

RESEARCH ARTICLE

Open Access

Common variants at 12p11, 12q24, 9p21, 9q31.2 and in *ZNF365* are associated with breast cancer risk for *BRCA1* and/or *BRCA2* mutation carriers

Antonis C Antoniou^{1*}, Karoline B Kuchenbaecker¹, Penny Soucy², Jonathan Beesley³, Xiaoqing Chen³, Lesley McGuffog¹, Andrew Lee¹, Daniel Barrowdale¹, Sue Healey³, Olga M Sinilnikova⁴, Maria A Caligo⁵, Niklas Loman⁶, Katja Harbst⁶, Annika Lindblom⁷, Brita Arver⁸, Richard Rosenquist⁹, Per Karlsson¹⁰, Kate Nathanson¹¹, Susan Domchek¹¹, Tim Rebbeck¹¹, Anna Jakubowska¹², Jan Lubinski¹², Katarzyna Jaworska¹², Katarzyna Durda¹³, Elżbieta Złowowcka-Perłowska¹², Ana Osorio¹⁴, Mercedes Durán¹⁵, Raquel Andrés¹⁶, Javier Benítez¹⁷, Ute Hamann¹⁸, Frans B Hogervorst¹⁹, Theo A van Os²⁰, Senno Verhoef²¹, Hanne EJ Meijers-Heijboer²², Juul Wijnen²³, Encarna B Gómez García²⁴, Marjolijn J Ligtenberg²⁵, Mieke Krieger²⁶, J Margriet Collée²⁷, Margreet GEM Ausems²⁸, Jan C Oosterwijk²⁹, Susan Peock¹, Debra Frost¹, Steve D Ellis¹, Radka Platte¹, Elena Fineberg¹, D Gareth Evans³¹, Fiona Lalloo³¹, Chris Jacobs³², Ros Eeles³³, Julian Adlard³⁴, Rosemarie Davidson³⁵, Trevor Cole³⁶, Jackie Cook³⁷, Joan Paterson³⁸, Fiona Douglas³⁹, Carole Brewer⁴⁰, Shirley Hodgson⁴¹, Patrick J Morrison⁴², Lisa Walker⁴³, Mark T Rogers⁴⁴, Alan Donaldson⁴⁵, Huw Dorkins⁴⁶, Andrew K Godwin⁴⁷, Betsy Bove⁴⁸, Dominique Stoppa-Lyonnet⁴⁹, Claude Houdayer⁵⁰, Bruno Buecher⁵¹, Antoine de Pauw⁵², Sylvie Mazoyer⁵³, Alain Calender⁵⁴, Mélanie Léoné⁵⁴, Brigitte Bressac- de Paillerets⁵⁵, Olivier Caron⁵⁶, Hagay Sobol⁵⁷, Marc Frenay⁵⁸, Fabienne Prieur⁵⁹, Sandra Fert Ferrer⁶⁰, Isabelle Mortemousque⁶¹, Sandra Buys⁶³, Mary Daly⁶⁴, Alexander Miron⁶⁵, Mary Beth Terry⁶⁶, John L Hopper⁶⁷, Esther M John⁶⁸, Melissa Southey⁶⁹, David Goldgar⁷⁰, Christian F Singer⁷¹, Anneliese Fink-Retter⁷¹, Muy-Kheng Tea⁷¹, Daphne Geschwantler Kaulich⁷¹, Thomas VO Hansen⁷², Finn C Nielsen⁷², Rosa B Barkardottir⁷³, Mia Gaudet⁷⁴, Tomas Kirchhoff⁷⁵, Vijai Joseph⁷⁶, Ana Dutra-Clarke⁷⁶, Kenneth Offit⁷⁶, Marion Piedmonte⁷⁷, Judy Kirk⁷⁸, David Cohn⁷⁹, Jean Hurteau⁸⁰, John Byron⁸¹, James Fiorica⁸², Amanda E Toland⁸³, Marco Montagna⁸⁴, Cristina Oliani⁸⁵, Evgeny Imyanitov⁸⁶, Claudine Isaacs⁸⁷, Laima Tihomirova⁸⁸, Ignacio Blanco⁸⁹, Conxi Lazaro⁹⁰, Alex Teulé⁸⁹, J Del Valle⁹⁰, Simon A Gayther⁹¹, Kunle Odunsi⁹², Jenny Gross⁹³, Beth Y Karlan⁹³, Edith Olah⁹⁴, Soo-Hwang Teo⁹⁵, Patricia A Ganz⁹⁶, Mary S Beattie⁹⁷, Cecelia M Dorfling⁹⁸, Elizabeth Jansen van Rensburg⁹⁸, Orland Diez⁹⁹, Ava Kwong¹⁰⁰, Rita K Schmutzler¹⁰¹, Barbara Wappenschmidt¹⁰¹, Christoph Engel¹⁰², Alfons Meindl¹⁰³, Nina Ditsch¹⁰⁴, Norbert Arnold¹⁰⁵, Simone Heidemann¹⁰⁶, Dieter Niederacher¹⁰⁷, Sabine Preisler-Adams¹⁰⁸, Dorothea Gadzicki¹⁰⁹, Raymonda Varon-Mateeva¹¹⁰, Helmut Deissler¹¹¹, Andrea Gehrig¹¹², Christian Sutter¹¹³, Karin Kast¹¹⁴, Britta Fiebig¹¹⁵, Dieter Schäfer¹¹⁶, Trinidad Caldes¹¹⁷, Miguel de la Hoya¹¹⁷, Heli Nevanlinna¹¹⁸, Taru A Muranen¹¹⁸, Bernard Lespérance¹¹⁹, Amanda B Spurdle³, Susan L Neuhausen¹²¹, Yuan C Ding¹²¹, Xianshu Wang¹²², Zachary Fredericksen¹²³, Vernon S Pankratz¹²³, Noralane M Lindor¹²⁴, Paolo Peterlongo¹²⁵, Siranoush Manoukian¹²⁶, Bernard Peissel¹²⁶, Daniela Zaffaroni¹²⁶, Bernardo Bonanni¹²⁷, Loris Bernard¹²⁸, Riccardo Dolcetti¹²⁹, Laura Papi¹³⁰, Laura Ottini¹³¹, Paolo Radice¹²⁵, Mark H Greene¹³², Jennifer T Loud¹³², Irene L Andrulis¹³³, Hilmi Ozcelik¹³⁴, Anna Marie Mulligan¹³⁵, Gord Glendon¹³⁶, Mads Thomassen¹³⁷, Anne-Marie Gerdes¹³⁸, Uffe B Jensen¹³⁹, Anne-Bine Skytte¹⁴⁰, Torben A Kruse¹³⁷, Georgia Chenevix-Trench³, Fergus J Couch¹⁴¹, Jacques Simard¹⁴² and Douglas F Easton¹ and for CIMBA, SWE-BRCA⁶ and for HEBON³⁰ and for EMBRACE¹ and for GEMO Study Collaborators⁶², for kConFab Investigators¹²⁰

* Correspondence: antonis@srl.cam.ac.uk

¹Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Worts Causeway, Cambridge CB1 8RN, UK
Full list of author information is available at the end of the article

Abstract

Introduction: Several common alleles have been shown to be associated with breast and/or ovarian cancer risk for *BRCA1* and *BRCA2* mutation carriers. Recent genome-wide association studies of breast cancer have identified eight additional breast cancer susceptibility loci: rs1011970 (9p21, *CDKN2A/B*), rs10995190 (*ZNF365*), rs704010 (*ZMIZ1*), rs2380205 (10p15), rs614367 (11q13), rs1292011 (12q24), rs10771399 (12p11 near *PTHLH*) and rs865686 (9q31.2).

Methods: To evaluate whether these single nucleotide polymorphisms (SNPs) are associated with breast cancer risk for *BRCA1* and *BRCA2* carriers, we genotyped these SNPs in 12,599 *BRCA1* and 7,132 *BRCA2* mutation carriers and analysed the associations with breast cancer risk within a retrospective likelihood framework.

Results: Only SNP rs10771399 near *PTHLH* was associated with breast cancer risk for *BRCA1* mutation carriers (per-allele hazard ratio (HR) = 0.87, 95% CI: 0.81 to 0.94, *P*-trend = 3×10^{-4}). The association was restricted to mutations proven or predicted to lead to absence of protein expression (HR = 0.82, 95% CI: 0.74 to 0.90, *P*-trend = 3.1×10^{-5} , *P*-difference = 0.03). Four SNPs were associated with the risk of breast cancer for *BRCA2* mutation carriers: rs10995190, *P*-trend = 0.015; rs1011970, *P*-trend = 0.048; rs865686, 2df-*P* = 0.007; rs1292011 2df-*P* = 0.03. rs10771399 (*PTHLH*) was predominantly associated with estrogen receptor (ER)-negative breast cancer for *BRCA1* mutation carriers (HR = 0.81, 95% CI: 0.74 to 0.90, *P*-trend = 4×10^{-5}) and there was marginal evidence of association with ER-negative breast cancer for *BRCA2* mutation carriers (HR = 0.78, 95% CI: 0.62 to 1.00, *P*-trend = 0.049).

Conclusions: The present findings, in combination with previously identified modifiers of risk, will ultimately lead to more accurate risk prediction and an improved understanding of the disease etiology in *BRCA1* and *BRCA2* mutation carriers.

Introduction

Pathogenic mutations in *BRCA1* and *BRCA2* confer high risks of breast and ovarian cancers [1,2]. Several lines of evidence suggest that these risks are modified by other genetic or environmental factors that cluster in families. Direct evidence for genetic modifiers of risk has been provided through studies that investigated the associations between common breast and ovarian cancer susceptibility variants, identified through genome-wide association studies (GWAS) or candidate gene studies in the general population, and cancer risk for *BRCA1* and *BRCA2* mutation carriers [3-8] and through GWAS in *BRCA1* and *BRCA2* mutation carriers [9-11]. Six loci (at *TOX3*, 2q35, 6q25.1, 19p13, *CASP8* and wild-type copy of *BRCA1*) are now known to be associated with breast cancer risk for *BRCA1* mutation carriers; a further 10 loci (at *FGFR2*, *TOX3*, *MAP3K1*, *LSP1*, 2q35, *SLC4A7*, 5p12, 1p11.2, *ZNF365* and *RAD51*) have been associated with breast cancer risk for *BRCA2* carriers. The association patterns between these common variants and breast cancer risk for *BRCA1* and *BRCA2* mutation carriers are in general different, and mostly reflect differences in the associations of these single-nucleotide polymorphism (SNPs) with estrogen receptor (ER) status of breast cancer [12-14].

GWAS in the general population have recently identified eight additional breast cancer susceptibility loci which have not been previously investigated in *BRCA1* and *BRCA2* mutation carriers. Turnbull *et al.* [15] identified five susceptibility loci on chromosomes 9 (rs1011970), 10 (rs2380205, rs10995190, rs704010) and

11 (rs614367) through a GWAS of breast cancer cases with a family history of the disease and unrelated controls. In a further follow-up of additional promising associations from that GWAS, the Breast Cancer Association Consortium (BCAC) has identified two additional loci at 12p11 (rs10771399) and 12q24 (rs1292011) which were associated with breast cancer risk in the general population [16]. The estimated odds ratios (OR) for ER-positive breast cancer for four of these SNPs (rs1011970 near *CDKN2A/CDKN2B* at chromosome 9, rs10995190 in *ZNF365* at chromosome 10, rs614367 at 11q13 and rs1292011 at 12q24) were higher than the OR estimates for ER-negative breast cancer. In contrast, the OR estimates were similar for ER-positive and ER-negative breast cancer for SNPs rs2380205 (near *ANKRD16* and *FBXO18*), rs704010 (upstream of *ZMIZ1*) and rs10771399 near *PTHLH*. In a separate GWAS that included mainly cases with two primary breast cancers or a family history of the disease, SNP rs865686 at 9q31.2 was found to be associated with risk for breast cancer, OR = 0.89 (95% CI: 0.85 to 0.92), but no estimates by ER status were reported [17].

The associations of these eight loci with breast cancer risk for *BRCA1* and *BRCA2* mutation carriers are still unknown. To evaluate these associations, we genotyped the eight SNPs in *BRCA1* and *BRCA2* mutation carriers participating in the Consortium of Investigators of Modifiers of *BRCA1/2* (CIMBA). We further investigated the associations with the risks of developing ER-positive and ER-negative breast cancer and the risk of ovarian cancer.

Materials and methods

Subjects

All carriers participated in clinical or research studies at the host institutions which have been approved by local ethics committees (list provided in Additional file 1 Table S1). Informed consent was obtained from all study participants. Subjects were *BRCA1* and *BRCA2* mutation carriers recruited by 40 study centres in 28 countries through CIMBA (Additional file 1 Table 2). The majority of carriers (97.58%) were recruited through cancer genetics clinics offering genetic testing, and enrolled into national or regional studies. Some carriers were identified by population-based sampling of cases (2.38%), and some by community recruitment (0.04%). Eligibility to participate in CIMBA is restricted to female carriers of pathogenic *BRCA1* or *BRCA2* mutations age 18 years old or older at recruitment. Information collected included the year of birth; mutation description, including nucleotide position and base change; self reported ethnic ancestry, age at last follow-up; ages at breast or ovarian cancer diagnoses; and age or date at bilateral prophylactic mastectomy and oophorectomy. Related individuals were identified through a unique family identifier. Women were included in the analysis if they carried mutations that were pathogenic according to generally recognized criteria [18]. Further details on CIMBA can be found elsewhere [19].

Women who carried pathogenic mutations in both *BRCA1* and *BRCA2* were excluded from the current analysis. The primary analysis was restricted to women self-reported as “white of European ancestry”. The number of mutation carriers of non-white ancestry was too small to allow separate analysis. We investigated possible overlap of carriers between studies by comparing the year of birth, exact mutation description and the reported ages, to identify potential duplicate individuals. Where possible we also used other genotype data on SNPs genotyped in the current round (at least 26 SNPs), in previous genotyping rounds or as part of GWAS to find hidden duplicates. When a potential duplicate was identified, we contacted the relevant centres for further information about these individuals, in a manner that protected the identity of the individuals in question, in order to determine precisely the extent of true overlap in subjects and families appearing more than once in the data set. Duplicate mutation carriers were included only once in the analysis. When in doubt, and when centres could not clarify a potential duplication, one of the samples was excluded from the analysis.

