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Rescue of perfluorooctanesulfonate (PFOS)-mediated Sertoli cell injury by overexpression of gap junction protein connexin 43

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Perfluorooctanesulfonate (PFOS) is an environmental toxicant used in developing countries, including China, as a stain repellent for clothing, carpets and draperies, but it has been banned in the U.S. and Canada since the late 2000s. PFOS perturbed the Sertoli cell tight junction (TJ)-permeability barrier, causing disruption of actin microfilaments in cell cytosol, perturbing the localization of cell junction proteins (e.g., occluden-ZO-1, N-cadherin- β -catenin). These changes destabilized Sertoli cell blood-testis barrier (BTB) integrity. These findings suggest that human exposure to PFOS might induce BTB dysfunction and infertility. Interestingly, PFOS-induced Sertoli cell injury associated with a down-regulation of the gap junction (GJ) protein connexin43 (Cx43). We next investigated if overexpression of Cx43 in Sertoli cells could rescue the PFOS-induced cell injury. Indeed, overexpression of Cx43 in Sertoli cells with an established TJ-barrier blocked the disruption in PFOS-induced GJ-intercellular communication, resulting in the re-organization of actin microfilaments, which rendered them similar to those in control cells. Furthermore, cell adhesion proteins that utilized F-actin for attachment became properly distributed at the cell-cell interface, resealing the disrupted TJ-barrier. In summary, Cx43 is a good target that might be used to manage PFOS-induced reproductive dysfunction.

The physiological significance of gap junctions (GJs) to support various cellular functions in mammalian cells and tissues has been well established (for reviews, see refs 1 and 2) including the testis (for reviews, see refs 3–6). While the testis is known to express more than ten different connexins for the construction of GJ-based intercellular communication channels^{7,8}, studies have shown that connexin 43 (Cx43)-based gap junctions (GJ)⁹ play important and unique physiological functions (for a review, see ref. 10), at least in Sertoli cells, which apparently cannot be superseded by other connexins. This notion is supported by studies in which Sertoli cell-specific deletion of Cx43 resulted in infertility in mice wherein spermatogonia failed to differentiate into spermatocytes and enter meiosis I/II¹¹. Sertoli cells also fail to become differentiated in the testis during adulthood¹¹, and the testis of these mice display defects in the expression of multiple genes based on gene profiling search¹², as well as mis-localization of proteins at the blood-testis barrier (BTB)¹³. Interestingly, fertility is maintained in mice following specific deletion of Cx43 in germ cells¹⁴, illustrating Cx26 and Cx45 expressed in these mice can supersede the lost function of Cx43 in germ cells¹⁴, in contrast to Cx43-specific KO in Sertoli cells which leads to infertility¹¹. Taken collectively, these studies using genetic models clearly illustrate the unique importance of Sertoli cell Cx43 in spermatogenesis (for a review, see ref. 3).

Other studies have also illustrated the significance of Cx43 in the maintenance of BTB function in the rat testis¹⁵, such as BTB homeostasis in particular reassembly of the Sertoli cell tight junction (TJ)-permeability barrier¹⁶. For instance, cell junctions at the BTB undergo continuous remodeling to support the transport of preleptotene spermatocytes connected in clones across the immunological barrier at stage VIII of the epithelial cycle, cell junction dynamics (disassembly, reassembly, stabilization) must be tightly coordinated and regulated through

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GJs. In a recent study using an animal model in which rats were exposed to an acute dose of adjuvins (1-(2,4-dichlorobenzyl)-1H-indazole-3-carbohydrazide, a potential male contraceptive under development (for reviews, see refs 17 and 18)) to induce irreversible BTB disruption and sterility due to meiotic arrest¹⁹. Overexpression of Cx43 was shown to be able to re-initiate spermatogenesis by re-booting meiosis I/II in adjuvins treated rats. For instance, round spermatids were detected in considerable number of tubules vs. adjuvins treated rats without Cx43 overexpression in the testis²⁰. Detailed analysis of tubules that displayed signs of meiosis in these adjuvins treated rats has shown that besides corrective spatiotemporal expression of Cx43 in the seminiferous epithelium similar to normal rat testes, F-actin organization was re-built through proper spatiotemporal expression of actin nucleation protein formin 1 and actin barbed end capping and bundling protein Eps8²⁰. These changes thus supported proper localization of TJ- (e.g., occludin, ZO-1) and basal ES (ectoplasmic specialization, a testis-specific anchoring junction (for reviews, see refs 21–23))- (e.g., N-cadherin, γ -catenin) proteins. These findings are significant because they illustrate that the Cx43-based GJ communication is crucial to multiple cellular events to maintain the homeostasis of the BTB, confirming findings of an earlier report regarding the likely involvement of Cx43 in providing cross-talk between various junctions at the BTB to support spermatogenesis¹⁶.

PFOS (perfluorooctanesulfonate) is an environmental toxicant with its use in consumer products (e.g., carpets, textiles, paints, leather and paper) being banned in the U.S. and Canada in the late 2000s due to its health risks. It continues to be widely used in China due to its ability to serve as a stain repellent, and thus is widely accepted by consumers (for a review, see ref. 24). It was shown that PFOS exerted its effects in Sertoli cells by perturbing F-actin organization in Sertoli cells, thereby disrupting localization of TJ- and basal ES-proteins at the Sertoli cell-cell interface, and associated with a disruption of the GJ-communication function based on a functional dye-transfer FRAP (fluorescence recovery after photobleaching) assay²⁵. These phenotypes are somewhat similar to adjuvins-induced aspermatogenesis in the testis *in vivo*^{19,20}. We thus sought to examine if overexpression of Cx43 could rescue the PFOS-mediated Sertoli cell injury, because if it could, these findings offer the basis for therapeutic management of PFOS-induced testis injury through gene therapy by overexpressing Cx43, such as the use of nanoparticles that contain mammalian expression vector with the full-length Cx43 cDNA^{26–28}. Herein, we report findings based on the use of an *in vitro* model, which are the basis of future *in vivo* studies.

Materials and Methods

Animals and Antibodies. Sprague-Dawley male pups at 15–18 days of age were purchased from Charles River Laboratories (Kingston, NY) and maintained at the Rockefeller University Comparative Bioscience Center (CBC). Each group of ten pups was supplied with a foster mother during shipment and housing at the Rockefeller University CBC, and they had free access to water and standard rat chow. The use of rats was approved by the Rockefeller University Laboratory Animal Care and Use Committee with Protocol Numbers: 12506 and 15780-H. Rats were euthanized by CO₂ asphyxiation using slow (20–30%/min) displacement of chamber air with compressed CO₂ using a regulator approved by the Rockefeller University Laboratory Safety & Environmental Health (LS&EH). All experimental protocols and methods including the use of animals for all the studies reported herein were approved by, and carried out in accordance with the relevant guidelines of the Rockefeller University LS&EH, the Rockefeller University Institutional Biosafety Committee (IBC), and the Rockefeller University Comparative Bioscience Center (CBC), which were detailed in the approved Protocols 12506 and 15780-H. Also, these procedures were also described in detail wherever applicable in the sections below. Antibodies were purchased and applied as listed in Table 1.

Toxicants. Perfluorooctanesulfonate (PFOS, heptadecafluorooctanesulfonic acid, potassium salt, Mr 538.22) was purchased from Sigma-Aldrich (St. Louis, MO) and dissolved in DMSO at 100 mM so that when the Sertoli cells were treated with 20 μ M PFOS, the concentration of DMSO in the culture medium was 0.02% (v/v). The same amount of DMSO was used in controls, including Sertoli cells transfected with the empty pCI-neo mammalian expression vector (Promega).

