

### **sPDZD2: A Novel Negative Modulator of Hedgehog Signaling**

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PDZD2 is a multi-PDZ domain-containing protein of unknown function in early development. It is proteolytically cleaved to generate its secreted form, sPDZD2. Human PDZD2 is mapped to chromosome 5p13.2, which co-localizes with the disease-associated gene in a family of Brachydactyly Type A1 (BDA1) patients, suggesting involvement of PDZD2 in limb development. Hedgehog (Hh) is an important morphogen that dictates tissue patterning during embryonic development and recent studies showed that mutations in Indian Hedgehog (IHH) resulted in BDA1. Interestingly, *in situ* hybridization revealed that *Pdzd2* was expressed in the distal mesenchyme partially overlapping with *Shh* in mouse limb bud. During digit patterning, *Pdzd2* was expressed in the interzone that flanked the *Ihh*/*Gli1*-expressing phalanx condensation. Moreover, *Pdzd2* was expressed in the paraxial mesoderm adjacent to the differentiating neural tube. It is worth noting that PDZD2 protein was detected at the neural tube away from its site of synthesis, indicating a non-cell autonomous role of PDZD2 possibly via sPDZD2. *Pdzd2* expression in various Hh-active tissues in mouse and chicken suggested an evolutionary conserved role of *Pdzd2* in modulating general Hh signaling during early development.

Functional studies showed that overexpression of sPDZD2 in the chicken neural tube leads to down-regulation of *NKX2.2* and *OLIG2* expression. sPDZD2 was shown to counteract the ectopic *NKX2.2* expression induced by long-range signaling of ectopic HH. Consistently, sPDZD2 exhibited an inhibitory effect on SHH-induced reporter activity in a *Gli*-luciferase cell line. Taken together, our results provided the first evidence that sPDZD2 is a negative modulator of Hedgehog signaling.

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### **Sufu and Gli3 repressor mediate the temporal basal-to-apical progression of hair cell differentiation in mammalian cochleae**

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The Sonic Hedgehog pathway plays important roles in mammalian inner ear development. Mutations of *Shh*, *Smo* and *Gli3* lead to severe defects in mouse inner ear morphogenesis. However, knockout of *Gli2* does not affect inner ear morphology or cochlear hair cell differentiation, suggesting that the *Gli* repressor function may be required for Hedgehog signaling during inner ear development. *Sufu* is a negative regulator of Hedgehog signaling and it functions to repress *Gli* activator and enhance *Gli* repressor activities. To evaluate the involvement of *Sufu* and *Gli* transcription factors in mediating cochlear hair cell differentiation, we have analyzed the *Pax2*Cre;*Sufu*flox/flox, *Gli3*P1-4/P1-4 and *Gli3*Δ699/Δ699 mutants using hair cell marker *Myosin7a* and supporting cell markers *Sox2*, *P75* and *Jag1*. At E16.5, only one row of inner hair cells could be observed at the basal region of cochleae in the *Pax2*Cre;*Sufu*flox/flox mutants. Nevertheless, normal hair cells appeared at the medial region at E18.5, indicating that deletion of *Sufu* delays cochlear hair cell differentiation. *Gli3* repressor is abolished in the *Gli3*P1-4/P1-4 mutant, in which cochlear hair cell differentiation was delayed. Interestingly, in the *Gli3*Δ699/Δ699 mutant with excessive *Gli3* repressor, hair cell differentiation was accelerated in the apical region of the cochlear duct. Our results suggest that *Sufu* and *Gli3* repressor are essential factors which regulate the temporal basal-to-apical progression of cochlear hair cell differentiation, supporting that Sonic Hedgehog signaling is required to control the dynamics of hair cell differentiation.