Genotyping

DNA samples (in almost all cases, obtained from blood) were genotyped using the iPLEX Mass Array platform at

four genotyping centres (Additional file 1 Table S2); the iPLEX included 26 SNPs as part of a larger study. All centres included at least 2% of the samples in duplicate, no template controls in every plate, and a random mixture of affected and unaffected carriers. Samples that failed for $\geq 20\%$ of all the SNPs typed (that is, five or more) were excluded from the analysis. A study was included in the analysis only if the call rate was over 95%, after samples that failed at multiple SNPs had been excluded. For each study, genotypes for at least 98% of the duplicate samples had to be concordant. To assess the accuracy of genotyping across genotyping centres, the four centres genotyped 95 DNA samples from a standard test plate (Coriell Institute) for all SNPs. If the genotyping was inconsistent for more than one sample in the test plate, all the studies genotyped at the centre were excluded from the analysis of that SNP. No SNPs failed this criterion. The present study included eight SNPs: rs1011970 (9p21, near *CDKN2A/B*), rs10995190 (10q21, near *ZNF365*), rs704010 (10q22, near *ZMIZ1*), rs2380205 (10p15), rs614367 (11q13), rs1292011 (12q24), rs10771399 (12p11 near *PTHLH*) and rs865686 (9q31.2). Based on the quality control criteria, 4 studies were excluded from the analysis of rs2380205 (one due to low duplicate concordance, 3 due to low call rate), 2 studies were excluded from the analysis of rs704010 (low call rate) and 13 studies were excluded from the analysis of rs1292011 (all due to low call rates). As an additional genotyping quality-control check, we also evaluated the deviation from Hardy-Weinberg equilibrium (HWE) for unrelated subjects separately for each SNP and study. Nine studies had HWE *P*-values in the range 0.005 to 0.05 (two studies for rs10995190, two studies for rs704010, one study for rs10771399, two for rs1292011 and two for rs865686). Upon examination of the cluster plots for these studies and SNPs, none revealed any unusual patterns and these studies were included in all the analyses. After the above exclusions, a total of 19,731 unique mutation carriers (12,599 *BRCA1* and 7,132 *BRCA2*) from 40 studies had an observed genotype for at least 1 of the SNPs and were included in the primary analysis.

Statistical analysis

The aim of the primary analysis was to evaluate the association between each genotype and breast cancer risk within a survival analysis framework. The time variable for each individual was defined to be the time to breast cancer diagnosis. Each individual was followed until the first breast cancer diagnosis, ovarian cancer diagnosis, or bilateral prophylactic mastectomy or the age at last observation. Only those with a first breast cancer diagnosis were considered as affected in the

analysis. Mutation carriers censored at ovarian cancer diagnosis were considered unaffected. Analysis was conducted by modelling the retrospective likelihood of the observed genotypes conditional on the disease phenotypes as previously described [18]. The effect of each SNP was modelled either as a per-allele hazard ratio (HR) (multiplicative model) or as separate HRs for heterozygotes and homozygotes, and these were estimated on the logarithmic scale. The HRs were assumed to be independent of age (that is, we used a Cox proportional-hazards model). The assumption of proportional hazards was tested by adding a “genotype x age” interaction term to the model in order to fit models in which the HR changed with age. Analyses were carried out with the pedigree-analysis software MENDEL [20]; details of this approach have been described previously [18,21]. We examined between-study heterogeneity by comparing the models that allowed for study-specific log-hazard ratios against models in which the same log-hazard ratio was assumed to apply to all studies.

To investigate whether our results were influenced by any of our assumptions we performed additional sensitivity analyses. If a SNP is associated with disease survival, the inclusion of prevalent cases may influence the HR estimates. Current data indicate that five-year survival after a breast cancer diagnosis is over 80% (Cancer Research - UK, Breast cancer survival statistics) and studies have suggested no difference in survival between mutation carriers and non-carriers [22]. We, therefore, repeated our analysis by excluding mutation carriers diagnosed more than five years prior to recruitment into the study. To examine whether SNP associations differed by type of mutation, we classified *BRCA1* mutations according to their potential functional effect [23-26]. Class 1 mutations were those likely to lead to the absence of protein expression due to i) reduced transcript level and/or degradation or instability of truncated proteins, or ii) absence of transcription. Class 1 mutations comprise truncating mutations expected to trigger nonsense-mediated mRNA decay (NMD) or translation re-initiation but no production of stable protein, and deletion of transcription regulatory regions. Class 2 mutations were those likely to generate stable mutant proteins with partial or total loss of function that might also have dominant negative effect. Class 2 mutations include missense substitutions, in-frame deletions and insertions, as well as truncating mutations with premature stop codons occurring in the last exon. Mutations whose consequences at transcript or protein level could not be inferred were not considered for this classification. These were mainly mutations located in splice sites but not characterised for their effect at the transcript level, or large deletions or insertions with undetermined boundaries.

The associations of these SNPs with ovarian cancer risk were evaluated within a competing risk analysis framework [8,9,21], by estimating HRs simultaneously for breast and ovarian cancers. In this model, each individual was at risk of developing either breast or ovarian cancer, by assuming that the probabilities of developing each disease were independent conditional on the underlying genotype. A different censoring process was used for the competing risk analysis, whereby individuals were followed up to the age of the first breast or ovarian cancer diagnosis and were considered to have developed the corresponding disease. No follow-up was considered after the first cancer diagnosis. Individuals were censored for breast cancer at the age of bilateral prophylactic mastectomy and for ovarian cancer at the age of bilateral oophorectomy and were assumed to be unaffected for the corresponding disease. The remaining individuals were censored at the age at last observation and were assumed to be unaffected for both diseases.

We further evaluated the associations of these SNPs with breast cancer subtypes defined by the estrogen receptor (ER) status of the tumours in *BRCA1* and *BRCA2* mutation carriers. The analysis was carried out by an extension of the retrospective likelihood approach to model the simultaneous effect of each SNP on more than one tumour subtype [14]. Briefly, this involves modelling the conditional likelihood of the observed SNP genotypes and tumour subtypes, given the disease phenotypes. Within this framework it is possible to estimate simultaneously the HRs for each tumour subtype and test for heterogeneity in the associations. Only studies that provided tumour pathology information and had genotype information were included in the analysis. To maximise the available information, genotyped mutation carriers that were missing information on tumour characteristics (within each study) were included in the analysis, and their disease subtype was assumed to be missing at random [14]. This is a reasonable assumption given that more than 90% of mutation carriers in our sample were recruited prior to 2007, when it was uncommon to use tumour pathology in selecting individuals for *BRCA1* and *BRCA2* mutation screening.

To ensure a sufficiently large number of mutation carriers within each stratum, we grouped studies from the same country. All analyses were stratified by country of residence and used calendar-year- and cohort-specific cancer incidences for *BRCA1* and *BRCA2* [27]. For sensitivity analyses, strata with small numbers of mutation carriers were grouped. We used a robust variance-estimation approach to allow for the non-independence among related carriers [28].

Results

The analysis included 12,599 *BRCA1* and 7,132 *BRCA2* mutation carriers who were genotyped successfully for

at least one of the eight SNPs. Table 1 summarises the characteristics of the mutation carriers used in the analysis. In evaluating associations with breast cancer, 10,200 mutation carriers had been diagnosed with a first breast cancer diagnosis, 1,869 were censored at an ovarian cancer diagnosis, 561 at age of bilateral prophylactic mastectomy and 7,101 at the age at last observation.

Associations with cancer risk for BRCA1 mutation carriers

Of the eight SNPs, only rs10771399 in *PTHLH* was associated with breast cancer risk for *BRCA1* mutation carriers (P -trend = 3×10^{-4} , Table 2). The association was consistent with a multiplicative model in which each copy of the minor allele was estimated to confer a HR of 0.87 (95% CI: 0.81 to 0.94). There was no evidence of heterogeneity in the HR estimates across studies (P -het = 0.24, Additional file 1 Supplementary Figure 1). There was no evidence that the HRs varied with age ($P = 0.68$). The association remained significant, with a similar HR estimate (HR = 0.85, 95% CI: 0.77 to 0.93, P -trend = 6×10^{-4} , Table 3), when long-term survivors were excluded from the analysis, suggesting no evidence of survival bias. Interestingly, the association was restricted to *BRCA1* carriers of Class 1

mutations (HR = 0.82, 95% CI: 0.74 to 0.90, P -trend = 3×10^{-5} , Table 3) with no evidence of association for Class 2 mutation carriers (HR = 1.00, 0.87 to 1.15, P -trend = 0.99, P -difference between Class 1 and Class 2 = 0.03).

We found no evidence of association between breast cancer risk for *BRCA1* mutation carriers and any of the other SNPs under the trend models (P -trend > 0.15). There was, however, some suggestion of an association under the genotype specific model for rs865686 (2df $P = 0.06$, Table 2), reflecting a lower HR for heterozygous carriers than for either homozygote genotype. There was marginal evidence of heterogeneity in the HRs across countries for rs704010 and rs865686 (P -het = 0.04 for both), but examination of the forest plots revealed that in each case this was mainly due to a single study/country of relatively small sample size, with the majority of the HR estimates being close to 1 (Additional file 1 Supplementary Figure 1). There was no evidence that the HRs varied by age for any of the SNPs ($P > 0.08$ for all).

We further evaluated the SNP associations with breast and ovarian cancer risk simultaneously (Table 4). The associations with breast cancer risk remained

Table 1 Summary characteristics for the 19731 eligible BRCA1 and BRCA2 carriers* used in the analysis

Characteristic	BRCA1		BRCA2	
	Unaffected	Breast cancer	Unaffected	Breast cancer
Number	6,209	6,390	3,322	3,810
Person-years follow-up	264,903	263,068	147,053	168,201
Median age at censure (IQR ¹)	42 (34 to 50)	40 (34 to 47)	43 (34 to 53)	43 (37 to 50)
Age at censure, N (%)				
< 30	1,189 (19.2)	691 (10.8)	611 (18.4)	306 (8.0)
30 to 39	1,161 (26.8)	2,445 (38.3)	834 (25.1)	1,141 (30.0)
40 to 49	1,765 (28.4)	2,191 (34.3)	865 (26.0)	1,394 (36.6)
50 to 59	1,058 (17.0)	812 (12.7)	566 (17.0)	687 (18.0)
60 to 69	380 (6.1)	198 (3.1)	302 (9.1)	226 (5.9)
70+	156 (2.5)	53 (0.8)	144 (4.3)	56 (1.5)
Year of birth, N (%)				
< 1920	28 (0.5)	30 (0.5)	23 (0.7)	44 (1.2)
1920 to 1929	131 (2.1)	196 (3.1)	99 (3.0)	167 (4.4)
1930 to 1939	369 (5.9)	516 (8.1)	232 (7.0)	430 (11.3)
1940 to 1949	832 (13.4)	1,341 (21.0)	458 (13.8)	896 (23.5)
1950 to 1959	1,409 (22.7)	1,989 (31.1)	691 (20.8)	1,160 (30.5)
1960 to 1969	1,703 (27.4)	1,666 (26.1)	902 (27.2)	868 (22.8)
1970+	1,737 (28.0)	652 (10.2)	917 (27.6)	245 (6.4)
Mutation Class, N (%)				
Class 1 ²	4,063 (65.4)	3,878 (60.7)	3,114 (93.7)	3,520 (92.4)
Class 2 ²	1,780 (28.7)	1,973 (30.9)	72 (2.2)	100 (2.6)
Other	366 (5.9)	539 (8.4)	136 (4.1)	190 (5.0)

¹ IQR: Interquartile Range

² See methods for definitions

* Carriers of self-reported white European ancestry only.

Table 2 SNP genotype distributions and associations with breast cancer risk.

Mutation	Genotype	Unaffected N (%)	Affected ^a N (%)	HR	95% CI	P-value
CDK2NA/B - rs1011970						
<i>BRCA1</i>	GG	4,318 (69.7)	4,460 (70.0)	1		
	GT	1,698 (27.4)	1,719 (27.0)	1.01	0.94 to 1.09	
	TT	180 (2.9)	195 (3.1)	1.11	0.91 to 1.35	
	2-df test					0.61
	per allele			1.03	0.96 to 1.09	0.45
<i>BRCA2</i>	GG	2,279 (68.7)	2,586 (67.9)	1		
	GT	943 (28.4)	1,098 (28.9)	1.08	0.98 to 1.19	
	TT	94 (2.8)	123 (3.2)	1.23	0.95 to 1.59	
	2-df test					0.12
	per allele			1.09	1.00 to 1.18	0.048
ZNF365 - rs10995190						
<i>BRCA1</i>	GG	4,394 (70.9)	4,556 (71.5)	1		
	GA	1,656 (26.7)	1,662 (26.1)	0.98	0.91 to 1.06	
	AA	147 (2.4)	156 (2.5)	0.98	0.79 to 1.20	
	2-df test					0.89
	per allele			0.99	0.93 to 1.05	0.64
<i>BRCA2</i>	GG	2,334 (70.4)	2,802 (73.7)	1		
	GA	913 (27.5)	923 (24.3)	0.86	0.78 to 0.96	
	AA	68 (20.1)	79 (2.1)	0.96	0.69 to 1.34	
	2-df test					0.019
	per allele			0.90	0.82 to 0.98	0.015
ZMIZ1 - rs704010						
<i>BRCA1</i>	CC	2,476 (40.3)	2,504 (39.8)	1		
	CT	2,814 (45.8)	2,894 (46.0)	1.03	0.96 to 1.10	
	TT	855 (13.9)	888 (14.1)	1.04	0.93 to 1.15	
	2-df test					0.69
	per allele			1.02	0.97 to 1.07	0.42
<i>BRCA2</i>	CC	1,286 (39.3)	1,443 (38.4)	1		
	CT	1,496 (45.7)	1,779 (47.3)	1.07	0.97 to 1.18	
	TT	494 (15.8)	539 (14.3)	0.99	0.86 to 1.14	
	2-df test					0.32
	per allele			1.01	0.95 to 1.08	0.73
10p15 - rs2380205						
<i>BRCA1</i>	CC	1,609 (32.5)	1,710 (32.1)	1		
	CT	2,410 (48.7)	2,625 (49.3)	1.01	0.93 to 1.09	
	TT	933 (18.8)	990 (18.6)	1.02	0.92 to 1.13	
	2-df test					0.95
	per allele			1.01	0.96 to 1.06	0.75
<i>BRCA2</i>	CC	1,013 (32.8)	1,163 (31.8)	1		
	CT	1,516 (49.1)	1,816 (49.7)	1.05	0.95 to 1.16	
	TT	560 (18.1)	681 (18.6)	1.03	0.90 to 1.17	
	2-df test					0.63
	per allele			1.02	0.96 to 1.09	0.57
11q13 - rs614367						
<i>BRCA1</i>	CC	4,516 (73.2)	4,581 (72.1)	1		
	CT	1,511 (24.5)	1,618 (25.5)	1.05	0.98 to 1.14	
	TT	146 (2.4)	154 (2.4)	1.07	0.87 to 1.32	
	2-df test					0.34
	per allele			1.05	0.98 to 1.12	0.15
<i>BRCA2</i>	CC	2,432 (73.6)	2,723 (71.8)	1		

Table 2 SNP genotype distributions and associations with breast cancer risk. (Continued)