Isolation of Sertoli cells and Treatment with PFOS. Sertoli cells were isolated from 20-day-old rat testes and cultured in serum-free chemically defined medium of F12/DMEM supplemented with growth factors, including bovine insulin (10 μ g/ml), human transferrin (5 μ g/ml), and epidermal growth factor (2.5 ng/ml), as well as bacitracin (5 μ g/ml) and gentamicin (20 μ g/ml) as earlier described²⁹. Sertoli cells from 20-day-old rat testes were used because Sertoli cells at this age are fully differentiated and ceased to divide^{30,31}, and these cells are also functionally similar to Sertoli cells isolated from adult rat testes^{32,33}. Furthermore, Sertoli cells isolated from 20-day-old rat testes were >98% pure with negligible contamination of germ, peritubular myoid or Leydig cells³⁴. Virtually all contaminating germ cells were lysed via a brief hypotonic treatment using 20 mM Tris, pH 7.4 at 22 °C for 2.5 min at room temperature as described³⁵. However, Sertoli cells isolated from adult rat testes using the established BSA gradient approach³⁶ had a purity of ~85% even after the hypotonic treatment^{32,33}. As such, most investigators use Sertoli cells isolated from 20-day-old rat testes for their studies^{37–42}. For various experiments, Sertoli cells were seeded on Matrigel (BD Biosciences, San Jose, CA; 1:7 diluted in F12/DMEM medium) coated: (i) 6-well dishes at 0.4×10^6 cells/cm², with each well containing 5 ml of F12/DMEM supplemented with growth factors (for immunoblotting); and (ii) cover glasses (round, 18-mm diameter) placed into 12-well dishes at 0.04×10^6 cells/cm² (for immunofluorescent (IF) analysis of F-actin, actin binding/regulatory proteins such as Arp3 and Eps8), 0.15 vs. 0.1×10^6 cells/cm² (for IF analysis of Cx43 vs. TJ- and basal ES-proteins), with each well containing 2 ml medium. These cell densities were selected based on pilot experiments to better visualize the corresponding target proteins either in the Sertoli cell cytosol or at the cell-cell interface. For instance, a Sertoli cell density of 0.15×10^6 cells/cm² was necessary to obtain optimal visualization of Cx43. Cover glasses were then removed from the 12-well dishes and processed for immunofluorescence microscopy. To avoid inter-experimental variations, all cover glasses within an experiment were processed simultaneously. For

Antibodies	Host species	Vendor	Catalog number	Dilution	
				IB	IF
Actin	Goat	Santa Cruz Biotechnology	sc-1616	1:200	
Arp3	Mouse	Sigma-Aldrich	A5979	1:3000	1:100
α -catenin	Rabbit	Santa Cruz Biotechnology	sc-7894	1:200	
β -catenin	Mouse	Invitrogen	13-8400	1:500	1:100
β 1-integrin	Rabbit	Santa Cruz Biotechnology	sc-8978	1:200	
Claudin 11	Rabbit	Invitrogen	36-4500	1:500	1:100
Cx43	Rabbit	Sigma-Aldrich	C6219	1:3000	1:100
Eps8	Mouse	BD Biosciences	610143	1:3000	1:100
N-cadherin	Mouse	Invitrogen	33-3900	1:500	1:100
N-WASP	Rabbit	Santa Cruz Biotechnology	sc-20770	1:200	
Occludin	Rabbit	Invitrogen	71-1500	1:250	
Vimentin	Mouse	Santa Cruz Biotechnology	sc-6260	1:1000	
ZO-1	Rabbit	Invitrogen	61-7300	1:500	1:100

Table 1. Antibodies used for different experiments in this report.

experiments to assess the Sertoli cell TJ-permeability function to monitor BTB integrity *in vitro*, Sertoli cells at 1.0×10^6 cells/cm² were seeded onto Matrigel-coated (1:5 diluted in F12/DMEM) Millicell HA bicameral units (12 mm diameter, 0.6 cm² effective surface area; 0.45 μ m pore size; EMD Millipore), and units were then placed in 24-well dishes with the apical and basal compartment containing 0.5-ml F12/DMEM supplemented with growth factors as described²⁹. Each time point had quadruple bicameral units. The time of cell plating onto the bicameral units or cover glasses was defined as day 0. On day 2, cells were subjected to a hypotonic treatment with 20 mM Tris (pH 7.4 at 22 °C) for 2 min to lyse residual germ cells as described³⁵. Thereafter cells were washed twice with F12/DMEM, and the purity of these Sertoli cells was >98% when primer pairs specific to markers of Sertoli, Leydig, germ or myoid cells were used for RT-PCR, which confirmed negligible cell contamination as described⁴³. On day 3, PFOS dissolved in DMSO was diluted in F12/DMEM supplemented with various growth factors and gentamicin as described²⁹ to obtain the desired final concentration of 20 μ M. Sertoli cells were exposed to PFOS for 24 hr before termination. For the control, vehicle (i.e., F12/DMEM containing 0.02% DMSO (v/v)) was used.

Transfection of Sertoli cells with Plasmid DNA for Cx43 Overexpression. The full length rat testicular Cx43 cDNA (GenBank Accession Number BC081842.1) obtained by PCR using Sertoli cell cDNAs with both start and stop codons was cloned into the pCI-neo mammalian expression vector (Promega, Madison, WI) as earlier described²⁰. Possible endotoxin contamination in the plasmid DNA was removed using the EndoFree Plasmid Mega Kit (Qiagen, Germantown, MD). The sequence of this Cx43 cDNA clone was confirmed by direct nucleotide sequencing analysis at Genewiz (South Plainfield, NJ). Sertoli cells were transfected with pCI-neo empty vector (Ctrl) vs. pCI-neo containing Cx43 plasmid DNA (pCI-neo/Cx43) by using Effectene Transfection Reagent (Qiagen) for 6 hr on day 2. For cultures to be used for immunoblot analysis, Sertoli cells were transfected with 1.6 μ g plasmid DNA in 200 μ l Buffer EC, 12.8 μ l Enhancer, 24 μ l Effectene and 2 ml F12/DMEM with supplements and antibiotics. For cell staining and TER measurement, cells were transfected with 0.2 μ g plasmid DNA in 25 μ l Buffer EC, 1.6 μ l Enhancer, 3 μ l Effectene and 0.5 ml F12/DMEM with supplements and antibiotics. To monitor successful transfection, plasmid DNA was labeled with Cy3 (red fluorescence) using a Label IT Tracker Intracellular Nucleic Acid Localization Kit (Mirus).

Functional Assay that Monitors Sertoli cell TJ-Permeability Barrier Function. The Sertoli cell TJ-permeability barrier function *in vitro* was monitored by quantifying the transepithelial electrical resistance (TER) across the cell epithelium with a Millicell ERS system (EMD Millipore, Billerica, MA) as described^{29,44}. In brief, Sertoli cells were seeded on Matrigel (1:5 diluted with F12/DMEM)-coated Millicell HA bicameral units at 1.0×10^6 cells/cm² and cultured for 2 days to allow the assembly of functional TJ-barrier. Sertoli cells were then transfected with plasmid DNA on day 2 for 6 hr. Thereafter, cells were washed twice with F12/DMEM to remove transfection reagents, and they were cultured for an additional 18 hr before cells were exposed to PFOS for 24 hr. TER reading was taken daily using quadruple bicameral units for each treatment group including controls, medium was replaced immediately after TER reading was taken. Each TER experiment was performed three independent times using different batches of Sertoli cells, which yielded similar results.

Immunoblotting and Immunofluorescent Microscopy (IF). Sertoli cell lysates were obtained by using immunoprecipitation (IP) lysis buffer [10 mM Tris, pH 7.4 at 22 °C, containing 0.15 M NaCl, 1% NP-40 (v/v), and 10% glycerol (v/v)] freshly supplemented with protease and phosphatase inhibitor cocktails (Sigma-Aldrich) as described⁴⁴. Following homogenization using a Cole-Parmer Ultrasonic Processor, cell lysates were centrifuged at 15,000 g for 60 min at 4 °C to remove cellular debris. About 20 μ g cell lysate protein was subjected to SDS-PAGE and immunoblotting^{44,45} to detect changes in the steady-state levels of selected target proteins, including Cx43 after treatment of PFOS with or without overexpression of Cx43 vs. controls. About 10 μ g protein of cell lysate was used per lane to visualize β -actin and vimentin, which also served as the protein loading control. IF using cultured Sertoli cells was performed as described⁴⁴. Primary antibodies used for different experiments in this report are listed in Table 1, with goat anti-mouse or goat anti-rabbit IgG conjugated to Alexa Fluor 488 (Invitrogen)

diluted at 1:250 in PBS (10 mM sodium phosphate, pH 7.4 at 22 °C, containing 0.15 M NaCl). Cells were mounted in Prolong Gold Antifade reagent containing DAPI (Invitrogen) to visualize cell nuclei, and images were acquired using MicroSuite FIVE software (Version 1.224, Olympus Soft Imaging Solutions Corp., Lakewood, DO) using an Olympus DP71 12.5 MP digital camera attached to an Olympus BX61 motorized microscope (Olympus America, Inc., Center Valley, PA). Representative micrographs are shown but each experiment was repeated at least three times using different preparations of Sertoli cells, which yielded similar results.

