ascospores in *Phaeosphaeria luctuosa* (Niessl) Otani & Mikawa are also 5-septate, yellow, of similar size, and have an enlarged third cell from apex (Webster 1957, Ridley 1988). *Phaeosphaeria luctuosa* however, has been reported from grass, and produces smaller ascomata with pseudoparenchymatous cells (Ridley 1988) and *Phaeoseptoria* Speg. and *Stagonospora* (Sacc.) Sacc. anamorphs (Webster 1957, Leuchtmann 1984).

Astrosphaeriella stellata (Pat.) Sacc., Syll. Fung. 24:938. (1928). FIGS. 31–39

Ascomata 200–250 μm high, 800–950 μm diam, superficial, erumpent, dome-shaped to mammiform, with a flattened base, with slit-like ostiole, gregarious, black (FIGS. 31, 32). Peridium 50–100 μm wide, cells of outer layer with blackish and amorphous material; inner layer composed of thin-walled, polygonal cells, cells are flattened at the base (FIG. 32). Pseudoparaphyses trabeculate, numerous, 2–3 μm wide, filamentous, septate (FIG. 35). Asci 150–250 \times 13–19 μm (\bar{x} = 188 \times 15 μm , n = 20), bitunicate, fissitunicate, ectoascus rupturing at the apex, 8-spored, cylindrical to cylindrical-clavate, pedicellate, (FIGS. 33–35). Asco-

spores 48–60 \times 7–9 μm (\bar{x} = 55 \times 7.5 μm , n = 35), overlapping uni- to biseriolate, 1-septate, elongate fusiform, guttulate, constricted at the septum, hyaline, surrounded by a mucilaginous sheath (FIGS. 36–39).

Colonies on corn meal agar (CMA) slow growing, reaching 3 cm diam after two months, superficial hyphal growth low convex, creamy, woolly, white reverse. No anamorphs produced.

Specimens examined. CHINA. HONG KONG: Tai Po, Pak Au Shek, on submerged bamboo in the Lam Tsuen River, Aug 1998, *K. M. Tsui & Jackie Chan*, KM 287 [HKU(M) 16118, culture (HKUCC 6856)], May 2000, *K. M. Tsui*, KM287 (HKU(M) 16113).

Notes. The placement of this taxon in *Astrosphaeriella* Syd. et P. Syd. is suitable because it has trabeculate pseudoparaphyses and hyaline, elongate fusiform ascospores with mucilaginous sheaths. The genus has also been expanded to include species with a slit-like ostiole (Hyde et al 2000) but this character has not been reported from collections of *A. stellata* on terrestrial bamboo (Hawksworth 1981) and palms (Hyde and Fröhlich 1997).

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LITERATURE CITED

- Ahn Y-M, Shearer CA. 1997. Reexamination of taxa in *Leptosphaeria* originally described on host species in Ranunculaceae, Papaveraceae, and Magnoliaceae. *Can J Bot* 76:258–280.
- Barr ME. 1989. *Clypeosphaeria* and the Clypeosphaeriaceae. *Syst Ascomy* 8:1–8.
- . 1990. Prodrum to non-lichenized, pyrenomycetous members of class Hymenoascomycetes. *Mycotaxon* 39:43–184.
- . 1993. Redisposition of some taxa described by J. B. Ellis. *Mycotaxon* 46:45–76.
- Choi YW, Hyde KD, Ho WH. 1999. Single spore isolation of fungi. *Fung Divers* 3:29–38.
- Crivelli PG. 1983. Über die heterogene Ascomycetengattung *Pleospora* Rabh.; Vorschlag für eine Aufteilung. *Diss. ETH Nr. 7318*. Flück-Wirth, Komm.-Verl., Teufen. 213 p.
- Goh TK, Hyde KD. 1996. Biodiversity of freshwater fungi. *J Industr Microbiol* 17:328–345.
- , ———. 1999. Fungi on submerged wood and bamboo in the Plover Cove Reservoir, Hong Kong. *Fung Divers* 3:57–85.
- Hawksworth DL. 1981. *Astrosphaeriella* Sydow, a misunder-

- stood genus of melanmomataceous pyrenomycetes. Bot J Linn Soc 82:35–59.
- Holm L. 1957. Études taxonomiques sur les Pléosporacés. Symb Bot Upsal 14:1–188.
- Hyde KD, Fröhlich J. 1997. Fungi from palms. XXXVII. The genus *Astrosphaeriella*, including ten new species. Sydowia 50:81–132.
- , ——, Taylor JE. 1998. Fungi from palms. XXXVI. Reflections on unitunicate ascomycetes with apiospores. Sydowia 50:21–80.
- , Aptroot A, Fröhlich J, Taylor JE. 2000. Fungi from palms. XLIII. *Lophiostoma* and *Astrosphaeriella* species with slit-like ostioles. Nova Hedwigia 70:143–160.
- Kang JC, Hyde KD, Kong RYC. 1999. Studies on Amphisphaeriales: the Amphisphaeriaceae (*sensu stricto*). Mycol Res 103:53–64.
- Leuchtmann A. 1984. Über *Phaeosphaeria* Miyake und andere bitunicate Ascomyceten mit mehrfach quersgetierten Ascosporen. Sydowia 37:75–184.
- Müller E. 1950. Die schweizerischen Arten der Gattung *Leptosphaeria* und ihrer Verwandten. Sydowia 4:185–319.
- Ridley GS. 1988. New records and species of Loculoascomycetes from New Zealand. New Zealand J Bot 26:409–422.
- Rossmann AY, Samuels GJ, Rogerson CT, Lowen R. 1999. Genera of Bionectriaceae, Hypocreaceae and Nectriaceae (Hypocreales, Ascomycetes). Stud Mycol 42:248.
- Samuels GJ, Barr ME. 1997. Notes on and additions to the Niessliaceae (Hypocreales). Can J Bot 75:2165–2176.
- Scheuer C. 1993. *Cryptoniesslia setulosa* gen. et sp. nov. Mycol Res 97:543–546.
- Shearer CA. 1993. The freshwater ascomycetes. Nova Hedwig 56:1–33.
- , Crane JL, Reddy CKR. 1990. Studies in *Leptosphaeria*. Lectotypification of *Sphaeria doliolum*. Mycologia 82:496–500.
- Shoemaker RA. 1984. Canadian and some extralimital *Leptosphaeria* species. Can J Bot 62:2688–2729.
- , Babcock CE. 1989. Phaeosphaeria. Can J Bot 67:1500–1599.
- Sivanesan A. 1983. Studies on ascomycetes. Trans Brit Mycol Soc 81:313–332.
- Triska FJ, Cromack K. 1980. The role of wood debris in forests and streams. In: Waring RH, ed. Forest: fresh perspectives from ecosystem analysis. Proc 40th Biol Colloq 1979. Corvallis, Oregon: Oregon University Press. p 171–190.
- Tsui CKM, Hyde KD, Hodgkiss IJ. 2000. Biodiversity of fungi on submerged wood in Hong Kong streams. Aqua Microb Ecol 21:289–298.
- Webster J. 1957. Graminicolous pyrenomycetes. IV. Conidia of *Ophiobolus herpotrichus*, *Leptosphaeria luctuosa*, *L. fuckelii*, *L. pontiformis* and *L. eustomoides*. Trans Brit Mycol Soc 40:509–522.
- Wong MKM, Goh TK, Hodgkiss IJ, Hyde KD, Raghoo VM, Tsui CKM, Ho W-H, Wong WSW, Yuen T-K. 1998. Role of fungi in freshwater ecosystem. Biodiv Conserv 7:1187–1206.