	CT	799 (24.1)	983 (26.0)	1.06	0.96 to 1.17	
	TT	76 (2.3)	83 (2.2)	0.97	0.72 to 1.30	
	2-df test					0.54
	per allele			1.03	0.95 to 1.12	0.46
12q24 - rs1292011						
<i>BRCA1</i>	AA	1,292 (34.3)	1,331 (35.4)	1		
	AG	1,825 (48.4)	1,775 (47.3)	0.98	0.89 to 1.07	
	GG	653 (17.3)	649 (17.3)	1.01	0.90 to 1.14	
	2-df test					0.80
	per allele			1.00	0.94 to 1.06	0.99
<i>BRCA2</i>	AA	824 (35.2)	908 (35.9)	1		
	AG	1,095 (46.7)	1,225 (48.4)	1.03	0.92 to 1.16	
	GG	423 (18.1)	397 (15.7)	0.84	0.72 to 0.99	
	2-df test					0.03
	per allele			0.94	0.87 to 1.01	0.10
PTHLH - rs10771399						
<i>BRCA1</i>	AA	4,913 (79.4)	5,221 (82.0)	1		
	AG	1,194 (19.3)	1,082 (17.0)	0.87	0.80 to 0.95	
	GG	83 (1.3)	65 (1.0)	0.77	0.57 to 1.04	
	2-df test					1.5 × 10⁻³
	per allele			0.87	0.81 to 0.94	3.2 × 10⁻⁴
<i>BRCA2</i>	AA	2,649 (80.0)	3,085 (81.2)	1		
	AG	620 (18.7)	679 (17.9)	0.95	0.85 to 1.07	
	GG	45 (1.4)	34 (0.9)	0.74	0.47 to 1.15	
	2-df test					0.31
	per allele			0.93	0.84 to 1.04	0.20
9q31.2 - rs865686						
<i>BRCA1</i>	TT	2,521 (40.1)	2,640 (41.4)	1		
	TG	2,872 (46.4)	2,849 (44.7)	0.95	0.88 to 1.01	
	GG	799 (12.9)	880 (13.8)	1.05	0.95 to 1.17	
	2-df test					0.06
	per allele			1	0.96 to 1.05	0.85
<i>BRCA2</i>	TT	1,277 (38.6)	1,581 (41.6)	1		
	TG	1,610 (48.6)	1,717 (45.2)	0.86	0.78 to 0.95	
	GG	425 (12.8)	501 (13.2)	0.96	0.84 to 1.11	
	2-df test					7.3 × 10⁻³
	per allele			0.95	0.89 to 1.01	0.10

^a Breast Cancer
 HR, hazard ratio

essentially unchanged in the competing risk analysis, with only the *PTHLH* SNP rs10771399 being significantly associated with breast cancer risk. There was some suggestion of a possible association between this SNP and ovarian cancer risk for *BRCA1* mutation carriers with risk in the opposite direction (HR for ovarian cancer = 1.14, 95% CI: 1.00 to 1.30, *P*-trend = 0.06) especially among rare homozygotes (ovarian cancer HR for GG = 1.67, 95% CI: 1.05 to 2.64, *P*-homozygotes = 0.03). This analysis also provided some weak evidence for an association between SNP rs614367 at 11q13 and ovarian cancer risk for *BRCA1* mutation

carriers under the genotype-specific model (2df *P*-value = 0.03). There was no evidence that any of the other SNPs are associated with ovarian cancer risk for *BRCA1* mutation carriers.

Associations with cancer risk for *BRCA2* mutation carriers

There was evidence of association with breast cancer risk for *BRCA2* mutation carriers for four SNPs (Table 2). The minor allele of rs10995190 in *ZNF365* was associated with a reduced risk of breast cancer, where each copy of allele "A" was estimated to confer a HR of 0.90 (95% CI: 0.82 to 0.98, *P*-trend = 0.015). There was also

Table 3 Associations with breast cancer risk, after excluding prevalent breast cancer cases, and *BRCA1* mutation class.

	Unaffected, N	Affected, N	HR	95% CI	P-value
Excluding prevalent breast cancer cases					
CDK2NA/B -rs1011970					
<i>BRCA1</i>	6,200	3,152	1.05	0.98 to 1.14	0.18
<i>BRCA2</i>	3,319	1,950	1.10	1.00 to 1.22	0.05
ZNF365 - rs10995190					
<i>BRCA1</i>	6,201	3,151	0.96	0.89 to 1.04	0.34
<i>BRCA2</i>	3,318	1,949	0.90	0.81 to 1.00	0.05
ZMIZ1 - rs704010					
<i>BRCA1</i>	6,149	3,094	1.02	0.96 to 1.08	0.53
<i>BRCA2</i>	3,276	1,919	0.98	0.91 to 1.06	0.64
10p15 - rs2380205					
<i>BRCA1</i>	4,955	2,764	1.02	0.95 to 1.08	0.64
<i>BRCA2</i>	3,092	1,884	1.00	0.93 to 1.08	0.92
11q13 - rs614367					
<i>BRCA1</i>	6,177	3,144	1.01	0.94 to 1.10	0.73
<i>BRCA2</i>	3,310	1,944	0.99	0.89 to 1.10	0.88
12q24 - rs1292011					
<i>BRCA1</i>	3,773	1,798	1.04	0.97 to 1.12	0.29
<i>BRCA2</i>	2,345	1,220	0.96	0.88 to 1.06	0.41
PTHLH - rs10771399					
<i>BRCA1</i>	6,194	3,152	0.85	0.77 to 0.93	5.8×10^{-4}
<i>BRCA2</i>	3,317	1,944	0.89	0.78 to 1.00	0.06
9q31.2 - rs865686					
<i>BRCA1</i>	6,196	3,149	1.01	0.95 to 1.07	0.72
<i>BRCA2</i>	3,315	1,946	0.94	0.87 to 1.02	0.15
<i>BRCA1</i> analysis by mutation class					
CDK2NA/B -rs1011970					
Class1	4,040	3,843	1.01	0.94 to 1.10	0.72
Class2	1,771	1,958	1.03	0.91 to 1.16	0.66
ZNF365 - rs10995190					
Class1	4,058	3,844	.99	0.92 to 1.07	0.80
Class2	1,774	1,957	0.97	0.86 to 1.09	0.59
ZMIZ1 - rs704010					
Class1	3,998	3,787	1.04	0.98 to 1.10	0.22
Class2	1,767	1,936	1.01	0.92 to 1.11	0.85
10p15 - rs2380205					
Class1	3,664	3,538	1.01	0.95 to 1.07	0.82
Class2	931	1,263	1.03	0.91 to 1.15	0.67
11q13 - rs614367					
Class1	4,024	3,833	1.10	1.02 to 1.19	0.02
Class2	1,764	1,948	0.94	0.84 to 1.06	0.32
12q24 - rs1292011					
Class1	2,812	2,521	0.99	0.92 to 1.06	0.71
Class2	642	797	0.97	0.84 to 1.12	0.68
PTHLH - rs10771399					
Class1	4,035	3,841	0.82	0.74 to 0.90	3.1×10^{-5}
Class2	1,770	1,953	1.00	0.87 to 1.15	0.99
9q31.2 - rs865686					
Class1	4,038	3,840	0.98	0.92 to 1.04	0.48
Class2	1,769	1,957	1.03	0.94 to 1.14	0.49

HR, hazard ratio

Table 4 Competing risk analysis*.

		Unaffected N (%)	Breast cancer N (%)	Ovarian cancer N (%)	HR	Breast cancer		Ovarian cancer		
						95% CI	P-value	HR	95% CI	P-value
CDK2NA/B - rs1011970										
<i>BRCA1</i>	GG	3,328 (69.6)	4,424 (69.9)	1,026 (70.1)	1			1		
	GT	1,309 (27.4)	1,710 (27.0)	398 (27.2)	1.03	0.95 to 1.11		1.10	0.95 to 1.26	
	TT	142 (3.0)	194 (3.1)	39 (2.7)	1.09	0.88 to 1.35		0.90	0.61 to 1.32	
	2-df test per allele						0.57			0.35
<i>BRCA2</i>	GG	1,972 (68.6)	2,578 (67.9)	315 (69.5)	1			1		
	GT	815 (28.4)	1,097 (28.9)	129 (28.5)	1.09	0.98 to 1.21		1.07	0.85 to 1.35	
	TT	86 (3.0)	122 (3.2)	9 (2.0)	1.19	0.91 to 1.57		0.84	0.40 to 1.77	
	2-df test per allele						0.15			0.74
					1.09	1.00 to 1.19	0.05	1.03	0.84 to 1.25	0.81
ZNF365 - rs10995190										
<i>BRCA1</i>	GG	3,408 (71.3)	4,523 (71.5)	1,019 (69.6)	1			1		
	GA	1,258 (26.3)	1,650 (26.1)	410 (28.0)	1.00	0.93 to 1.08		1.12	0.98 to 1.28	
	AA	113 (2.4)	155 (2.5)	35 (2.4)	0.96	0.78 to 1.20		0.90	0.61 to 1.33	
	2-df test per allele						0.94			0.23
<i>BRCA2</i>	GG	2,033 (70.4)	2,795 (73.7)	318 (70.2)	1			1		
	GA	794 (27.7)	920 (24.3)	122 (26.9)	0.86	0.78 to 0.96		0.99	0.78 to 1.25	
	AA	55 (1.9)	79 (2.1)	13 (2.9)	1.03	0.74 to 1.43		1.58	0.83 to 3.03	
	2-df test per allele						0.02			0.37
					0.90	0.82 to 0.99	0.03	1.06	0.87 to 1.31	0.55
ZMIZ1 - rs704010										
<i>BRCA1</i>	CC	1,904 (40.2)	2,493 (40.0)	583 (40.1)	1			1		
	CT	2,172 (45.9)	2,871 (46.0)	665 (45.7)	1.02	0.95 to 1.10		1.01	0.89 to 1.15	
	TT	660 (13.9)	877 (14.1)	206 (14.2)	1.02	0.92 to 1.14		1.01	0.84 to 1.22	
	2-df test per allele						0.67			0.99
<i>BRCA2</i>	CC	1,109 (39.0)	1,439 (38.4)	181 (40.1)	1			1		
	CT	1,306 (46.0)	1,774 (47.3)	192 (43.4)	1.06	0.96 to 1.17		0.96	0.76 to 1.20	
	TT	426 (15.0)	538 (14.3)	69 (15.6)	1.00	0.86 to 1.15		1.05	0.77 to 1.43	
	2-df test per allele						0.46			0.81
					1.01	0.95 to 1.08	0.72	1.01	0.87 to 1.18	0.90
10p15 - rs2380205										
<i>BRCA1</i>	CC	1,183 (32.2)	1,698 (32.1)	438 (33.2)	1			1		
	CT	1,796 (48.9)	2,605 (49.3)	634 (48.1)	0.99	0.91 to 1.08		0.91	0.79 to 1.05	
	TT	696 (18.9)	981 (18.6)	246 (18.7)	1.00	0.90 to 1.12		0.94	0.78 to 1.14	
	2-df test per allele						0.96			0.45
<i>BRCA2</i>	CC	872 (32.6)	1,161 (31.8)	143 (33.7)	1			1		
	CT	1,321 (49.4)	1,812 (49.6)	199 (46.9)	1.04	0.94 to 1.16		0.94	0.74 to 1.19	
	TT	481 (18.0)	678 (18.6)	82 (19.3)	1.03	0.90 to 1.18		1.01	0.75 to 1.38	
	2-df test per allele						0.74			0.82
					1.02	0.95 to 1.09	0.61	1.00	0.85 to 1.16	0.98
11q13 - rs614367										
<i>BRCA1</i>	CC	3,439 (72.3)	4,547 (72.1)	1,111 (76.2)	1			1		
	CT	1,212 (25.5)	1,606 (25.5)	311 (21.3)	1.02	0.94 to 1.11		0.83	0.72 to 0.96	
	TT	109 (2.3)	154 (2.4)	37 (2.5)	1.12	0.91 to 1.39		1.20	0.81 to 1.76	
	2-df test						0.52			0.03

Table 4 Competing risk analysis*. (Continued)

	per allele				1.03	0.91 to 1.10	0.35	0.91	0.80 to 1.03	0.13
<i>BRCA2</i>	CC	2,106 (73.5)	2,716 (71.9)	333 (73.8)	1			1		
	CT	693 (24.2)	981 (26.0)	108 (24.0)	1.05	0.95 to 1.16		0.95	0.74 to 1.21	
	TT	66 (2.3)	83 (2.2)	10 (2.2)	0.96	0.71 to 1.28		0.87	0.46 to 1.63	
	2-df test						0.62			0.84
	per allele				1.03	0.94 to 1.12	0.56	0.94	0.77 to 1.15	0.56
12q24 - rs1292011										
<i>BRCA1</i>	AA	997 (34.1)	1,321 (35.4)	305 (34.9)	1			1		
	AG	1,406 (48.2)	1,765 (47.3)	429 (49.1)	1.00	0.90 to 1.10		1.11	0.93 to 1.31	
	GG	517 (17.7)	645 (17.3)	140 (16.0)	1.01	0.89 to 1.15		0.98	0.78 to 1.24	
	2-df test						0.97			0.39
	per allele				1.00	0.94 to 1.07	0.91	1.01	0.91 to 1.13	0.82
<i>BRCA2</i>	AA	715 (35.0)	907 (35.9)	110 (36.5)	1			1		
	AG	961 (47.0)	1,222 (48.4)	137 (45.5)	1.02	0.90 to 1.15		0.94	0.71 to 1.25	
	GG	370 (18.1)	396 (15.7)	54 (17.9)	0.83	0.70 to 0.97		0.89	0.61 to 1.28	
	2-df test						0.03			0.80
	per allele				0.93	0.86 to 1.00	0.07	0.94	0.78 to 1.13	0.51
PTHLH - rs10771399										
<i>BRCA1</i>	AA	3,810 (79.8)	5,179 (81.9)	1,145 (78.4)	1			1		
	AG	909 (19.0)	1,078 (17.1)	289 (19.8)	0.89	0.81 to 0.97		1.09	0.93 to 1.26	
	GG	56 (1.2)	65 (1.0)	27 (1.9)	0.86	0.63 to 1.16		1.67	1.05 to 2.64	
	2-df test						0.02			0.06
	per allele				0.90	0.83 to 0.97	6.4 × 10⁻³	1.14	1.00 to 1.30	0.06
<i>BRCA2</i>	AA	2,289 (79.7)	3,076 (81.2)	369 (81.5)	1			1		
	AG	545 (19.0)	678 (17.9)	76 (16.8)	0.94	0.84 to 1.06		0.88	0.67 to 1.16	
	GG	37 (1.3)	34 (0.9)	8 (1.8)	0.79	0.49 to 1.26		1.48	0.63 to 3.46	
	2-df test						0.38			0.43
	per allele				0.93	0.84 to 1.04	0.19	0.96	0.75 to 1.23	0.75
9q31.2 - rs865686										
<i>BRCA1</i>	TT	1,935 (40.5)	2,621 (41.5)	605 (41.3)	1			1		
	TG	2,206 (46.2)	2,825 (44.7)	690 (47.1)	0.94	0.88 to 1.01		0.99	0.87 to 1.12	
	GG	633 (13.3)	877 (13.9)	169 (11.5)	1.03	0.93 to 1.15		0.85	0.70 to 1.03	
	2-df test						0.12			0.23
	per allele				1.00	0.95 to 1.05	0.88	0.94	0.86 to 1.03	0.17
<i>BRCA2</i>	TT	1,103 (38.4)	1,576 (41.6)	179 (39.6)	1			1		
	TG	1,400 (48.8)	1,712 (45.2)	215 (47.6)	0.85	0.77 to 0.94		0.91	0.73 to 1.14	
	GG	367 (12.8)	501 (13.2)	58 (12.8)	0.97	0.84 to 1.12		0.98	0.71 to 1.35	
	2-df test						4.6 × 10⁻³			0.70
	per allele				0.94	0.88 to 1.01	0.10	0.97	0.83 to 1.13	0.67

Associations with breast and ovarian cancer risk for *BRCA1* and *BRCA2* carriers.