Real-Time PCR (qPCR). qPCR was performed as earlier described²⁰. In short, Sertoli cells cultured alone for 3 days were treated with PFOS (20 μ M) for 24 hr and terminated on day 4. Total RNA was isolated from these Sertoli cells using Trizol (Life Technologies) and reversed transcribed to cDNAs using Moloney murine leukemia virus reverse transcriptase (Promega). qPCR was performed using the QuantStudio™ 12 K Flex Real-time PCR System (Thermo Fisher) at the Rockefeller University Genomics Resource Center ($n = 3$ experiments, each in triplicate) using a primer pair specific to rat Cx43 (sense: 5'-ACTCTCGCCTATGTCTCTCT-3', nucleotides 1023–1041; antisense: 5'-CGAGTTGGAGATGGTGCTT-3', nucleotides 1164–1182; GenBank Accession Number: BC081842.1), and a primer pair specific to GAPDH for co-amplification as described²⁰ which served as the internal control for normalization. The specificity of the fluorescence signal was confirmed by both melting curve analysis and gel electrophoresis. The steady-state level of Cx43 was then determined using the $2^{-\Delta\Delta CT}$ method.

GJ Communication Assay by Fluorescent Dye-transfer. A fluorescence-based GJ-intercellular communication assay to assess the effects of overexpression of Cx43 on Sertoli cells treated with PFOS was performed as described^{16,25,46}. Sertoli cells cultured at 0.15×10^6 cells/cm² on Matrigel-coated (diluted 1:7 with F12/DMEM) glass-bottom dishes (Cat #: P35G-0-20-C, MatTek) were transfected with plasmid DNA for overexpression of Cx43 vs. empty pCI-neo vector (control) on day 2 for 6 hr, and then exposed to PFOS vs. without PFOS (control) on day 3 for 24 hr. Cells were labeled with 5 μ M calcein AM (Mr 994.87, Invitrogen) for 30 min at 35 °C. Cell membrane-permeable calcein AM would be converted in living cells into membrane-impermeable and fluorescent calcein. Extracellular calcein AM was then removed after incubation and cells were rinsed with F12/DMEM twice. A few selected Sertoli cells were photobleached and the FRAP (fluorescence recovery after photobleaching) was performed by monitoring changes in the fluorescent intensity of the selected Sertoli cell that assessed the transfer of fluorescent dye from adjacent Sertoli cells to the photobleached cell. GJ communication assay was performed at the Rockefeller University BioImaging Resource Center using confocal microscopy (Zeiss/Perkin-Elmer) equipped with a Digital Mosaic system (Photonics Instruments) for photobleaching and to acquire images from 0 to 120 sec with Sertoli cells maintained in a culture chamber at 35 °C in a humidified atmosphere of 5% CO₂/95% air (v/v). Images were acquired using an Andor iXon EMCCD camera and MetaMorph software package (Molecular Devices), and stacked images were aligned using the StackReg plug-in in ImageJ for analysis.

Image analysis. Image analysis to assess the fluorescent intensity (FI) or fluorescent signal (FS) at the Sertoli cell-cell interface was performed as described⁴⁴. At least 200 cells were randomly selected and examined in control vs. experimental groups with $n = 3$ experiments (i.e., ~70 randomly selected cells per experiment). For changes in protein localization near the Sertoli cell surface, the distribution of the fluorescent signals at the cell-cell interface was measured at the two opposite ends (i.e., 4 measurements) of the Sertoli cell nucleus, which was then averaged to obtain the mean width. The fluorescent intensity of a target protein in Sertoli cells was quantified using ImageJ 1.45 software (NIH, Bethesda, MD; <http://rsbweb.nih.gov/ij>).

Statistical Analysis. Statistical significance was determined by two-way ANOVA, followed by Dunnett's procedure using GB-STAT Statistical Analysis Software Package (Version 7.0, Dynamic Microsystems, Silver Spring, MD). All experiments were repeated at least three times using different batches of Sertoli cell cultures, and each time point had triplicate or quadruplicate (for TER measurements that monitored TJ-permeability barrier function) wells/dishes.

Results

PFOS (Perfluorooctanesulfonate) Down-Regulates Cx43 expression and Perturbs the Organization of Actin Microfilaments at the Sertoli cell Basal ES (Ectoplasmic Specialization)/BTB (Blood-Testis Barrier).

The BTB, unlike other blood-tissue barriers such as the blood-brain barrier which is constituted exclusively by the TJ-barrier of the microvessels with support from pericytes in the brain (for a review, see ref. 47), is created by coexisting TJ, basal ES and GJ between adjacent Sertoli cells close to the base of the seminiferous tubule (for reviews, see refs 48–50). The most typical feature of the basal ES/BTB is the network of actin microfilaments that are extensively bundled that lie perpendicular to the Sertoli cell plasma membrane and sandwiched in between cisternae of endoplasmic reticulum and the Sertoli cell plasma membrane (for reviews, see refs 48 and 51), illustrating the significance of the Sertoli cell F-actin network in supporting BTB function. Studies have shown that Sertoli cells cultured *in vitro* establish a functional TJ-barrier that mimics the Sertoli BTB *in vivo*, which have been used extensively in the field by investigators as a model to study BTB function for almost three decades^{40–42,52–55}. Exposure of Sertoli cells cultured *in vitro* with an established functional TJ-permeability barrier to PFOS at 20 μ M (a concentration selected based on an earlier dosing study which yielded a distinctive phenotype on the Sertoli cell TJ-barrier function without unwanted cytotoxicity²⁵) using the regimen summarized in Fig. 1A, PFOS was found to down-regulate Cx43 expression considerably, in particular the Cx43 p2 isoform of Mr 43 kDa, to be followed by the p1 isoform of Mr 41 kDa, but a mild up-regulation of p0 isoform (Mr 39 kDa) (Fig. 1B). Our findings that Cx43 is a heterogeneous protein, displaying apparent Mr of 39, 41 and 43 kDa corresponding to p0, p1 and p2 (Fig. 1B) are consistent with earlier reports^{56,57}. This down-regulation of Cx43 by PFOS was also supported by findings using qPCR, illustrating a considerable reduction in its steady-state