*Censoring process described in the methods

HR, hazard ratio

some marginal evidence that the minor allele of rs1011970 near *CDKN2A/CDKN2B* was associated with increased breast cancer risk (HR = 1.09, 95% CI: 1.00 to 1.18, *P*-trend = 0.048). None of the other polymorphisms was associated with breast cancer risk for *BRCA2* mutation carriers under the multiplicative model. However, SNPs rs865686 and rs1292011 were associated with risk under the genotype specific model (2df-*P* = 0.007 and 0.03 respectively, Table 2). There was some

evidence of heterogeneity in the HRs across countries for rs1011970 (*P*-het = 0.005). This appeared to be mainly due to the USA stratum. The heterogeneity was no longer significant after removal of that stratum (*P*-het = 0.42) and the HR estimate for the association with breast cancer risk increased to 1.20 (95% CI: 1.09 to 1.32, *P*-trend = 1×10^{-4}). There was no heterogeneity for any of the other SNPs (*P*-het > 0.12 for all, Additional file 1 Supplementary Figure 2). The HR estimates

for the four associated SNPs were similar when long-term survivors were excluded from the analysis (Table 3). Consistent with the results of the main analysis, rs10995190 in *ZNF365* and rs1011970 near *CDKN2A/CDKN2B* provided marginal evidence of association using the trend-test statistic (P -trend = 0.05 for both) and SNPs rs865686 was associated with breast cancer risk under the genotype specific model (2df- P = 0.03). SNP rs1292011 was not associated with breast cancer risk in this analysis. A somewhat smaller HR estimate was obtained for the *PTHLH* SNP rs10771399 compared to the main analysis (per-allele HR = 0.89, 95% CI: 0.78 to 1.00, P -trend = 0.06). The attenuation of the association in the overall analysis could have occurred if the SNP is also associated with prognosis. However, the difference in the HRs was small. The results for the remaining SNPs were similar and non-significant. None of SNPs were associated with ovarian cancer risk for *BRCA2* mutation carriers (Table 4).

Associations by tumour ER-status

Table 5 summarises the associations of the eight SNPs with breast cancer ER status in *BRCA1* and *BRCA2* mutation carriers. Only the *PTHLH* SNP rs10771399 was associated with ER-negative breast cancer for *BRCA1* mutation carriers (ER-negative HR = 0.81, 95% CI: 0.74 to 0.90, P -trend = 3.8×10^{-5}). There was also marginal evidence that SNP rs704010 near *ZMIZ1* was associated with ER-positive breast cancer for *BRCA1* mutation carriers (ER-positive HR = 1.12, 95% CI: 1.00 to 1.26, P -trend = 0.046). However, the associations between ER-negative and ER-positive breast cancer among *BRCA1* mutation carriers were only significantly different for SNP rs1292011 at 12q24 (P -heterogeneity = 0.045).

Despite the small number of *BRCA2* ER-negative breast cancers, there was a suggestion that the minor allele of the *PTHLH* SNP rs10771399 is protective for ER-negative breast cancer for *BRCA2* mutation carriers (HR for ER-negative = 0.78, 95% CI: 0.62 to 1.00, P -

Table 5 Associations with estrogen receptor-positive and estrogen receptor-negative breast cancer risk for *BRCA1* and *BRCA2* carriers.

	Unaffected N	ER-positive N	ER-negative N	ER status unknown N	ER-positive			ER-negative			P-dif
					HR	95% CI	P-value	HR	95% CI	P-value	
CDK2NA/B - rs1011970											
<i>BRCA1</i>	4,893	559	1,888	2,841	0.95	0.81 to 1.12	0.56	1.03	0.95 to 1.11	0.47	0.41
<i>BRCA2</i>	2,928	1,372	424	1,649	1.10	1.00 to 1.22	0.05	1.15	0.96 to 1.37	0.12	0.70
ZNF365 - rs10995190											
<i>BRCA1</i>	4,895	559	1,887	2,843	0.88	0.74 to 1.04	0.14	1.01	0.94 to 1.10	0.75	0.16
<i>BRCA2</i>	2,927	1,370	406	1,648	0.89	0.80 to 1.00	0.043	0.87	0.71 to 1.07	0.19	0.84
ZMIZ1 - rs704010											
<i>BRCA1</i>	4,842	548	1,846	2,811	1.12	1.00 to 1.26	0.046	1.00	0.94 to 1.06	0.91	0.08
<i>BRCA2</i>	2,887	1,347	401	1,636	1.01	0.93 to 1.09	0.91	1.00	0.87 to 1.14	0.95	0.91
10p15 - rs2380205											
<i>BRCA1</i>	4,465	540	1,812	2,513	0.90	0.80 to 1.01	0.08	1.02	0.96 to 1.09	0.46	0.06
<i>BRCA2</i>	2,701	1,341	396	1,543	1.02	0.95 to 1.10	0.60	0.94	0.82 to 1.08	0.39	0.31
11q13 - rs614367											
<i>BRCA1</i>	4,879	557	1,886	2,832	1.09	0.93 to 1.29	0.30	1.04	0.96 to 1.12	0.40	0.59
<i>BRCA2</i>	2,921	1,365	405	1,639	1.06	0.96 to 1.17	0.26	0.84	0.69 to 1.04	0.10	0.05
12q24 - rs1292011											
<i>BRCA1</i>	3,429	308	1,043	2,031	0.87	0.74 to 1.02	0.09	1.05	0.98 to 1.13	0.12	0.046
<i>BRCA2</i>	2,065	813	239	1,170	0.95	0.86 to 1.04	0.28	0.98	0.82 to 1.16	0.78	0.79
PTHLH - rs10771399											
<i>BRCA1</i>	4,889	557	1,887	2,842	0.94	0.78 to 1.13	0.52	0.81	0.74 to 0.90	3.8×10^{-5}	0.20
<i>BRCA2</i>	2,926	1,366	406	1,648	0.97	0.86 to 1.10	0.68	0.78	0.62 to 1.00	0.049	0.12
9q31.2 - rs865686											
<i>BRCA1</i>	4,892	559	1,888	2,836	0.92	0.81 to 1.03	0.15	1.01	0.95 to 1.08	0.68	0.16
<i>BRCA2</i>	2,924	1,370	405	1,645	0.91	0.84 to 0.99	0.028	1.07	0.92 to 1.25	0.40	0.08

ER, estrogen receptor; HR, hazard ratio

trend = 0.049), but there was no association with ER-positive breast cancer. There was evidence that SNPs rs10995190 near *ZNF365*, rs865686 at 9q31.2 and rs1011970 near *CDKN2A/B* are associated with ER-positive breast cancer for *BRCA2* mutation carriers (P -trend = 0.043, 0.028 and 0.05 respectively). However, the HR estimates were not significantly different from those for ER-negative breast cancer.

Discussion

We have investigated eight novel breast cancer susceptibility loci identified through breast cancer GWAS [15-17] for their associations with breast cancer risk for *BRCA1* and *BRCA2* mutation carriers using data from the CIMBA. The estimated per-allele ORs associated with the minor allele of each SNP from the population-based studies varied from 0.85 to 1.15, and only four of the eight SNPs had ORs of less than 0.90 or greater than 1.10 (rs10995190, rs614367, rs865686 and rs10771399) [15-17]. For *BRCA1* mutation carriers, only SNP rs10771399 at 12p11 was associated with the overall risk of breast cancer, whereas SNPs rs10995190 at 10q21, rs1011970 at 9p21, rs865686 at 9q31.2 and rs1292011 at 12q24 were associated with breast cancer risk for *BRCA2* mutation carriers. The magnitude of the estimated HRs for all these SNPs were consistent with the OR estimates for the risk of breast cancer in the general population. The power to detect associations with SNPs conferring relative risks in the range of 0.90 to 1.10 was limited by our sample size, particularly among *BRCA2* mutation carriers [29].

Based on the HR estimates and associated 95% confidence intervals, given our sample size of *BRCA1* mutation carriers, it is unlikely that the relative risks for overall *BRCA1* breast cancer risk are of similar magnitude to those estimated in the general population for SNPs rs10995190 at 10q21 (estimated odds ratio (OR) from the replication stage of the GWAS = 0.76), rs2380205 at 10p15 (OR = 0.94), rs614367 at 11q13 (OR = 1.15), rs1292011 at 12q24 (OR = 0.92) and rs865686 at 9q31.2 (OR = 0.89), since the 95% confidence intervals for the HRs do not include the estimated OR from the population-based studies. Similarly, the HRs for *BRCA2* breast cancer risk exclude the ORs from the general population for SNPs rs2380205 at 10p15 and rs614367 at 11q13. Taken together, these findings suggest that SNPs rs2380205 at 10p15 and rs614367 at 11q13 do not modify breast cancer risk in either *BRCA1* or *BRCA2* mutation carriers. A replication study by BCAC, involving close to 50,000 breast cancer cases and 50,000 controls, found only weak evidence for association of rs2380205 at 10p15 with breast cancer risk in the general population [Lambrechts and Easton personal communication, manuscript submitted] suggesting that

the original finding (OR = 0.94, $P = 5 \times 10^{-7}$ [15]) may have been a false positive. If this were true, the absence of an association in carriers would be expected. The lack of evidence for an association with the 11q13 SNP rs614367 with *BRCA1* and *BRCA2* breast cancer risk is more surprising since the association in the general population is relatively strong and consistently replicated (OR 1.21, 95% CI 1.17 to 1.25 in the recent BCAC analysis [Lambrechts and Easton personal communication, manuscript submitted]). The association in the general population appears to be restricted to ER-positive disease, which would explain the lack of association for *BRCA1* carriers but not *BRCA2* carriers. This is perhaps the clearest evidence so far of a departure from a multiplicative interaction between a common susceptibility locus and a *BRCA2* mutation on the risk of developing breast cancer. The lack of an association in *BRCA1* carriers for rs1292011 and rs865686 is also consistent with the observation that these associations are stronger for ER-positive disease in the general population [16]. The absence of an association for *ZNF365* rs10995190 in *BRCA1* carriers is more surprising since this association appears to be unrelated to ER status in the general population [Lambrechts and Easton personal communication, manuscript submitted] [30].

Of the eight SNPs investigated, the strongest association was found between SNP rs10771399 at 12p11 and breast cancer risk for *BRCA1* mutation carriers. Other loci previously found to be associated with *BRCA1* breast cancer risk include the 19p13 and 6q25.1 loci [6,9], *TOX3* and *CASP8* [3,5,7]. Analysis by tumour ER-status revealed that rs10771399 at 12p11 has a stronger association with ER-negative than ER-positive breast cancer for both *BRCA1* and *BRCA2* mutation carriers. The ER-specific HRs were similar for both genes, suggesting that this SNP is primarily associated with ER-negative breast cancer, although results from the general population suggested similar ORs for ER-positive and ER-negative breast cancer (0.87 for ER-positive disease, 0.85 for ER-negative disease [16]). Interestingly, the association among *BRCA1* mutation carriers was restricted to those carrying mutations proven or predicted to lead to absence of protein expression (Class 1) with no evidence for an association in carriers of *BRCA1* mutations likely to generate stable mutant proteins (Class 2) (P -diff = 0.03). This observation suggests that the modifying effect of SNP rs10771399 at 12p11 might be attenuated for tumours that retain residual *BRCA1* function or that retain the capacity to bind to some of its partners. rs10771399 lies in a region at 12p11 that contains *PTHLH* (parathyroid hormone-like hormone isoform 1, also known as *PTHRP* - parathyroid hormone-related protein) and *CCDC91*. *PTHLH* is a plausible candidate cancer susceptibility gene. It encodes

a protein that regulates endochondral bone development and epithelial-mesenchymal interactions during the formation of the mammary glands. The receptor of this hormone, PTHR1, is responsible for most cases of humoral hypercalcemia of malignancy [31]. It is produced by various types of carcinomas [32], and is an important factor in the development of bone metastasis [33].

We found that SNP rs10995190 *ZNF365* is associated with *BRCA2* breast cancer risk. A different SNP (rs16917302) in *ZNF365*, which is only weakly correlated with rs10995190 (pairwise r^2 is approximately 0.10 in the present sample) was previously identified via a GWAS of breast cancer in *BRCA2* mutation carriers [10]. These results suggest that there could be a causal associated variant correlated with both rs10995190 and rs16917302, or alternatively more than one causal disease variant in this locus. SNP rs10995190 has also recently been found to be associated with mammographic density in the general population [34]. Previous studies found that mammographic density modifies breast cancer risk for *BRCA2* mutation carriers [35], raising the possibility that this locus modifies breast cancer risk for *BRCA2* mutation carriers through its influence on mammographic density. However, mammographic density has also been shown to modify the breast cancer risk for *BRCA1* carriers, which also makes the absence of association for rs10995190 in *BRCA1* carriers somewhat surprising. Mammographic density data are not available in the CIMBA sample to test this hypothesis explicitly.

There was no evidence of association with ovarian cancer risk for *BRCA1* or *BRCA2* mutation carriers for any of the SNPs, with the exception of some weak evidence for SNPs rs10771399 and rs614367 for *BRCA1* carriers. This is not surprising, since all SNPs were selected on the basis of prior evidence of association with breast cancer risk in the general population and none of these SNPs have so far been found to be associated with ovarian cancer in general population through the ongoing GWAS [36-38].

Conclusions

The per-allele HRs estimated for each of the associated loci in the present report are modest, and in isolation would have only a small impact on the absolute risks of developing breast cancer. However, we have shown previously that modifier SNPs in combination can result in large differences in the absolute risk of developing breast cancer for carriers at the extreme percentiles of the combined SNP distribution [5,39]. Furthermore, the causal variants underlying these loci may confer larger relative risks. Considering all reported modifying loci by the CIMBA consortium, there are now six loci in total

that are associated with breast cancer risk for *BRCA1* mutation carriers (19p13, 6q25.1, 12p11, *TOX3*, 2q35 and *CASP8*) and 13 loci which are known to be associated with *BRCA2* breast cancer risk (*FGFR2*, *TOX3*, *MAP3K1*, *LSP1*, 2q35, *SLC4A7*, 5p12, 1p11.2, *ZNF365*, *CDKN2A/B*, 9q31.2, 12q24 and *RAD51*). Ongoing GWAS in *BRCA1* and *BRCA2* mutation carriers and in the general population are likely to identify further modifier loci and taken together, they may lead to more accurate risk predictions in mutation carriers with implications for clinical management, and to a better understanding of the biology of tumour development in mutation carriers.

Additional material

Additional file 1: Supplementary tables and figures. Table S1 List of local ethics committees that granted approval for the access and use of the data in current study. Supplementary figure 1 Forest plot of the country-specific per-allele HR estimates for breast cancer for *BRCA1* mutation carriers. Supplementary figure 2 Forest plot of the country-specific per-allele HR estimates for breast cancer for *BRCA2* mutation carriers.

Abbreviations

BCAC: Breast Cancer Association Consortium; CIMBA: Consortium of Investigators of Modifiers of *BRCA1/2*; ER: estrogen receptor; GWAS: genome-wide association studies; HR: hazard ratio; HWE: Hardy-Weinberg equilibrium; NMD: nonsense-mediated mRNA decay; OR: odds ratio; SNPs: single-nucleotide polymorphism.

Acknowledgements

This work was supported by Cancer Research UK grants C12292/A11174 and C1287/A10118. The research leading to these results has received funding from the European Community's Seventh Framework Programme under grant agreement n° 223175 (HEALTH-F2-2009-223175), from the Canadian Institutes of Health Research for the "CIHR Team in Familial Risks of Breast Cancer" program and by the Canadian Breast Cancer Research Alliance-grant #019511. This research was also supported by NIH grant CA128978, an NCI Specialized Program of Research Excellence (SPORE) in Breast Cancer (CA116201), a U.S. Department of Defence Ovarian Cancer Idea award (W81XWH-10-1-0341) and grants from the Breast Cancer Research Foundation and the Komen Foundation for the Cure. ACA is a CR-UK Senior Cancer Research Fellow, DFE is CR-UK Principal Research Fellow, GCT is a NHMRC Senior Principal Research Fellow, J.S. is Chairholder of the Canada Research Chair in Oncogenetics.

Study specific

Baltic Familial Breast and Ovarian Cancer Consortium

We acknowledge the Genome Database of Latvian Population, Latvian Biomedical

Research and Study Centre and Ramunas Janavicius (Vilnius University Hospital Santariskiu Clinics, Lithuania) for data and DNA samples for BFOCC. The work was supported in part by a grant from the European Social Fund Nr.2009/0220/1DP/1.1.1.2.0/09/APIA/VIAA/016.

BMBSA was supported by grants from the Cancer Association of South Africa (CANSA) to Elizabeth J. van Rensburg.

Breast Cancer Family Registry (BCFR)

This work was supported by the National Cancer Institute, National Institutes of Health under RFA-CA-06-503 and through cooperative agreements with members of the Breast Cancer Family Registry (BCFR) and Principal Investigators, including Cancer Care Ontario (U01 CA69467), Columbia University (U01 CA69398), Fox Chase Cancer Center (U01 CA69631), Huntsman Cancer Institute (U01 CA69446), Cancer Prevention Institute of

California (formerly the Northern California Cancer Center) (U01 CA69417), University of Melbourne (U01 CA69638), and Research Triangle Institute Informatics Support Center (RFP No. N02PC45022-46). Samples from the FCCC, HCI and CPIC were processed and distributed by the Coriell Institute for Medical Research. The content of this manuscript does not necessarily reflect the views or policies of the National Cancer Institute or any of the collaborating centres in the BCFR, nor does mention of trade names, commercial products or organizations imply endorsement by the US Government or the BCFR.

CNIO

The research leading to these results has been partially funded by Mutua Madrileña Foundation, "Red de Investigación en Cáncer RD06/0020/1160" and Spanish Ministry of Science and Innovation (FIS PI08 1120 and SAF2010-20493).

Copenhagen Breast Cancer Study (CBCS)

We would like to thank Bent Ejertsen for clinical data and acknowledge the NEYE foundation for financial support.

Deutsches Krebsforschungszentrum (DKFZ) study

The DKFZ study was supported by the DKFZ.

Epidemiological study of BRCA1 & BRCA2 mutation carriers (EMBRACE)

Douglas F. Easton is the PI of the study. EMBRACE Collaborating Centres are: Coordinating Centre, Cambridge: Susan Peock, Debra Frost, Steve D. Ellis, Elena Fineberg, Radka Platte, Clare Oliver. North of Scotland Regional Genetics Service, Aberdeen: Zosia Miedzzybrodzka, Helen Gregory. Northern Ireland Regional Genetics Service, Belfast: Patrick Morrison, Lisa Jeffers. West Midlands Regional Clinical Genetics Service, Birmingham: Trevor Cole, Kai-ren Ong, Jonathan Hoffman. South West Regional Genetics Service, Bristol: Alan Donaldson, Margaret James. East Anglian Regional Genetics Service, Cambridge: Joan Paterson, Sarah Downing, Amy Taylor. Medical Genetics Services for Wales, Cardiff: Alexandra Murray, Mark T. Rogers, Emma McCann. St James's Hospital, Dublin and National Centre for Medical Genetics, Dublin: M. John Kennedy, David Barton. South East of Scotland Regional Genetics Service, Edinburgh: Mary Porteous, Sarah Drummond. Peninsula Clinical Genetics Service, Exeter: Carole Brewer, Emma Kivuva, Anne Searle, Selina Goodman, Kathryn Hill. West of Scotland Regional Genetics Service, Glasgow: Rosemarie Davidson, Victoria Murday, Nicola Bradshaw, Lesley Snadden, Mark Longmuir, Catherine Watt, Sarah Gibson, Eshika Haque, Ed Tobias, Alexis Duncan. South East Thames Regional Genetics Service, Guy's Hospital London: Louise Izatt, Chris Jacobs, Caroline Langman. North West Thames Regional Genetics Service, Harrow: Huw Dorkins. Leicestershire Clinical Genetics Service, Leicester: Julian Barwell. Yorkshire Regional Genetics Service, Leeds: Julian Adlard, Gemma Serra-Feliu. Cheshire & Merseyside Clinical Genetics Service, Liverpool: Ian Ellis, Catherine Houghton. Manchester Regional Genetics Service, Manchester: D Gareth Evans, Fiona Laloo, Jane Taylor. North East Thames Regional Genetics Service, NE Thames, London: Lucy Side, Alison Male, Cheryl Berlin. Nottingham Centre for Medical Genetics, Nottingham: Jacqueline Eason, Rebecca Collier. Northern Clinical Genetics Service, Newcastle: Fiona Douglas, Oonagh Claber, Irene Jobson. Oxford Regional Genetics Service, Oxford: Lisa Walker, Diane McLeod, Dorothy Halliday, Sarah Durell, Barbara Stayner. The Institute of Cancer Research and Royal Marsden NHS Foundation Trust: Ros Eeles, Susan Shanley, Nazneen Rahman, Richard Houlston, Elizabeth Bancroft, Elizabeth Page, Audrey Arden-Jones, Kelly Kohut, Jennifer Wiggins, Elena Castro, Anita Mitra, Lisa Robertson. North Trent Clinical Genetics Service, Sheffield: Jackie Cook, Oliver Quarrell, Cathryn Bardsley. South West Thames Regional Genetics Service, London: Shirley Hodgson, Sheila Goff, Glen Brice, Lizzie Winchester, Charlotte Eddy, Vishakha Tripathi, Virginia Attard. Wessex Clinical Genetics Service, Princess Anne Hospital, Southampton: Diana Eccles, Anneke Lucassen, Gillian Crawford, Donna McBride, Sarah Smalley. EMBRACE is supported by Cancer Research UK Grants C1287/A10118 and C1287/A11990. D. Gareth Evans and Fiona Laloo are supported by an NIHR grant to the Biomedical Research Centre, Manchester. The Investigators at The Institute of Cancer Research and The Royal Marsden NHS Foundation Trust are supported by an NIHR grant to the Biomedical Research Centre at The Institute of Cancer Research and The Royal Marsden NHS Foundation Trust. Ros Eeles is supported by Cancer Research UK Grant C5047/A8385.

GEORGETOWN

CI received support from the Familial Cancer Registry and the Tissue Culture Shared Registry at Georgetown University (NIH/NCI grant P30-CA051008), the Cancer Genetics Network (HHSN261200744000C), and Swing Fore the Cure. Gynecologic Oncology Group (GOG)

This study was supported by National Cancer Institute grants to the Gynecologic Oncology Group (GOG) Administrative Office and the GOG Tissue Bank (CA 27469), and to the GOG Statistical and Data Center (CA 37517 and CA 101165). We thank the investigators of the Australia New Zealand Gynaecological Oncology Group (ANZGOG). GOG's participation was sponsored by GOG's Cancer Prevention and Control Committee, and supported through funding provided by both intramural (Clinical Genetics Branch, DCEG) and extramural (Community Oncology and Prevention Trials Program - COPTRG) NCI programs.

Hospital Clinico San Carlos (HCSC)

The HCSC study was partially supported by Instituto de Salud Carlos III; RD06/0020/0021. We wish to thank Dr. Pedro Perez-Segura and Dr. Atocha Romero for their contribution to this study.

The Hereditary Breast and Ovarian Cancer Research Group Netherlands (HEBON)

HEBON Collaborating Centres: Coordinating center: Netherlands Cancer Institute, Amsterdam, NL: F.B.L. Hogervorst, S. Verhoef, M. Verheus, L.J. van 't Veer, F.E. van Leeuwen, M.A. Rookus; Erasmus Medical Center, Rotterdam, NL: M. Collée, A.M.W. van den Ouweland, A. Jager, M.J. Hoening, M.M.A. Tilanus-Linthorst, C. Seynaeve; Leiden University Medical Center, NL, Leiden: C.J. van Asperen, J.T. Wijnen, M.P. Vreeswijk, R.A. Tollenaar, P. Devilee; Radboud University Nijmegen Medical Center, Nijmegen, NL: M.J. Ligtenberg, N. Hoogerbrugge; University Medical Center Utrecht, Utrecht, NL: M.G. Ausems, R.B. van der Luijt; Amsterdam Medical Center, NL: C.M. Aalfs, T.A. van Os; VU University Medical Center, Amsterdam, NL: J.J.P. Gille, Q. Waisfisz, H.E.J. Meijers-Heijboer; University Hospital Maastricht, Maastricht, NL: E.B. Gomez-Garcia, C.E. van Roozendaal, Marinus J. Blok, B. Caanen; University Medical Center Groningen University, NL: J.C. Oosterwijk, A.H. van der Hout, M.J. Mourits; The Netherlands Foundation for the detection of hereditary tumours, Leiden, NL: H.F. Vasen. The HEBON study is supported by the Dutch Cancer Society grants NK1998-1854, NK12004-3088, NK12007-3756 and the ZonMW grant 91109024.

Helsinki Breast Cancer Study (HEBCS)

HEBCS acknowledge Drs. Kristiina Aittomäki, Kirsimari Aaltonen and Carl Blomqvist and Tuomas Heikkinen and research nurse Irja Erkkilä for their help with the patient data and samples. The HEBCS study has been financially supported by the Helsinki University Central Hospital Research Fund, Academy of Finland (132473), the Finnish Cancer Society, and the Sigrid Juselius Foundation.

ICO

Contract grant sponsor: Asociación Española Contra el Cáncer, Spanish Health Research Fund; Carlos III Health Institute; Catalan Health Institute and Autonomous Government of Catalonia. Contract grant numbers: ISCIII/RETIC RD06/0020/1051, PI10/01422, PI10/31488 and 2009SGR290.

ILUH

The ILUH group was supported by the Icelandic Association "Walking for Breast Cancer Research" and by the Landspítali University Hospital Research Fund.

INHERIT

We would like to thank Stéphane Dubois, Dr Martine Dumont, Martine Tranchant (Cancer Genomics Laboratory, CRCHUQ) for sample management and skillful technical assistance, Sylvie Desjardins and Marc-André Rodrigue (Plateforme de séquençage et de génotypage des génome du CRCHUL/CHUQ) for iPLEX genotyping and Pascal Belleau for data quality control analyses.

Istituto Oncologico Veneto Hereditary Breast and Ovarian Cancer Study (IOVHBOCS)

This study was supported by Ministero dell'Istruzione, dell'Università e della Ricerca (MIUR) and "Ministero della Salute" ("Progetto Tumori Femminili and grant numbers RFPS 2006-5-341353, ACC2/R6.9")

kConFab

We wish to thank Heather Thorne, Eveline Niedermayr, all the kConFab research nurses and staff, the heads and staff of the Family Cancer Clinics, and the Clinical Follow Up Study (funded 2001-2009 by NHMRC and currently by the National Breast Cancer Foundation and Cancer Australia #628333) for their contributions to this resource, and the many families who contribute to kConFab. kConFab is supported by grants from the National Breast Cancer Foundation, the National Health and Medical Research Council (NHMRC) and by the Queensland Cancer Fund, the Cancer Councils of New South Wales, Victoria, Tasmania and South Australia, and the Cancer

Foundation of Western Australia. ABS is funded by an NHMRC Senior Research Fellowship.

Memorial Sloan-Kettering Cancer Center (MSKCC)

We acknowledge the Starr Cancer Consortium, the Breast Cancer Research Foundation, the Norman and Carol Stone Cancer Research Initiative, the Kate and Robert Niehaus Clinical Cancer Research Initiative, the Lymphoma Foundation, and the Sabin Family Research Initiative.

National Cancer Institute (NCI)

The research of Drs. Greene and Loud was supported by the Intramural Research Program of the US National Cancer Institute at the National Institutes of Health, and by support services contracts NO2-CP-11019-50 and NO2-CP-65504 with Westat, Inc, Rockville, MD.

N.N. Petrov Institute of Oncology (NNPIO)

This work has been supported by the Russian Federation for Basic Research (grants 10-04-92601, 10-04-92110, 11-04-00227) the Federal Agency for Science and Innovations (contract 16.512.11.2237) and through a Royal Society International Joint grant (JP090615).

The *Hong Kong Hereditary Breast Cancer Family Registry* thank Dr. Ellen Li Charitable Foundation for their support

Hungarian Breast and Ovarian Cancer Study (HUNBOCS)

The study was supported by Norwegian EEA Financial Mechanism (HU0115/NA/2008-3/ÖP-9) and by Hungarian Research Grant KTIÁ-OTKA (CK-80745).