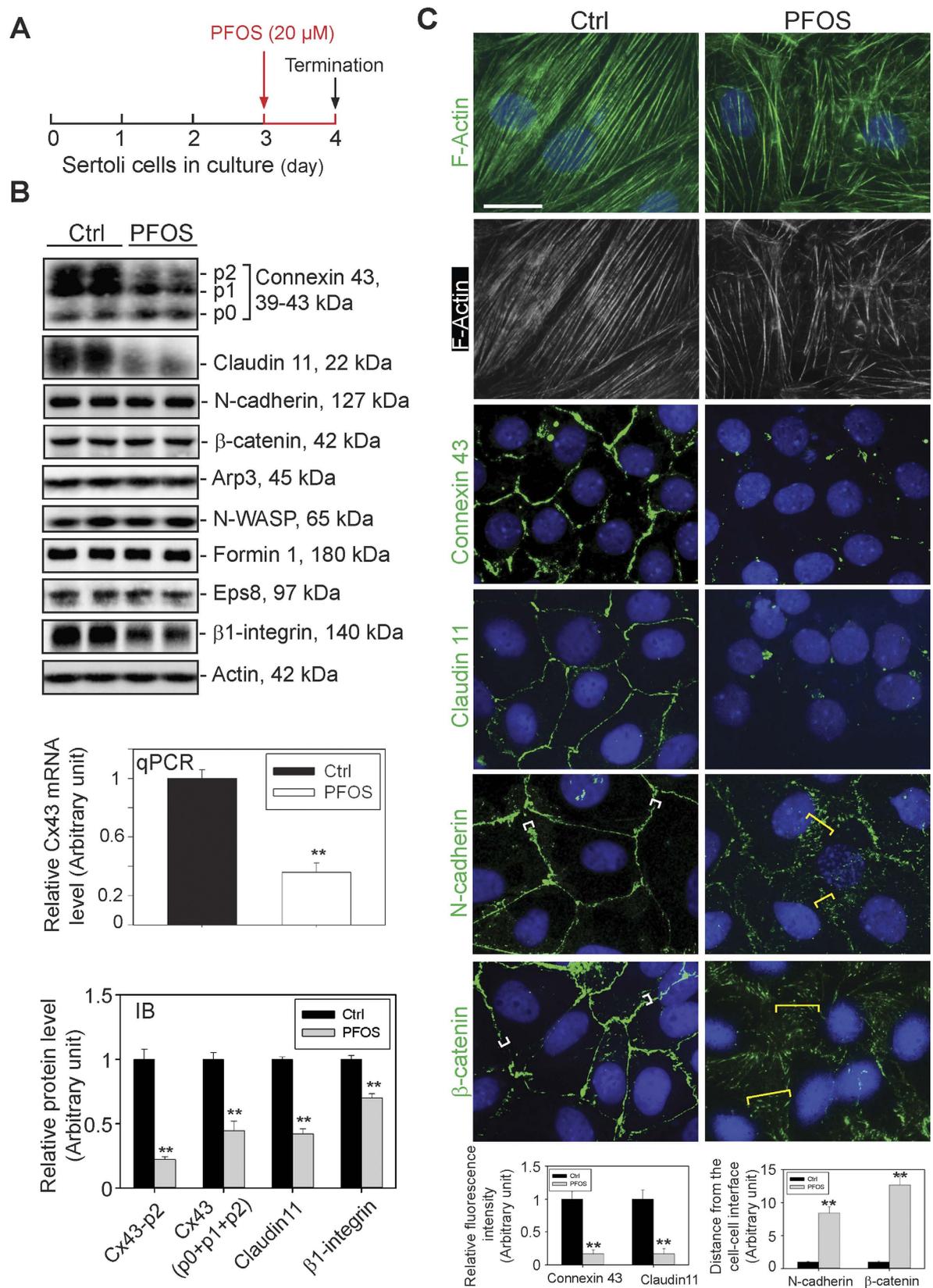


Figure 1. Disruptive effects of PFOS on Sertoli cell F-actin organization and distribution of BTB-associated proteins at the cell-cell interface. (A) Treatment regimen used for this experiment. Sertoli cells cultured alone at 0.4×10^6 cell/cm² for 3 days following the establishment of a functional TJ-barrier were treated with 20 μ M PFOS for 24 hr. Thereafter, cells were harvested for immunoblotting or for immunofluorescent microscopy. (B) Immunoblot analysis to assess changes in the steady-state level of Cx43 (a GJ protein), TJ- (e.g., claudin 11), basal ES (e.g., N-cadherin, β -catenin), and actin binding/regulatory proteins (e.g., Arp3, N-WASP, formin 1 and Eps8) with β -actin serving as the protein loading control. Cx43 appeared as three immunoreactive bands

designated as p0, p1 and p2, corresponding to Mr of 39, 41, and 43 kDa due to differential phosphorylation, which is consistent with earlier reports^{56,57}. A study by qPCR (with GAPDH serving as the internal control for normalization) also confirmed a down-regulation of Cx43 expression. Immunoblot analysis also illustrated a down-regulation on the steady-state level of Cx43 protein when p0, p1 and p2 isoforms were analyzed as a whole, with a more considerable reduction for p2. Besides Cx43, claudin 11 and β 1-integrin were also down-regulated. Each bar in the two bar graphs is a mean \pm SD of $n = 5$ experiments. $**P < 0.01$. (C) F-actin organization in Sertoli cells was assessed by phalloidin-FITC staining, illustrating PFOS induced truncation and defragmentation of actin microfilaments. PFOS treatment also considerably down-regulated the expression of Cx43 and claudin 11 at the Sertoli-Sertoli cell interface, consistent with data shown in (B). While the levels of N-cadherin and β -catenin were not considerably affected based on immunoblotting results, these basal ES proteins localized diffusely at the Sertoli cell-cell interface, apparently rapidly internalized. Different cell densities were used to optimize the visualization of different target proteins including F-actin as noted in *Materials and Methods*. Micrographs shown herein are representative results of an experiment. All three independent experiments yielded similar results. Scale bar, 25 μ m, which applies to all other micrographs. Each bar graph is a mean \pm SD of $n = 3$ experiments. $**P < 0.01$.

mRNA level, consistent with immunoblotting data shown in Fig. 1B. For instance, a considerable down-regulation on the steady-state protein of not just p2, but Cx43 as a whole was noted. In short, the expression of Cx43 was down-regulated by PFOS, not just its phosphorylation status. PFOS also down-regulated claudin 11 considerably, apparently a less phosphorylated form due to a reduced Mr (Fig. 1B). However, PFOS did not alter the expression of several other BTB-associated proteins, including actin binding and regulatory proteins Arp3 (an actin barbed end nucleation protein causing actin filament branching), N-WASP (an upstream activator of the Arp2/3 complex), formin 1 (an actin nucleation protein inducing long stretches of actin microfilaments) and Eps8 (an actin barbed end capping and bundling protein), except β 1-integrin which is a hemidesmosome protein³⁸ was found to be down-regulated (Fig. 1B). More important, the organization of actin microfilaments in Sertoli cells was found to be grossly affected in which defragmentation was extensively found in the cell cytosol in which actin microfilaments were truncated, appearing as shorter stretches of fragments (see both the green fluorescence and black-and-white images in Fig. 1C). Truncation of actin microfilaments in Sertoli cells failed to support localization of GJ protein Cx43, TJ protein claudin 11 and also basal ES proteins N-cadherin and β -catenin so that these adhesion proteins no longer tightly localized at the Sertoli cell-cell interface (Fig. 1C).

Overexpression of Cx43 in Sertoli cells Promotes TJ-Barrier Function, Rescuing PFOS-Induced BTB Disruption. Studies have shown that GJ-based Cx43 is crucial to confer BTB function in the rat testis by mediating cross-talk between various types of junctions at the BTB, such as TJ and basal ES, in order to maintain BTB homeostasis¹⁵. Additionally, the expression of Cx43 is essential to confer TJ reassembly during BTB remodeling¹⁶, such as during the transport of preleptotene spermatocytes across the immunological barrier. A recent study also reported that overexpression of Cx43 rescued toxicant-induced BTB disruption by resealing the disrupted BTB, re-initiating meiosis using the adjuvins model²⁰ since an acute dose of adjuvins was shown to cause meiotic arrest¹⁹. Using the regimen shown in Fig. 2A, overexpression of Cx43 in Sertoli cells was confirmed by immunoblotting (Fig. 2B). Interestingly, TJ protein claudin 11 expression was also moderately up-regulated, and also hemidesmosome protein β 1-integrin but not other BTB-associated proteins examined following overexpression of Cx43 (Fig. 2B). These findings were also summarized and shown in the bar graphs on the right panel (Fig. 2B). It was noted that the efficiency of overexpression of Cx43 using primary Sertoli cells cultured *in vitro* was ~25–35% as noted in the bar graph shown in Fig. 2B (upper panel) when cells were harvested on days 4 and 5 at 48 and 72 hr following the transfection of Sertoli cells with plasmid DNA. This relatively low efficiency may be the result of timing to terminate the cells. Nonetheless, overexpression of Cx43 was found to promote the Sertoli cell TJ-permeability barrier (Fig. 2C). Furthermore, Cx43 overexpression also rescued PFOS-induced Sertoli cell TJ-permeability barrier disruption (Fig. 2C). These findings were also corroborated by quantifying changes in the Cx43 steady-state using Sertoli cell lysates obtained on day 5 in the four experimental groups, illustrating overexpression of Cx43 partially restored the PFOS-induced Cx43 down-regulation (Fig. 2C, right panel).

Overexpression of Cx43 rescues the PFOS-mediated GJ communication dysfunction. Using the regimen shown in Fig. 3A, PFOS was found to impede Sertoli cell GJ communication function based on the use of a dye-transfer functional assay by quantifying the ability of GJs between Sertoli cells to transfer fluorescent dye across the GJ to a photobleached cell (Fig. 3B) in a FRAP assay. Importantly, overexpression of Cx43 was found to rescue the PFOS-induced GJ-based intercellular communication dysfunction (Fig. 3B), and also supported by findings of fluorescent dye transfer assay of $n = 3$ experiments (Fig. 3C), illustrating Cx43 plays a critical role in GJ communication function in the testis.