Ohio State University Clinical Cancer Genetics (OSU-CCG)

Leigha Senter and Kevin Sweet were instrumental in accrual of study participants, ascertainment of medical records and database management. We thank the Human Genetics Sample Bank for preparation of samples. This study was supported by the Ohio State University Comprehensive Cancer Center.

Ontario Cancer Genetics Network Study (OCGN)

This work was supported by Cancer Care Ontario, the "CIHR Team in Familial Risks of Breast Cancer" program, and the US National Cancer Institute, National Institutes of Health under RFA # CA-06-503 and through cooperative agreements with members of the Breast Cancer Family Registry (BCFR) and Principal Investigators. The content of this manuscript does not necessarily reflect the views or policies of the National Cancer Institute or any of the collaborating centres in the BCFR, nor does mention of trade names, commercial products, or organizations imply endorsement by the US Government or the BCFR. We wish to thank Teresa Selander, Nayana Weerasooriya and members of the Ontario Cancer Genetics Network for their contributions to the study.

Beckman Research Institute of City of Hope (BRICOH)

The study was supported by the National Institutes of Health (R01 CA74415 to SLN) and the Morris and Horowitz Families Endowment.

SWE-BCRA

SWE-BCRA collaborators: Per Karlsson, Margareta Nordling, Annika Bergman and Zakaria Einbeigi, Gothenburg, Sahlgrenska University Hospital; Marie Stenmark-Askalm and Sigrun Liedgren, Linköping University Hospital; Åke Borg, Niklas Loman, Håkan Olsson, Maria Solter, Helena Jernström, Katja Harbst and Karin Henriksson, Lund University Hospital; Annika Lindblom, Brita Arver, Anna von Wachenfeldt, Annelie Liljegren, Gisela Barbany-Bustintza and Johanna Rantala, Stockholm, Karolinska University Hospital; Beatrice Melin, Henrik Grönberg, Eva-Lena Stattin and Monica Emanuelsson, Umeå University Hospital; Hans Ehrencrona, Richard Rosenquist and Niklas Dahl, Uppsala University Hospital.

U.K. and Gilda Radner Familial Ovarian Cancer Registry (UKGRFOCR)

UKFOCR was supported by a project grant from CRUK to Paul Pharoah. We thank Paul Pharoah, Susan Ramus, Carole Pye, Patricia Harrington and Eva Wozniak for their contributions towards the UKFOCR. We would like to acknowledge the Roswell Park Alliance Foundation for their continued support of the Gilda Radner Ovarian Family Cancer Registry. GRFOCR would like to acknowledge Kirsten Moysich and Lara Sucheston (Department of Cancer Prevention and Control).

University of Kansas Medical Center (KUMC)

We thank Ms. JoEllen Weaver for her help collecting patient data and samples. AKG was funded by U01CA69631, 5U01CA113916, and the Eileen Stein Jacoby Fund while at FCCC. The author acknowledges support from The University of Kansas Cancer Center and the Kansas Bioscience Authority Eminent Scholar Program. AKG is the Chancellors Distinguished Chair in Biomedical Sciences endowed Professor.

University of Pennsylvania (UPENN)

This research was supported by the Breast Cancer Research Foundation (to KLN) and the Komen Foundation for the Cure (to SMD).

Women's Cancer Program - Cedars-Sinai Medical Center (WCRI)

This work is supported by funding from the American Cancer Society Clinical Research Professorship (SIOP-06-258-COUN).

Author details

¹Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Worts Causeway, Cambridge CB1 8RN, UK

²Cancer Genomics Laboratory, Centre Hospitalier Universitaire de Québec, 2705

Laurier Boulevard, T3-57, Quebec City, QC Canada. ³Genetics and Population

Health Division, Queensland Institute of Medical Research, 300 Herston Rd,

Herston, Brisbane, QLD 4006, Australia. ⁴Unité Mixte de Génétique

Constitutionnelle des Cancers Fréquents, Centre Hospitalier Universitaire de Lyon/

Centre Léon Bérard, 28 rue Laënnec, Lyon 69373, France and INSERM U1052,

CNRS UMR5286, Université Lyon 1, Cancer Research Center of Lyon, 28 rue

Laënnec, Lyon 69373, France. ⁵Section of Genetic Oncology, Dept. of Laboratory

Medicine, University and University Hospital of Pisa, Via Roma 57, 56125 Pisa, Italy.

⁶Department of Oncology, Lund University Hospital, Lund, Sweden. ⁷Department of

Clinical Genetics, Karolinska University Hospital, Stockholm, Sweden.

⁸Department of Oncology, Karolinska University Hospital, Stockholm, Sweden.

⁹Department of Genetics and Pathology, Rudbeck Laboratory, Uppsala University,

Uppsala, Sweden. ¹⁰Department of Oncology, Sahlgrenska University Hospital,

Gothenburg, Sweden. ¹¹Abramson Cancer Center, Perelman School of Medicine

at the University of Pennsylvania, Philadelphia, PA, USA. ¹²Department of Genetics

and Pathology, Pomeranian Medical University, Szczecin, Poland. ¹³Department of

Genetics and Pathology, Pomeranian Medical University, Szczecin and

Postgraduate School of Molecular Medicine, Warsaw Medical University, Warsaw,

Poland. ¹⁴Human Genetics Group, Human Cancer Genetics Programme, Spanish

National Cancer Research Centre, Madrid, Spain and Spanish Network on Rare

Diseases (CIBERER). ¹⁵Institute of Biology and Molecular Genetics. Universidad de

Valladolid (IBGM-UVA), Valladolid, Spain. ¹⁶Oncology unit. Hospital clinico

Universitario "Lozano Blesa", Zaragoza, Spain. ¹⁷Human Genetics Group and

Genotyping Unit, Human Cancer Genetics Programme, Spanish National Cancer

Research Centre, Madrid, Spain and Spanish Network on Rare Diseases (CIBERER).

¹⁸Molecular Genetics of Breast Cancer, Deutsches Krebsforschungszentrum

(DKFZ), Heidelberg, Germany. ¹⁹Family Cancer Clinic, Netherlands Cancer Institute,

Amsterdam, The Netherlands. ²⁰Department of Clinical Genetics, Academic Meical

Center, Amsterdam, The Netherlands. ²¹Department of Clinical Genetics,

Netherlands Cancer Institute, Amsterdam, The Netherlands. ²²Department of

Clinical Genetics, VU Medical Center, Amsterdam, The Netherlands. ²³Department

of Clinical Genetics and GROM, School for Oncology and Developmental Biology,

MUMC, Maastricht, The Netherlands. ²⁴Department of Clinical Genetics and

GROM, School for Oncology and Developmental Biology, MUMC, Maastricht, The

Netherlands. ²⁵Department of Human Genetics, Radboud University Nijmegen

Medical Center, Nijmegen, The Netherlands. ²⁶Department of Clinical Genetics,

Family Cancer Clinic, Erasmus University Medical Center, Rotterdam, The

Netherlands. ²⁷Department of Clinical Genetics, Family Cancer Clinic, Erasmus

University Medical Center, Rotterdam, The Netherlands. ²⁸Department of Medical

Genetics, University Medical Center Utrecht, PO Box 85090, 3508 AB Utrecht, The

Netherlands. ²⁹Department of Genetics, University Medical Center, Groningen

University, Groningen, The Netherlands. ³⁰Netherlands Cancer Institute,

Amsterdam, The Netherlands. ³¹Genetic Medicine, Manchester Academic Health

Sciences Centre, Central Manchester University Hospitals NHS Foundation Trust,

Manchester, UK. ³²Clinical Genetics, Guy's and St. Thomas' NHS Foundation Trust,

London, UK. ³³Oncogenetics Team, The Institute of Cancer Research and Royal

Marsden NHS Foundation Trust, UK. ³⁴Yorkshire Regional Genetics Service, Leeds,

UK. ³⁵Ferguson-Smith Centre for Clinical Genetics, Yorkhill Hospitals, Glasgow, UK.

³⁶West Midlands Regional Genetics Service, Birmingham Women's Hospital

Healthcare NHS Trust, Edgbaston, Birmingham, UK. ³⁷Sheffield Clinical Genetics

Service, Sheffield Children's Hospital, Sheffield, UK. ³⁸Department of Clinical

Genetics, East Anglian Regional Genetics Service, Addenbrookes Hospital,

Cambridge, UK. ³⁹Institute of Genetic Medicine, Centre for Life, Newcastle Upon

Tyne Hospitals NHS Trust, Newcastle upon Tyne, UK. ⁴⁰Department of Clinical

Genetics, Royal Devon & Exeter Hospital, Exeter, UK. ⁴¹Medical Genetics Unit, St

George's, University of London, UK. ⁴²Northern Ireland Regional Genetics Centre,

Belfast Health and Social Care Trust, and Department of Medical Genetics,

Queens University Belfast, Belfast UK. ⁴³Oxford Regional Genetics Service, Churchill

Hospital, Oxford, UK. ⁴⁴All Wales Medical Genetics Services, University Hospital of

Wales, Cardiff, UK. ⁴⁵Clinical Genetics Department, St Michael's Hospital, Bristol, UK

⁴⁶North West Thames Regional Genetics Service, Kennedy-Galton Centre, Harrow,

UK. ⁴⁷Department of Pathology and Laboratory Medicine, University of Kansas Medical Center, Kansas City, KS, USA. ⁴⁸Clinical Molecular Genetics Laboratory, Fox Chase Cancer Center, Philadelphia, PA, USA. ⁴⁹Service de Génétique Oncologique, Institut Curie, Paris, France, Unité INSERM U830, Institut Curie, Paris, France, Université Paris Descartes, Faculté de Médecine, Paris, France. ⁵⁰Service de Génétique Oncologique, Institut Curie, Paris, France and Université Paris Descartes, Faculté de Pharmacie, Paris, France. ⁵¹Service de Génétique Oncologique, Institut Curie, 26 rue d'Ulm, Paris, France. ⁵²Service de Génétique Oncologique, Institut Curie, Paris, France. ⁵³INSERM U1052, CNRS UMR5286, Université Lyon 1, Centre de Recherche en Cancérologie de Lyon, Lyon, France. ⁵⁴Unité Mixte de Génétique Constitutionnelle des Cancers Fréquents, Hospices Civils de Lyon / Centre Léon Bérard, Lyon, France. ⁵⁵Service de Génétique, Institut de Cancérologie Gustave Roussy, Villejuif, France and INSERM U946, Fondation Jean Dausset, Paris, France. ⁵⁶Consultation de Génétique, Département de Médecine, Institut de Cancérologie Gustave Roussy, Villejuif, France. ⁵⁷Département Oncologie génétique, Prévention et Dépistage, INSERM CIC-P9502, Institut Paoli-Calmettes/Université d'Aix-Marseille II, Marseille, France. ⁵⁸Centre Antoine Lacassagne, Nice, France. ⁵⁹Service de Génétique Clinique Chromosomique et Moléculaire, Centre Hospitalier Universitaire de St Etienne, St Etienne, France. ⁶⁰Laboratoire de Génétique Chromosomique, Hôtel Dieu Centre Hospitalier, BP 1125 Chambéry, France. ⁶¹Service de Génétique, Centre Hospitalier Universitaire Bretonneau, Tours, France. ⁶²Cancer Genetics Network "Groupe Génétique et Cancer", Fédération Nationale des Centres de Lutte Contre le Cancer, France. ⁶³Huntsman Cancer Institute, 2000 Circle of Hope, Salt Lake City, UT 84112, USA. ⁶⁴Division of Population Science, Fox Chase Cancer Center, 333 Cottman Avenue, Philadelphia, PA 19111, USA. ⁶⁵Department of Cancer Biology, Dana-Farber Cancer Institute, and Department of Surgery, Harvard Medical School, 27 Drydock Avenue, Boston, MA 02210, USA. ⁶⁶Department of Epidemiology, Columbia University, New York, NY, USA. ⁶⁷Centre for Molecular, Environmental, Genetic and Analytic (MEGA) Epidemiology, Melbourne School of Population Health, Level 1, 723 Swanston Street, The University of Melbourne, Victoria 3010, Australia. ⁶⁸Department of Epidemiology, Cancer Prevention Institute of California, 2201 Walnut Avenue, Suite 300, Fremont, CA 94538, USA. ⁶⁹Genetic Epidemiology Laboratory, Department of Pathology, University of Melbourne, Australia. ⁷⁰Department of Dermatology, University of Utah School of Medicine, 30 North 1900 East, SOM 4B454, Salt Lake City, UT 84132, USA. ⁷¹Dept of OB/GYN and Comprehensive Cancer Center, Medical University of Vienna, Vienna, Austria. ⁷²Center for Genomic Medicine, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark. ⁷³Department of Pathology, Landspítali - University Hospital, Reykjavik Iceland and Faculty of Medicine, University of Iceland, Reykjavik, Iceland. ⁷⁴Epidemiology Research Program, American Cancer Society, Atlanta, GA, USA. ⁷⁵Department of Environmental Medicine, NYU Cancer Institute, New York University School of Medicine, New York, NY. ⁷⁶Clinical Cancer Genetics Laboratory, Memorial Sloan Kettering Cancer Center, New York, NY. ⁷⁷Statistical and Data Center, Roswell Park Cancer Institute, Buffalo, USA. ⁷⁸Australia New Zealand (ANZGOG), Westmead Hospital, Sydney, Australia. ⁷⁹Ohio State University, Columbus Cancer Council, OH, USA. ⁸⁰Evanston CCOOP - NorthShore University Health System; University of Chicago, USA. ⁸¹Southern Pines Women's Health Center, P.C., University of North Carolina at Chapel Hill, USA. ⁸²Sarasota Memorial Healthcare, Tufts Medical Center, USA. ⁸³Departments of Molecular Virology, Immunology & Medical Genetics and Internal Medicine, Comprehensive Cancer Center, The Ohio State University, Columbus, OH 43210, USA. ⁸⁴Immunology and Molecular Oncology Unit, Istituto Oncologico Veneto IOV - IRCCS, Padua, Italy. ⁸⁵U.O.C. di Oncologia, ULSS5 Ovest Vicentino, Italy. ⁸⁶Laboratory of Molecular Oncology, N.N. Petrov Institute of Oncology, St.-Petersburg, Russia. ⁸⁷Lombardi Comprehensive Cancer Center, Georgetown University, Washington DC, USA. ⁸⁸Latvian Biomedical Research and Study Centre, Latvia. ⁸⁹Genetic Counselling Unit, Hereditary Cancer Program, IDIBELL-Catalan Institute of Oncology. ⁹⁰Molecular Diagnostic Unit, Hereditary Cancer Program, IDIBELL-Catalan Institute of Oncology. ⁹¹Department of Preventive Medicine, Keck School of Medicine, University of Southern California, California, USA. ⁹²Department of Gynecologic Oncology, Roswell Park Cancer Institute, Buffalo, NY, USA. ⁹³Women's Cancer Program at the Samuel Oschin Comprehensive Cancer Institute at Cedars-Sinai Medical Center, Los Angeles, USA. ⁹⁴Department of Molecular Genetics, National Institute of Oncology, Budapest, Hungary. ⁹⁵Cancer Research Initiatives Foundation, Sime Darby Medical Centre, Malaysia and University Malaysia Cancer Research Institute, University Malaysia Medical Centre, Malaysia. ⁹⁶Jonsson Comprehensive Cancer Center at UCLA, Los Angeles, CA, USA. ⁹⁷UCSF Cancer Risk Program, University of California, San Francisco; UCSF Departments of Medicine, Epidemiology, and Biostatistics, USA. ⁹⁸Cancer Genetics Laboratory, Department of Genetics, University of Pretoria,