Overexpression of Cx43 re-organizes TJ- and basal ES-protein distribution at the Sertoli cell-cell interface disrupted by PFOS. Following exposure of Sertoli cell epithelium *in vitro* to PFOS, the localization of Cx43, TJ- (e.g., claudin 11, ZO-1) and basal ES- (e.g., N-cadherin, β -catenin) proteins at the Sertoli cell-cell interface was found to be grossly perturbed, either considerably down-regulated (e.g., Cx43, claudin 11) or mis-localized (e.g., ZO-1, N-cadherin, β -catenin) (Fig. 4). Overexpression of Cx43 moderately promoted localization of Cx43, as well as TJ and basal ES proteins at the Sertoli cell-cell interface (Fig. 4). More important, overexpression of Cx43 in Sertoli cells before these cells were exposed to PFOS was found to rescue these cells from the disruptive effects of PFOS. For instance, Cx43 overexpressed Sertoli cells displayed proper expression

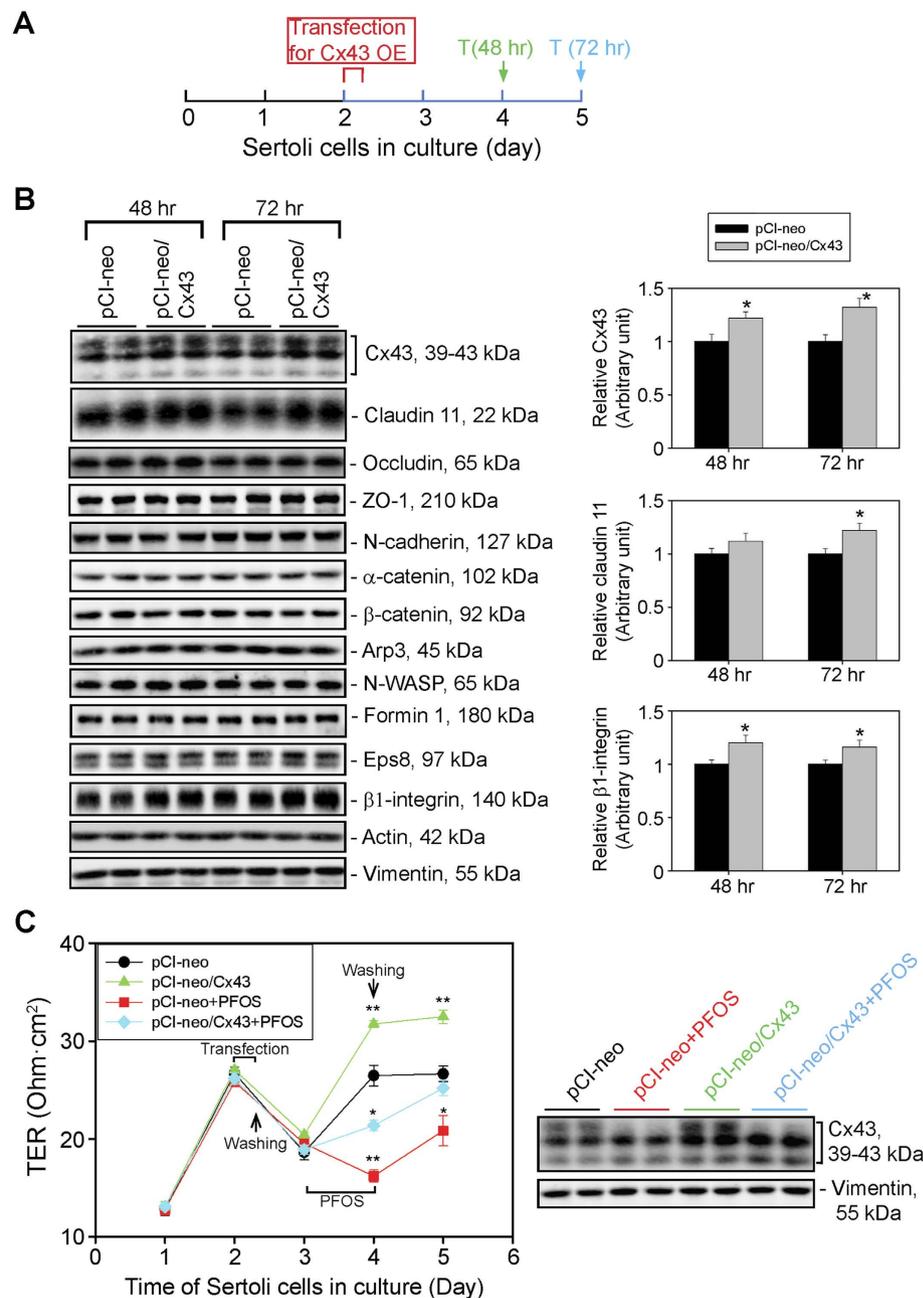


Figure 2. Overexpression of Cx43 in Sertoli cells with an established TJ-permeability barrier promotes barrier function, rescuing PFOS-induced barrier disruption. (A) Treatment regimen used to assess the effects of overexpression of Cx43 in Sertoli cells on the steady-state levels of BTB-associated proteins by immunoblot analysis. T, termination. (B) Immunoblot analysis using Sertoli cell lysates to assess changes in the steady-state levels of selected BTB-associated proteins, including Cx43 following its overexpression at 48- and 72-hr after transfection (i.e., on days 4 and 5, respectively). Among all the BTB-associated proteins examined, only the TJ protein claudin 11 and the hemidesmosome protein β 1-integrin displayed a moderate up-regulation following Cx43 overexpression as illustrated in the bar graphs on the right panel. Each bar in the bar graphs is a mean \pm SD of $n = 3$ experiments. * $P < 0.05$. (C) Sertoli cells were cultured on Matrigel-coated bicameral units at 1.0×10^6 cells/cm² for two days to establish a functional TJ-permeability barrier. Cells were then transfected with pCI-neo empty vector vs. pCI-neo vector containing Cx43 for overexpression of Cx43 (pCI-neo/Cx43) in these cells for 6 hr. Thereafter, cells were washed twice with F12/DMEM and cultured for another 18 hr, followed by an incubation with or without (i.e., equal amount of DMSO-containing medium) 20 μ M PFOS for 24 hr. Cells were then rinsed twice to remove PFOS and cultured in F12/DMEM for another 24 hr. TER was recorded daily using quadruple bicameral units for each time-point. Overexpression of Cx43 in Sertoli cells was found to promote the TJ-permeability barrier and alleviate PFOS-induced TJ-disruption. Each data point is a mean \pm SD of quadruple bicameral units of a representative experiment from $n = 3$ experiments, which yielded similar results. * $P < 0.05$; ** $P < 0.01$. Sertoli cell lysates obtained on day 5 (see left panel, and 2A for the treatment regimen) were also used for immunoblotting, illustrating overexpression of Cx43 partially rescued the PFOS-induced Cx43 down-regulation. Vimentin served as the protein loading control.

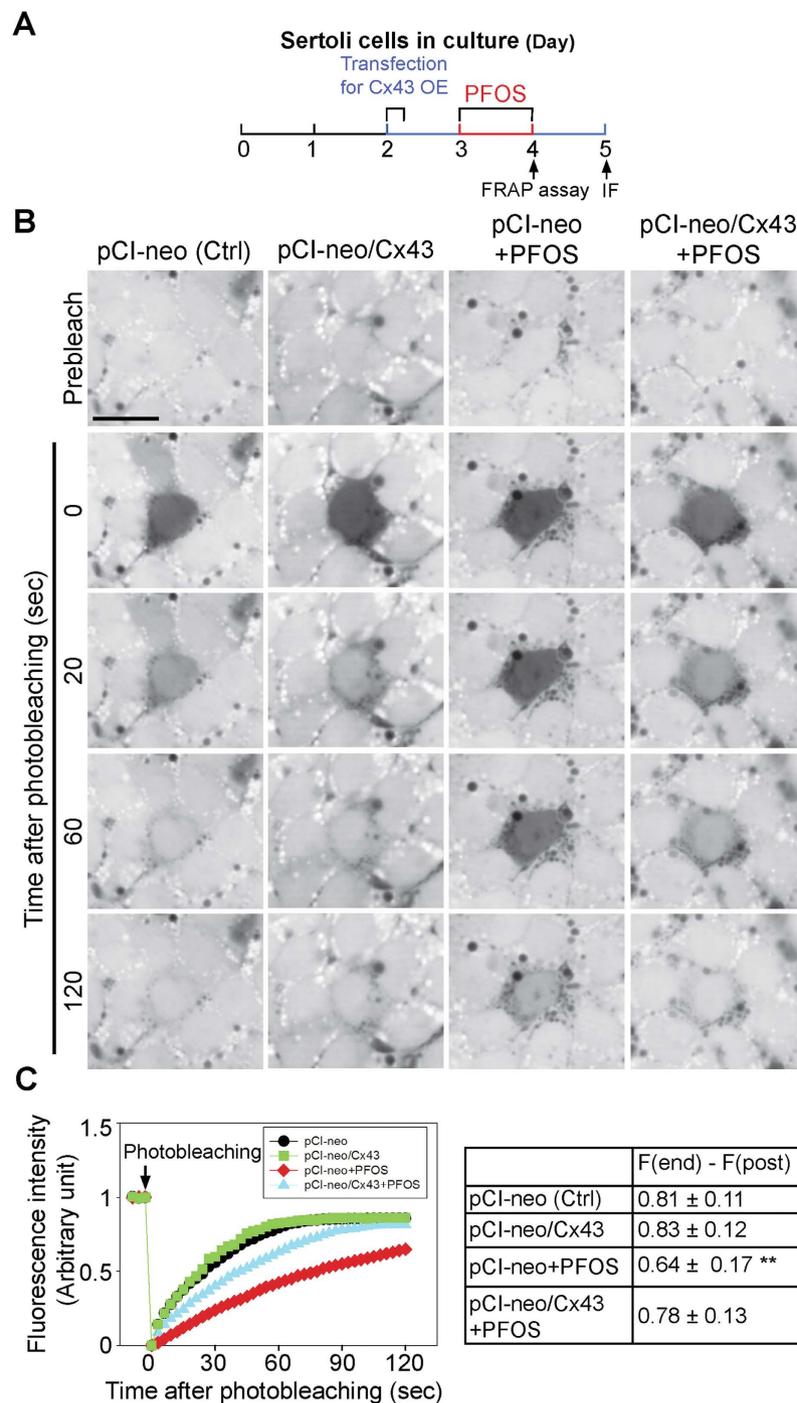


Figure 3. Overexpression of Cx43 in Sertoli cells rescues PFOS-induced GJ communication dysfunction. (A) Treatment regimen used to determine if overexpression (OE) of Cx43 can rescue PFOS-induced GJ communication function based on a dye-transfer FRAP (fluorescence recovery after photobleaching) functional assay. (B) GJ intercellular communication function between Sertoli cells cultured at 0.15×10^6 cells/cm² was assessed by a dye-transfer FRAP assay based on the transfer of fluorescent dye (calcein AM) from neighboring cells via GJ following photobleaching of a single Sertoli cell. Sertoli cells were cultured at 0.15×10^6 cells/cm² on Matrigel-coated glass-bottom dishes for 2 days. The cells were then transfected with pCI-neo/Cx43 vs. pCI-neo empty vector for 6 hr. Thereafter, cells were washed twice with F12-DMEM and cultured for another 18 hr, followed by treatment with 20 μ M PFOS for 24 hr (with or without Cx43 overexpression vs. Cx43 overexpression alone or empty vector alone). Results from a typical FRAP assay are shown with $n = 3$ experiments, which yielded similar results. Scale bar, 40 μ m, which applies to all other micrographs. (C) Typical results of a representative FRAP assay from $n = 3$ experiments that yielded similar results, illustrating GJ communication was impaired by PFOS treatment, but overexpression of Cx43 rescued the PFOS-induced GJ dysfunction. Each data point on the right panel is a mean \pm SD of $n = 4$ independent experiments for the four experimental groups using different batches of Sertoli cells for the FRAP assay. ** $P < 0.01$.

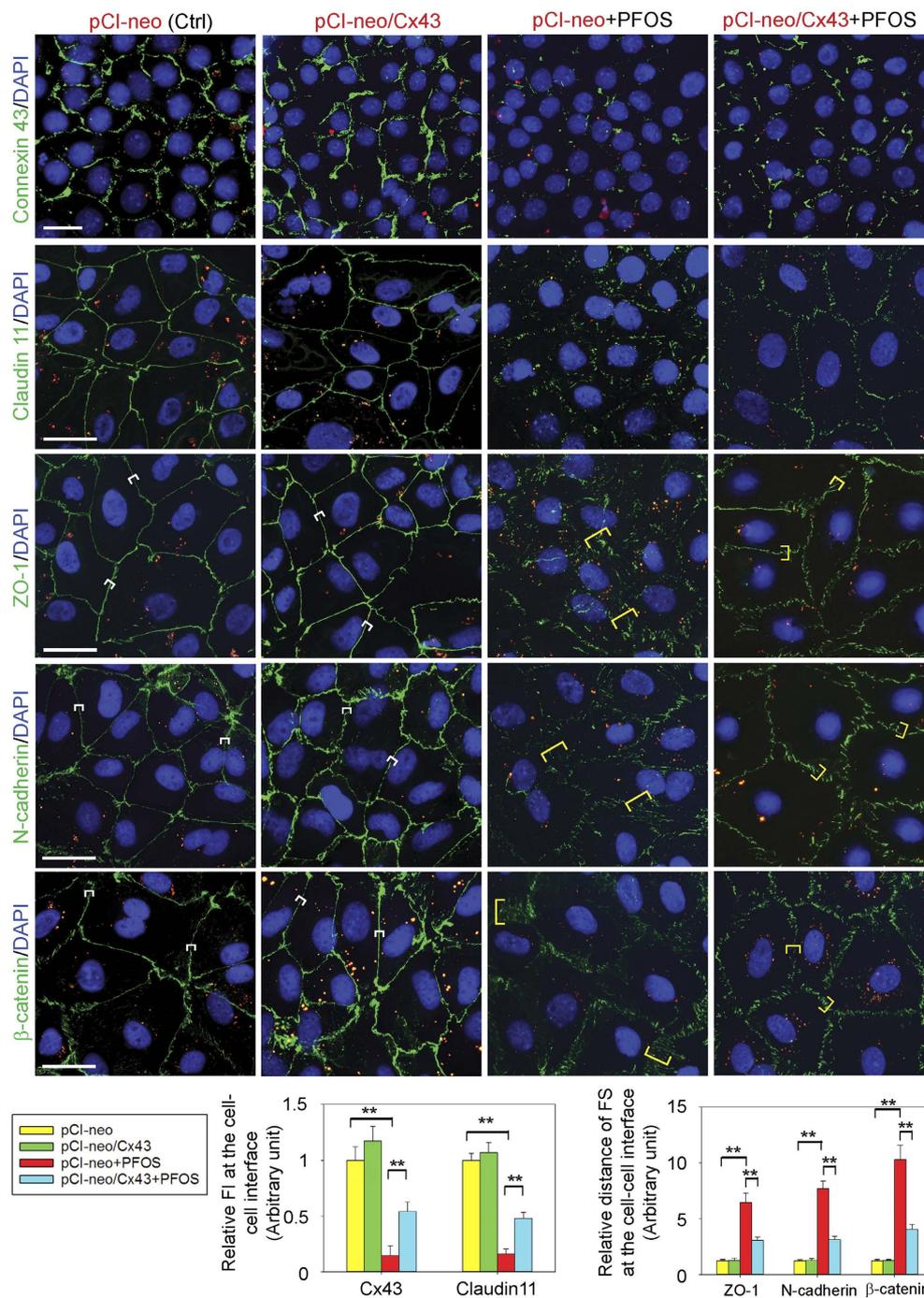


Figure 4. Overexpression of Cx43 in Sertoli cells rescues PFOS-induced Sertoli cell BTB disruption through proper localization of adhesion proteins at the cell-cell interface. Sertoli cells were cultured at $0.1\text{--}0.15 \times 10^6$ cells/cm² for two days. They were transfected with pCI-neo/Cx43 vs. pCI-neo empty vector (control, Ctrl) and then subjected to PFOS treatment, and cells were harvested for IF analysis as noted in the regimen shown in Fig. 3A. In cells overexpressed with Cx43 (pCI-neo/Cx43), the fluorescent intensity for Cx43 at cell-cell interface was moderately stronger than control (pCI-neo empty vector alone) cells. However, PFOS induced considerable down-regulation of Cx43 and claudin 11 at the cell-cell interface (pCI-neo+PFOS), PFOS also perturbed the localization of TJ protein ZO-1, and basal ES proteins N-cadherin and β-catenin. Importantly, overexpression of Cx43 in PFOS treated cells (pCI-neo/Cx43+PFOS) rescued the PFOS-mediated BTB disruption by maintaining the expression of Cx43 and claudin 11. This also induced proper localization of TJ protein ZO-1 and basal ES proteins N-cadherin and β-catenin at the cell-cell interface. Plasmid DNA (pCI-neo vs. pCI-neo/Cx43) was labeled with Cy3 and appeared as red fluorescence to annotate successful transfection. Scale bar, 30 μm, in the first micrograph, which applies to the remaining micrographs in the same panel. These results are representative micrographs from an experiment, and $n = 3$ independent experiments were performed using different batches of Sertoli cells and yielded similar results. Each bar graph is a mean \pm SD of $n = 3$ experiments. ** $P < 0.01$. FI, fluorescent intensity; FS, fluorescent signal.