South Africa. ⁹⁹Oncogenetics Laboratory, Vall d'Hebron Institute of Oncology (VHIO), Vall d'Hebron University Hospital, Barcelona Spain. ¹⁰⁰The Hong Kong Hereditary Breast Cancer Family Registry; The University of Hong Kong; Cancer Genetics Center, Hong Kong Sanatorium and Hospital, Hong Kong. ¹⁰¹Centre of Familial Breast and Ovarian Cancer, Department of Gynaecology and Obstetrics and Centre for Integrated Oncology (CIO), University hospital of Cologne, Germany. ¹⁰²Institute for Medical Informatics, Statistics and Epidemiology, University of Leipzig, Germany. ¹⁰³Department of Gynaecology and Obstetrics, Division of Tumor Genetics, Klinikum rechts der Isar, Technical University Munich, Germany. ¹⁰⁴Department of Gynaecology and Obstetrics, Ludwig-Maximilian University Munich, Germany. ¹⁰⁵Department of Gynaecology and Obstetrics, University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Germany. ¹⁰⁶Institute of Human Genetics, University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Germany. ¹⁰⁷Department of Gynaecology and Obstetrics, University Hospital Düsseldorf, Heinrich-Heine University Düsseldorf, Germany. ¹⁰⁸Institute of Human Genetics, University of Münster, Münster, Germany. ¹⁰⁹Institute of Cell and Molecular Pathology, Hannover Medical School, Hannover, Germany. ¹¹⁰Institute of Human Genetics, Campus Virchow Klinikum, Charite Berlin, Germany. ¹¹¹Department of Gynaecology and Obstetrics, University Hospital Ulm, Germany. ¹¹²Centre of Familial Breast and Ovarian Cancer, Department of Medical Genetics, Institute of Human Genetics, University Würzburg, Germany. ¹¹³Institute of Human Genetics, Department of Human Genetics, University Hospital Heidelberg, Germany. ¹¹⁴Department of Gynaecology and Obstetrics, University Hospital Carl Gustav Carus, Technical University Dresden, Germany. ¹¹⁵Institute of Human Genetics, University Regensburg, Germany. ¹¹⁶Institute of Human Genetics, University Hospital Frankfurt a.M., Germany. ¹¹⁷Molecular Oncology Laboratory, Hospital Clinico San Carlos, Madrid, Spain. ¹¹⁸Department of Obstetrics and Gynecology, University of Helsinki and Helsinki University Central Hospital, Biomedicum Helsinki, P.O. BOX 700, 00029 HUS, Helsinki, Finland. ¹¹⁹Faculty of Medicine - Medicine and Medical Specialties, Université de Montréal. ¹Hemato-oncology service, Hôpital du Sacré-Coeur de Montréal, 5400 Gouin Blvd West Montreal, Quebec, Canada. ¹²⁰Peter MacCallum Cancer Center, Melbourne, Australia. ¹²¹Department of Population Sciences, Beckman Research Institute of City of Hope, Duarte, CA, USA. ¹²²Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN, USA. ¹²³Department of Medical Genetics, Mayo Clinic, Rochester, MN, USA. ¹²⁴Department of Health Sciences Research, Mayo Clinic, Rochester, MN, USA. ¹²⁵Unit of Molecular Bases of Genetic Risk and Genetic Testing, Department of Preventive and Predicted Medicine, Fondazione IRCCS Istituto Nazionale Tumori (INT), Milan, Italy and IFOM, Fondazione Istituto FIRC di Oncologia Molecolare, Milan, Italy. ¹²⁶Unit of Medical Genetics, Department of Preventive and Predictive Medicine, Fondazione IRCCS Istituto Nazionale Tumori (INT), Milan, Italy. ¹²⁷Division of Cancer Prevention and Genetics, Istituto Europeo di Oncologia (IEO), Milan Italy. ¹²⁸Department of Experimental Oncology, Istituto Europeo di Oncologia, Milan, Italy and Consortium for Genomics Technology (Cogentech), Milan, Italy. ¹²⁹Cancer Bioimmunotherapy Unit, Centro di Riferimento Oncologico, IRCCS, ¹Aviano (PN), Italy. ¹³⁰Medical Genetics Unit, Department of Clinical Physiopathology, University of Florence, Firenze, Italy. ¹³¹Department of Molecular Medicine, "Sapienza" University of Rome, Rome, Italy. ¹³²Clinical Genetics Branch, DCEG, NCI; Room EPS 7032, Rockville MD 20852 USA. ¹³³Samuel Lunenfeld Research Institute, Mount Sinai Hospital, Toronto, Ontario; Cancer Care Ontario, Departments of Molecular Genetics and Laboratory Medicine and Pathobiology, University of Toronto, Ontario, Canada. ¹³⁴Samuel Lunenfeld Research Institute, Mount Sinai Hospital, Toronto, Ontario, Department of Laboratory Medicine and Pathobiology, University of Toronto, Ontario, Canada. ¹³⁵Department of Laboratory Medicine and Pathobiology, University of Toronto, Toronto, ON, Canada; Department of Laboratory Medicine, and the Keenan Research Centre of the Li Ka Shing Knowledge Institute, St Michael's Hospital, Toronto, ON, Canada. ¹³⁶Ontario Cancer Genetics Network: Cancer Care Ontario, Canada. ¹³⁷Department of Clinical Genetics, Odense University Hospital, Denmark. ¹³⁸Department of Clinical Genetics, Rigshospitalet and Copenhagen University, Denmark. ¹³⁹Department of Clinical Genetics, Skejby Hospital, Aarhus, Denma. ¹⁴⁰Department of Clinical Genetics, Vejle Hospital, Denmark. ¹⁴¹Department of Laboratory Medicine and Pathology, and Health Sciences Research, Mayo Clinic, Rochester, MN, USA. ¹⁴²Cancer Genomics Laboratory, Centre Hospitalier Universitaire de Québec, 2705 Laurier Boulevard, T3-57, Quebec City and Canada Research Chair in Oncogenetics, Department of Molecular Medicine, Faculty of Medicine, Laval University, Quebec, Canada.

Authors' contributions

ACA, KBK and DFE wrote the manuscript. KBK performed the statistical analysis. ACA supervised the statistical analysis and data management. ACA, GCT and DFE developed the study design. LM and DB are the CIMBA database managers. AL wrote computer programs for the analysis. SH and OMS reviewed, recoded and classified the BRCA1 and BRCA2 mutations in CIMBA. GCT initiated and coordinates CIMBA. PS, JB, XC and YCD performed the genotyping. AJ, SLN, GCT and JS supervised the genotyping of samples. MAC, NL, KH, AL, BA, RR, PK, KN, SD, TR, AJ, JL, KJ, KD, EZ, AO, MD, RA, JB, UH, FBH, TAVO, SV, HEJMH, JW, EBGG, MJL, MK, JMC, MGEMA, JCO, SP, DF, SDE, RP, EF, DGE, FL, CJ, RE, JA, RD, TC, JC, JP, FD, CB, SH, PJM, LW, MTR, AD, HD, AKG, BB, DS, CH, BB, AdP, SM, AC, ML, BBdeP, OC, HS, MF, FP, SFF, IM, SB, MD, AM, MBT, JLH, EMJ, MS, DG, CFS, AFR, MKT, DGK, TvoH, FCN, RBB, MG, TK, VJ, ADC, KO, MP, JK, DC, JH, JB, JF, AET, MM, CO, EI, CI, LT, IB, CL, AT, JDV, SAG, KO, JG, BYK, EO, SHT, PAG, MSB, CMD, EJV, OD, AK, RKS, BW, CE, AM, ND, NA, SH, DN, SPA, DG, RVM, HD, AG, CS, KK, BF, DS, TC, MdIH, HN, TAM, BL, ABS, SLN, YCD, XW, ZF, VSP, NML, PR, MHG, JTL, ILA, HO, AMM, GG, MT, AMG, UBJ, ABS, TAK, GCT and FJC acquired phenotypic data and DNA samples or designed the centre-specific studies. All authors read and approved the final manuscript for publication.

Competing interests

The authors declare that they have no competing interests.

Received: 21 September 2011 Revised: 15 November 2011

Accepted: 20 February 2012 Published: 20 February 2012

References

- Antoniou A, Pharoah PD, Narod S, Risch HA, Eyfjord JE, Hopper JL, Loman N, Olsson H, Johannsson O, Borg A, Pasini B, Radice P, Manoukian S, Eccles DM, Tang N, Olah E, Anton-Culver H, Warner E, Lubinski J, Gronwald J, Gorski B, Tulinius H, Thorlacius S, Eerola H, Nevanlinna H, Syrjakoski K, Kallioniemi OP, Thompson D, Evans C, Peto J, *et al*: **Average risks of breast and ovarian cancer associated with BRCA1 or BRCA2 mutations detected in case Series unselected for family history: a combined analysis of 22 studies.** *Am J Hum Genet* 2003, **72**:1117-1130.
- Begg CB, Haile RW, Borg A, Malone KE, Concannon P, Thomas DC, Langholz B, Bernstein L, Olsen JH, Lynch CF, Anton-Culver H, Capanu M, Liang X, Hummer AJ, Sima C, Bernstein JL: **Variation of breast cancer risk among BRCA1/2 carriers.** *JAMA* 2008, **299**:194-201.
- Antoniou AC, Spurdle AB, Sinilnikova OM, Healey S, Pooley KA, Schmutzler RK, Versmold B, Engel C, Meindl A, Arnold N, Hofmann W, Sutter C, Niederacher D, Deissler H, Caldes T, Kampjarvi K, Nevanlinna H, Simard J, Beesley J, Chen X, Neuhausen SL, Rebbeck TR, Wagner T, Lynch HT, Isaacs C, Weitzel J, Ganz PA, Daly MB, Tomlinson G, Olopade OI, *et al*: **Common breast cancer-predisposition alleles are associated with breast cancer risk in BRCA1 and BRCA2 mutation carriers.** *Am J Hum Genet* 2008, **82**:937-948.
- Antoniou AC, Sinilnikova OM, McGuffog L, Healey S, Nevanlinna H, Heikkinen T, Simard J, Spurdle AB, Beesley J, Chen X, Neuhausen SL, Ding YC, Couch FJ, Wang X, Fredericksen Z, Peterlongo P, Peissel B, Bonanni B, Viel A, Bernard L, Radice P, Szabo CI, Foretova L, Zikan M, Claes K, Greene MH, Mai PL, Rennert G, Lejbkovicz F, Andrulis IL, *et al*: **Common variants in LSP1, 2q35 and 8q24 and breast cancer risk for BRCA1 and BRCA2 mutation carriers.** *Hum Mol Genet* 2009, **18**:4442-4456.
- Antoniou AC, Beesley J, McGuffog L, Sinilnikova OM, Healey S, Neuhausen SL, Ding YC, Rebbeck TR, Weitzel JN, Lynch HT, Isaacs C, Ganz PA, Tomlinson G, Olopade OI, Couch FJ, Wang X, Lindor NM, Pankratz VS, Radice P, Manoukian S, Peissel B, Zaffaroni D, Barile M, Viel A, Allavena A, Dall'olio V, Peterlongo P, Szabo CI, Zikan M, Claes K, *et al*: **Common breast cancer susceptibility alleles and the risk of breast cancer for BRCA1 and BRCA2 mutation carriers: implications for risk prediction.** *Cancer Res* 2010, **70**:9742-9754.
- Antoniou AC, Kartsonaki C, Sinilnikova OM, Soucy P, McGuffog L, Healey S, Lee A, Peterlongo P, Manoukian S, Peissel B, Zaffaroni D, Cattaneo E, Barile M, Pensotti V, Pasini B, Dolcetti R, Giannini G, Laura PA, Varesco L, Radice P, Mai PL, Greene MH, Andrulis IL, Glendon G, Ozelik H, Thomassen M, Gerdes AM, Kruse TA, Birk JU, Cruger DG, *et al*: **Common alleles at 6q25.1 and 1p11.2 are associated with breast cancer risk for BRCA1 and BRCA2 mutation carriers.** *Hum Mol Genet* 2011, **20**:3304-3321.
- Engel C, Versmold B, Wappenschmidt B, Simard J, Easton DF, Peock S, Cook M, Oliver C, Frost D, Mayes R, Evans DG, Eeles R, Paterson J, Brewer C, McGuffog L, Antoniou AC, Stoppa-Lyonnet D, Sinilnikova OM, Barjhoux L, Frenay M, Michel C, Leroux D, Dreyfus H, Toulas C, Gladieff L, Uhrhammer N, Bignon YJ, Meindl A, Arnold N, Varon-Mateeva R, *et al*: **Association of the variants CASP8 D302H and CASP10 V410I with breast and ovarian cancer risk in BRCA1 and BRCA2 mutation carriers.** *Cancer Epidemiol Biomarkers Prev* 2010, **19**:2859-2868.
- Ramus SJ, Kartsonaki C, Gayther SA, Pharoah PD, Sinilnikova OM, Beesley J, Chen X, McGuffog L, Healey S, Couch FJ, Wang X, Fredericksen Z, Peterlongo P, Manoukian S, Peissel B, Zaffaroni D, Roversi G, Barile M, Viel A, Allavena A, Ottini L, Papi L, Gismondi V, Capra F, Radice P, Greene MH, Mai PL, Andrulis IL, Glendon G, Ozelik H, *et al*: **Genetic variation at 9p22.2 and ovarian cancer risk for BRCA1 and BRCA2 mutation carriers.** *J Natl Cancer Inst* 2010, **103**:105-116.
- Antoniou AC, Wang X, Fredericksen ZS, McGuffog L, Tarrell R, Sinilnikova OM, Healey S, Morrison J, Kartsonaki C, Lesnick T, Ghousaini M, Barrowdale D, Peock S, Cook M, Oliver C, Frost D, Eccles D, Evans DG, Eeles R, Izatt L, Chu C, Douglas F, Paterson J, Stoppa-Lyonnet D, Houdayer C, Mazoyer S, Giraud S, Lasset C, Remenieras A, Caron O, *et al*: **A locus on 19p13 modifies risk of breast cancer in BRCA1 mutation carriers and is associated with hormone receptor-negative breast cancer in the general population.** *Nat Genet* 2010, **42**:885-892.
- Gaudet MM, Kirchoff T, Green T, Vijai J, Korn JM, Guiducci C, Segre AV, McGee K, McGuffog L, Kartsonaki C, Morrison J, Healey S, Sinilnikova OM, Stoppa-Lyonnet D, Mazoyer S, Gauthier-Villars M, Sobol H, Longy M, Frenay M, Gemo SC, Hogervorst FB, Rookus MA, Collee JM, Hoogerbrugge N, van Roozendaal KE, Piedmonte M, Rubinstein W, Nerenstone S, Van Le L, Blank SV, *et al*: **Common genetic variants and modification of penetrance of BRCA2-associated breast cancer.** *PLoS Genet* 2010, **6**:e1001183.
- Cox DG, Simard J, Sinnott D, Hamdi Y, Soucy P, Ouimet M, Barjhoux L, VERNY-Pierre C, McGuffog L, Healey S, Szabo C, Greene MH, Mai PL, Andrulis IL, Thomassen M, Gerdes AM, Caligo MA, Friedman E, Laitman Y, Kaufman B, Paluch SS, Borg A, Karlsson P, Askmalin MS, Bustinza GB, Nathanson K, Domchek SM, Rebbeck TR, Benitez J, Hamann U, *et al*: **Common variants of the BRCA1 wild-type allele modify the risk of breast cancer in BRCA1 mutation carriers.** *Hum Mol Genet* 2011, **20**:4732-4747.
- Broeks A, Schmidt MK, Sherman ME, Couch FJ, Hopper JL, Dite GS, Apicella C, Smith LD, Hammet F, Southey MC, van 't Veer LJ, de Groot R, Smit VT, Fasching PA, Beckmann MW, Jud S, Ekici AB, Hartmann A, Hein A, Schulz-Wendtland R, Burwinkel B, Marme F, Schneeweiss A, Sinn HP, Sohn C, Tchatchou S, Bojesen SE, Nordestgaard BG, Flyger H, Orsted DD, *et al*: **Low penetrance breast cancer susceptibility loci are associated with specific breast tumour subtypes: findings from the Breast Cancer Association Consortium.** *Hum Mol Genet* 2011, **20**:3289-3303.
- Garcia-Closas M, Hall P, Nevanlinna H, Pooley K, Morrison J, Richesson DA, Bojesen SE, Nordestgaard BG, Axelsson CK, Arias JI, Milne RL, Ribas G, Gonzalez-Neira A, Benitez J, Zamora P, Brauch H, Justenhoven C, Hamann U, Ko YD, Bruening T, Haas S, Dork T, Schurmann P, Hillemanns P, Bogdanova N, Bremer M, Karstens JH, Fagerholm R, Aaltonen K, Aittomaki K, *et al*: **Heterogeneity of breast cancer associations with five susceptibility loci by clinical and pathological characteristics.** *PLoS Genet* 2008, **4**: e1000054.
- Mulligan AM, Couch FJ, Barrowdale D, Domchek SM, Eccles D, Nevanlinna H, Ramus SJ, Robson M, Sherman M, Spurdle AB, Wappenschmidt B, Lee A, McGuffog L, Healey S, Sinilnikova OM, Janavicius R, Hansen TV, Nielsen FC, Ejlersen S, Osorio A, Munoz-Repeto I, Duran M, Godino J, Pertesi M, Benitez J, Peterlongo P, Manoukian S, Peissel B, Zaffaroni D, Cattaneo E, *et al*: **Common breast cancer susceptibility alleles are associated with tumour subtypes in BRCA1 and BRCA2 mutation carriers: results from the Consortium of Investigators of Modifiers of BRCA1/2.** *Breast Cancer Res* 2011, **13**:R110.
- Turnbull C, Ahmed S, Morrison J, Pernet D, Renwick A, Maranian M, Seal S, Ghousaini M, Hines S, Healey CS, Hughes D, Warren-Perry M, Tapper W, Eccles D, Evans DG, Hoening M, Schutte M, van den Ouweland A, Houlston R, Ross G, Langford C, Pharoah PD, Stratton MR, Dunning AM, Rahman N, Easton DF: **Genome-wide association study identifies five new breast cancer susceptibility loci.** *Nat Genet* 2010, **42**:504-507.