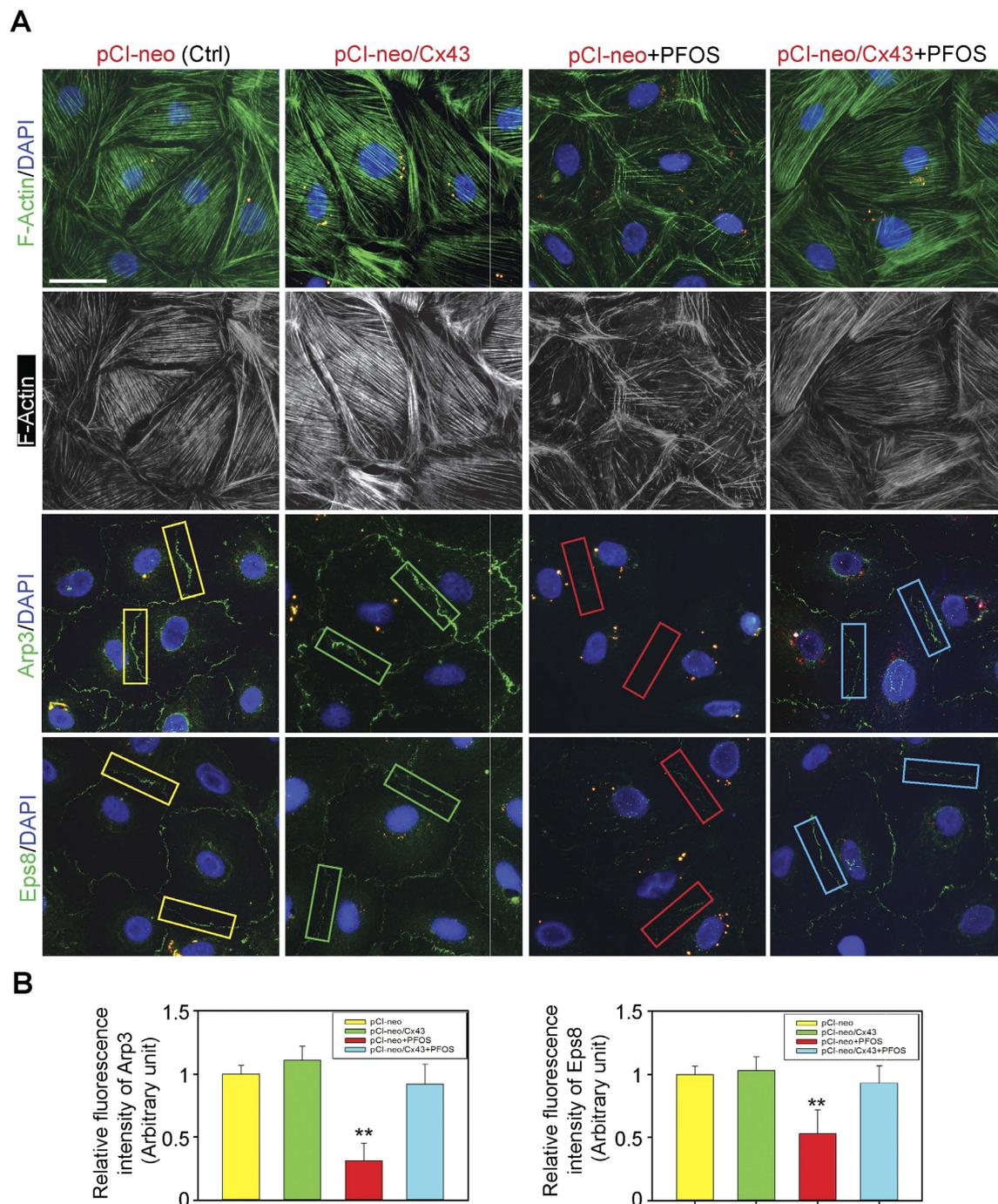


Figure 5. Overexpression of Cx43 in Sertoli cells restores actin microfilament organization via proper spatiotemporal expression of actin binding/regulatory proteins Arp3 and Eps8. (A) We next examined the mechanism by which Cx43 overexpression rescues the PFOS-induced Sertoli cell TJ-barrier disruption. Exposure of Sertoli cells with an established functional TJ-permeability barrier to PFOS for 24 hr was found to induce mis-organization of actin microfilaments wherein they displayed extensive defragmentation across the Sertoli cell cytosol. However, overexpression of Cx43 restored organization of actin microfilaments in Sertoli cells, apparently via proper spatiotemporal expression of actin binding and regulatory proteins Arp3 and Eps8, such that these proteins localized to the Sertoli cell-cell interface, similar to the control cells expressed with the empty pCI-neo vector. Plasmid DNA (pCI-neo vs. pCI-neo/Cx43) was labeled with Cy3 and appeared as red fluorescence to annotate successful transfection. Scale bar, 30 μ m, which applies to all other micrographs. (B) Relative fluorescent intensity of Arp3 and Eps8 at the Sertoli cell-cell interface was quantified in the PFOS treatment group (pCI-neo+PFOS) vs. control (Ctrl, transfected with pCI-neo vector alone), Cx43 overexpression (pCI-neo/Cx43) and Cx43 overexpression plus PFOS (pCI-neo/Cx43+PFOS) groups. Each bar is a mean \pm SD by taking the average fluorescent intensity from two opposite ends between adjacent Sertoli cells (see corresponding colored boxed area for Arp3 vs. Eps8 from different groups) from 50 randomly selected cells in each experiment with $n = 3$ independent experiments. ** $P < 0.01$.

and/or localization of BTB-associated proteins at the Sertoli cell-cell interface exposed to PFOS (Fig. 4), making them more similar to control cells transfected with empty vector (Fig. 4). These findings thus support earlier observations that overexpression of Cx43 rescued the PFOS-induced Sertoli cell TJ-permeability barrier disruption as noted in Fig. 2C.

Overexpression of Cx43 Induces reorganization of Actin Microfilaments in Sertoli Cells Exposed to PFOS Through Proper Spatiotemporal Expression of Arp3 and Eps8.

To better understand the molecular mechanism by which overexpression of Cx43 that rescues PFOS-induced F-actin dis-organization, we next examined the possible involvement of Arp3 and Eps8 (Fig. 5). Overexpression of Cx43 in Sertoli cells exposed to PFOS induced re-organization of actin microfilaments in which microfilaments were no longer found to be considerably truncated vs. PFOS treated cells without Cx43 overexpression (Fig. 5A), which thus supported proper localization of adhesion proteins as noted in Fig. 4. More important, this corrective organization of actin microfilaments in Sertoli cells following Cx43 overexpression in PFOS treated cells was mediated by proper spatial expression of barbed end actin nucleation protein Arp3 in Sertoli cells and also actin barbed end capping and bundling protein Eps8, making their pattern of localization analogous to control cells (Fig. 5A), and the corrective spatial localization and/or expression of these two actin regulatory proteins were summarized in Fig. 5B. These findings thus support the notion that Cx43 exerts its effects by conferring proper organization of actin microfilaments in Sertoli cells to rescue the PFOS-induced TJ-barrier dysfunction. It is noted that this reduced Arp3 vs. Eps8 expression at the Sertoli cell-cell interface as noted herein (Fig. 5A,B) represented a considerable shift in their distribution instead of a down-regulation in expression since these two actin binding/regulatory proteins appeared to be internalized and possibly distributed evenly across the Sertoli cell cytosol. This notion was indeed supported by no significant change in the overall expression of Arp3 or Eps8 after PFOS treatment (see Fig. 1B).