16. Ghossaini M, Fletcher O, Michailidou K, Turnbull C, Schmidt MK, Dicks E, Dennis J, Wang Q, Humphreys MK, Luccarini C, Baynes C, Conroy D, Maranian M, Ahmed S, Driver K, Johnson N, Orr N, dos Santos Silva I, Waisfisz Q, Meijers-Heijboer H, Uitterlinden AG, Rivadeneira F, Netherlands Collaborative Group on Hereditary Breast and Ovarian Cancer (HEBON), Hall P, Czene K, Irwanto A, Liu J, Nevanlinna H, Aittomäki K, Blomqvist C, Meindl A, *et al*: **Genome-wide association analysis identifies three new breast cancer susceptibility loci.** *Nat Genet* 2012.
17. Fletcher O, Johnson N, Orr N, Hosking FJ, Gibson LJ, Walker K, Zelenika D, Gut I, Heath S, Palles C, Coupland B, Broderick P, Schoemaker M, Jones M, Williamson J, Chilcott-Burns S, Tomczyk K, Simpson G, Jacobs KB, Chanock SJ, Hunter DJ, Tomlinson IP, Godwin A, Ashworth A, Ross G, dos Santos Silva I, Lathrop M, Houlston RS, Peto J: **Novel breast cancer susceptibility locus at 9q31.2: results of a genome-wide association study.** *J Natl Cancer Inst* 2011, **103**:425-435.
18. Antoniou AC, Sinilnikova OM, Simard J, Leone M, Dumont M, Neuhausen SL, Struewing JP, Stoppa-Lyonnet D, Barjhoux L, Hughes DJ, Coupier I, Belotti M, Lasset C, Bonadona V, Bignon YJ, Rebbeck TR, Wagner T, Lynch HT, Domchek SM, Nathanson KL, Garber JE, Weitzel J, Narod SA, Tomlinson G, Olopade OI, Godwin A, Isaacs C, Jakubowska A, Lubinski J, Gronwald J, *et al*: **RAD51 135G > C modifies breast cancer risk among BRCA2 mutation carriers: results from a combined analysis of 19 studies.** *Am J Hum Genet* 2007, **81**:1186-1200.
19. Chenevix-Trench G, Milne RL, Antoniou AC, Couch FJ, Easton DF, Goldgar DE: **An international initiative to identify genetic modifiers of cancer risk in BRCA1 and BRCA2 mutation carriers: the Consortium of Investigators of Modifiers of BRCA1 and BRCA2 (CIMBA).** *Breast Cancer Res* 2007, **9**:104.
20. Lange K, Weeks D, Boehnke M: **Programs for pedigree analysis: MENDEL, FISHER, and dGENE.** *Genet Epidemiol* 1988, **5**:471-472.
21. Barnes D, Lee A, EMBRACE Investigators, kConFab Investigators, Easton D, Antoniou AC: **Evaluation of association methods for analysing modifiers of disease risk in carriers of high-risk mutations.** *Genet Epidemiol* 2011.
22. Rennert G, Bisland-Naggan S, Barnett-Griness O, Bar-Joseph N, Zhang S, Rennert HS, Narod SA: **Clinical outcomes of breast cancer in carriers of BRCA1 and BRCA2 mutations.** *N Engl J Med* 2007, **357**:115-123.
23. Buisson M, Anczukow O, Zetoune AB, Ware MD, Mazoyer S: **The 185delAG mutation (c.68_69delAG) in the BRCA1 gene triggers translation reinitiation at a downstream AUG codon.** *Hum Mutat* 2006, **27**:1024-1029.
24. Mazoyer S, Puget N, Perrin-Vidoz L, Lynch HT, Serova-Sinilnikova OM, Lenoir GM: **A BRCA1 nonsense mutation causes exon skipping.** *Am J Hum Genet* 1998, **62**:713-715.
25. Perrin-Vidoz L, Sinilnikova OM, Stoppa-Lyonnet D, Lenoir GM, Mazoyer S: **The nonsense-mediated mRNA decay pathway triggers degradation of most BRCA1 mRNAs bearing premature termination codons.** *Hum Mol Genet* 2002, **11**:2805-2814.
26. Anczukow O, Ware MD, Buisson M, Zetoune AB, Stoppa-Lyonnet D, Sinilnikova OM, Mazoyer S: **Does the nonsense-mediated mRNA decay mechanism prevent the synthesis of truncated BRCA1, CHK2, and p53 proteins?** *Hum Mutat* 2008, **29**:65-73.
27. Antoniou AC, Cunningham AP, Peto J, Evans DG, Lalloo F, Narod SA, Risch HA, Eyfjord JE, Hopper JL, Southey MC, Olsson H, Johannsson O, Borg A, Passini B, Radice P, Manoukian S, Eccles DM, Tang N, Olah E, Anton-Culver H, Warner E, Lubinski J, Gronwald J, Gorski B, Tryggvadottir L, Syrjakoski K, Kallioniemi OP, Eerola H, Nevanlinna H, Pharoah PD, *et al*: **The BOADICEA model of genetic susceptibility to breast and ovarian cancers: updates and extensions.** *Br J Cancer* 2008, **98**:1457-1466.
28. Boos DD: **On generalised score tests.** *Am Stat* 1992, **46**:327-333.
29. Antoniou AC, Chenevix-Trench G: **Common genetic variants and cancer risk in Mendelian cancer syndromes.** *Curr Opin Genet Dev* 2010, **20**:299-307.
30. Stevens KN, Vachon CM, Lee AM, Slager S, Lesnick T, Olsowd C, Fasching PA, Miron P, Eccles D, Carpenter JE, Godwin AK, Ambrosone C, Winqvist R, Brauch H, Schmidt MK, Cox A, Cross SS, Sawyer E, Hartmann A, Beckmann MW, Schulz-Wendtland R, Ekici AB, Tapper WJ, Gerty SM, Durcan L, Graham N, Hein R, Nickels S, Flesch-Janys D, Heinz J, *et al*: **Common breast cancer susceptibility loci are associated with triple negative breast cancer.** *Cancer Res* 2011, **71**:6240-6249.
31. Suva LJ, Winslow GA, Wattenhall RE, Hammonds RG, Moseley JM, Diefenbach-Jagger H, Rodda CP, Kemp BE, Rodriguez H, Chen EY, *et al*: **A parathyroid hormone-related protein implicated in malignant hypercalcemia: cloning and expression.** *Science* 1987, **237**:893-896.
32. Asadi F, Kukreja S: **Parathyroid hormone-related protein in prostate cancer.** *Crit Rev Eukaryot Gene Expr* 2005, **15**:15-28.
33. Linforth R, Anderson N, Hoey R, Nolan T, Downey S, Brady G, Ashcroft L, Bundred N: **Coexpression of parathyroid hormone related protein and its receptor in early breast cancer predicts poor patient survival.** *Clin Cancer Res* 2002, **8**:3172-3177.
34. Lindstrom S, Vachon CM, Li J, Varghese J, Thompson D, Warren R, Brown J, Leyland J, Audley T, Wareham NJ, Loos RJ, Paterson AD, Rommens J, Waggott D, Martin LJ, Scott CG, Pankratz VS, Hankinson SE, Hazra A, Hunter DJ, Hopper JL, Southey MC, Chanock SJ, Silva IS, Liu J, Eriksson L, Couch FJ, Stone J, Apicella C, Czene K, *et al*: **Common variants in ZNF365 are associated with both mammographic density and breast cancer risk.** *Nat Genet* 2011, **43**:185-187.
35. Mitchell G, Antoniou AC, Warren R, Peock S, Brown J, Davies R, Mattison J, Cook M, Warsi I, Evans DG, Eccles D, Douglas F, Paterson J, Hodgson S, Izatt L, Cole T, Burgess L, Eeles R, Easton DF: **Mammographic density and breast cancer risk in BRCA1 and BRCA2 mutation carriers.** *Cancer Res* 2006, **66**:1866-1872.
36. Bolton KL, Tyrer J, Song H, Ramus SJ, Notaridou M, Jones C, Sher T, Gentry-Maharaj A, Wozniak E, Tsai YY, Weidhaas J, Paik D, Van Den Berg DJ, Stram DO, Pearce CL, Wu AH, Brewster W, Anton-Culver H, Ziogas A, Narod SA, Levine DA, Kaye SB, Brown R, Paul J, Flanagan J, Sieh W, McGuire V, Whittemore AS, Campbell I, Gore ME, *et al*: **Common variants at 19p13 are associated with susceptibility to ovarian cancer.** *Nat Genet* 2010, **42**:880-884.
37. Goode EL, Chenevix-Trench G, Song H, Ramus SJ, Notaridou M, Lawrenson K, Widschwendter M, Vierkant RA, Larson MC, Kjaer SK, Birrer MJ, Berchuck A, Schildkraut J, Tomlinson I, Kiemeny LA, Cook LS, Gronwald J, Garcia-Closas M, Gore ME, Campbell I, Whittemore AS, Sutphen R, Phelan C, Anton-Culver H, Pearce CL, Lambrechts D, Rossing MA, Chang-Claude J, Moysich KB, Goodman MT, *et al*: **A genome-wide association study identifies susceptibility loci for ovarian cancer at 2q31 and 8q24.** *Nat Genet* 2010, **42**:874-879.
38. Song H, Ramus SJ, Tyrer J, Bolton KL, Gentry-Maharaj A, Wozniak E, Anton-Culver H, Chang-Claude J, Cramer DW, Diciocco R, Dork T, Goode EL, Goodman MT, Schildkraut JM, Sellers T, Baglietto L, Beckmann MW, Beesley J, Blaakaer J, Carney ME, Chanock S, Chen Z, Cunningham JM, Dicks E, Doherty JA, Durst M, Ekici AB, Fenstermacher D, Fridley BL, Giles G, *et al*: **A genome-wide association study identifies a new ovarian cancer susceptibility locus on 9p22.2.** *Nat Genet* 2009, **41**:996-1000.
39. Milne RL, Antoniou AC: **Genetic modifiers of cancer risk for BRCA1 and BRCA2 mutation carriers.** *Ann Oncol* 2011, **22**(Suppl 1):i11-i17.

doi:10.1186/bcr3121

Cite this article as: Antoniou *et al*: Common variants at 12p11, 12q24, 9p21, 9q31.2 and in ZNF365 are associated with breast cancer risk for BRCA1 and/or BRCA2 mutation carriers. *Breast Cancer Research* 2012 **14**:R33.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