Discussion

GJs are important cell junctions that confer cell-cell communication and are known to coordinate cellular events across a cell epithelium, such as the seminiferous epithelium during the epithelial cycle of spermatogenesis. For instance, studies have shown that GJs serve as the ultrastructure to transfer chemicals and/or signaling molecules up to ~1000 daltons between adjacent cells in an epithelium (for reviews, see refs 4,59 and 60). Studies by physiologists and toxicologists have also shown that GJs play a critical role in transcriptional regulation and/or toxicant-mediated epigenetic gene modifications since gap junctional intercellular communication (GJIC) is an important mechanism that integrate signaling pathways, which in turn control gene expression between epithelial cells of an epithelium (for reviews, see refs 61–64). Thus, if a toxicant, such as PFOS, cadmium or BPA (bisphenol A), perturbs GJIC function in the Sertoli cell of the testis, multiple signaling pathways can be compromised. This might result in male reproductive dysfunction. Studies that define the molecular mechanisms by which PFOS mediates its toxicity in mammalian cells such as the Sertoli cell in the testis are important since PFOS, an environmental toxicant, is still commonly used in China and other developing countries because of its beneficial properties in consumer products such as serving as a stain repellent in clothing, carpets and draperies. However, its use in the U.S. and Canada has been banned in the late 2000s. Herein, PFOS was shown to block GJIC between Sertoli cells using a dye-transfer cell-cell communication assay based on FRAP²⁵. Since studies have shown that Cx43 is an important GJ-based protein in the testis and studies have shown that it is also a target of environmental toxicants (for a review, see ref. 10). Furthermore, a recent report using adjuvin-induced sterility via the use of an acute dose as a model, the phenotypes of the testis^{19,20} mimicked many of the disruptive changes in the testis found in Sertoli cell-specific Cx43 KO mice^{11,13}. Interestingly, overexpression of Cx43 in the adjuvin-treated testes was found to re-initiate spermatogenesis in particular re-booting meiosis since round spermatids were detected in a significant number of tubules²⁰. However, spermiogenesis failed to occur since no elongating/elongated spermatids were detected in any of the tubules with signs of meiosis²⁰. Nonetheless, studies using the adjuvin model vs. the Sertoli cell-specific Cx43 KO genetic models^{11,13} have demonstrated the significance of Cx43-based GJ function since the presence of at least nine other GJs found in the testis⁷ failed to replace the lost function of Cx43. However, it is of interest to note that while treatment of Sertoli cell epithelium with PFOS reduces GJ-based communication function, the evidence that this partial uncoupling is responsible for the effects of the toxicant is corroborative in nature since PFOS is plausibly mediated its toxicity through multiple pathways. However, overexpression of Cx43 was found to re-establish the disrupted actin microfilament organization, reseal the Sertoli TJ-barrier, and re-distribute TJ and basal ES proteins to the Sertoli cell-cell interface. Moreover, these changes were mediated by proper spatiotemporal expression of Arp3 and Eps8. These findings thus support the notion that Cx43 disruption induced by PFOS is one of the major mechanisms underlying PFOS-induced Sertoli cell injury.

These earlier findings thus prompted us to examine the role of Cx43 in correcting PFOS-mediated Sertoli cell injury by overexpressing Cx43 in the Sertoli cells exposed to this environmental toxicant as reported herein. It was noted that successful overexpression of Cx43 in Sertoli cells would repair the disrupted GJIC between these cells. More important, this re-establishment of GJIC in the Sertoli cell epithelium *in vitro* with an established functional TJ-permeability barrier that mimics the Sertoli BTB *in vivo* is crucial to re-organize the Sertoli cell actin microfilaments, making their organization similar to control Sertoli cells in which long stretches of microfilaments lay across the cell cytosol instead of grossly truncated following PFOS exposure, thereby supporting the proper adhesion protein complex localization (e.g., occludin/ZO-1, N-cadherin/ β -catenin which utilize F-actin for their attachments) via corrective spatial expression of actin binding and regulatory proteins such as Arp3 and Eps8. These findings thus support the notion that the re-established GJ-based communication following Cx43 overexpression is used to maintain proper communications between Sertoli cells. This re-established GJ function in turn supports better organization of the cytoskeleton to maintain the Sertoli cell TJ-barrier function.

It is noted that Cx43 global KO leads to perinatal death due to defects in neural crest formation, vascular development, and heart malformation^{65–68}. A recent report has shown that using mouse embryonic fibroblasts (MEFs) obtained from Cx43 knockout (KO) mice for an *in vitro* wound closure assay, these MEFs were found to display defects in cell polarity and directional cell movement due to defects in microtubule organizing center (MOC) as a result of Cx43 deficiency⁶⁹. Furthermore, actin microfilaments at the wound edge of these MEFs also failed to properly align to confer directional cell movement, and these defects in actin- and microtubule-based cytoskeletal functions could be restored, at least in part, following overexpression of Cx43⁶⁹. These findings thus support our observations that Cx43 mediates its effects through proper GJIC to re-establish the necessary signaling function to confer proper cytoskeletal organization. Work is now in progress to identify the downstream signaling function involved in these events.

In summary, we have demonstrated that PFOS exerts its toxic effects in the testis by disrupting the Cx43-based GJIC, which in turn perturbs the underlying actin-based cytoskeletal function. This thus affects cell adhesion protein complexes at the Sertoli cell-cell interface which utilize F-actin for their attachment, thereby disrupting the Sertoli cell TJ-permeability barrier function. However, these disruptive effects can be rescued via overexpression of Cx43 in PFOS-treated Sertoli cells. Further investigations should include studies to manipulate cytoskeletal function through their binding and regulatory proteins, such as Arp3, Eps8 and palladin for F-actin and EB1 and MARKs for MTs, attempting to block the PFOS-induced reproductive dysfunction.

References

- Bai, D. Structural analysis of key gap junction domains - lessons from genome data and disease-linked mutants. *Semin Cell Dev Biol*, doi: 10.1016/j.semcdb.2015.11.015 (2015).
- Balse, E. *et al.* Dynamic of ion channel expression at the plasmamembrane of cardiomyocytes. *Physiol Rev* **92**, 1317–1358 (2012).
- Gerber, J., Heinrich, J. & Brehm, R. Blood-testis barrier and Sertoli cell function: lessons from SCCx43KO mice. *Reproduction* **151**, R15–27, doi: 10.1530/REP-15-0366 (2016).
- Li, M. W. M., Mruk, D. D. & Cheng, C. Y. Gap junctions and blood-tissue barriers. *Adv Exp Med Biol* **763**, 260–280 (2012).
- Pointis, G., Gilleron, J., Carette, D. & Segretain, D. Physiological and physiopathological aspects of connexins and communicating gap junctions in spermatogenesis. *Philos Trans R Soc Lond B Biol Sci* **365**, 1607–1620 (2010).
- Pointis, G. & Segretain, D. Role of connexin-based gap junction channels in testis. *Trends Endocrinol Metab* **16**, 300–306 (2005).
- Risley, M. Connexin gene expression in seminiferous tubules of the Sprague-Dawley rat. *Biol Reprod* **62**, 748–754 (2000).
- Chung, S. S. W., Lee, W. M. & Cheng, C. Y. Study on the formation of specialized inter-Sertoli cell junctions *in vitro*. *J Cell Physiol* **181**, 258–272 (1999).
- Risley, M., Tan, I., Roy, C. & Saez, J. Cell-, age-, and stage-dependent distribution of connexin 43 gap junctions in testes. *J Cell Sci* **103**, 81–96 (1992).
- Pointis, G., Gilleron, J., Carette, D. & Segretain, D. Testicular connexin 43, a precocious molecular target for the effect of environmental toxicants on male fertility. *Spermatogenesis* **1**, 303–317 (2011).
- Brehm, R. *et al.* A Sertoli cell-specific knockout of connexin43 prevents initiation of spermatogenesis. *Am J Pathol* **171**, 19–31 (2007).
- Giese, S. *et al.* Sertoli-cell-specific knockout of connexin 43 leads to multiple alterations in testicular gene expression in prepubertal mice. *Disease models & mechanisms* **5**, 895–913, doi: 10.1242/dmm.008649 (2012).
- Carette, D. *et al.* Major involvement of connexin 43 in seminiferous epithelial junction dynamics and male fertility. *Dev Biol* **346**, 54–67 (2010).
- Gunther, S., Fietz, D., Weider, K., Bergmann, M. & Brehm, R. Effects of a murine germ cell-specific knockout of Connexin 43 on Connexin expression in testis and fertility. *Transgenic Res* **22**, 631–641, doi: 10.1007/s11248-012-9668-1 (2013).
- Li, M. W. M., Mruk, D. D., Lee, W. M. & Cheng, C. Y. Connexin 43 and plakophilin-2 as a protein complex that regulates blood-testis barrier dynamics. *Proc Natl Acad Sci USA* **106**, 10213–10218 (2009).
- Li, M. W. M., Mruk, D. D., Lee, W. M. & Cheng, C. Y. Connexin 43 is critical to maintain the homeostasis of blood-testis barrier via its effects on tight junction reassembly. *Proc Natl Acad Sci USA* **107**, 17998–18003 (2010).
- Cheng, C. Y. Toxicants target cell junctions in the testis - insights from the indazole-carboxylic acid model. *Spermatogenesis* **4**, e981485, (doi: 10.4161/21565562.2014.9814895) (2014).
- Mruk, D. D., Silvestrini, B. & Cheng, C. Y. Anchoring junctions as drug targets: Role in contraceptive development. *Pharmacol Rev* **60**, 146–180 (2008).
- Mok, K. W., Mruk, D. D., Lee, W. M. & Cheng, C. Y. Spermatogonial stem cells alone are not sufficient to re-initiate spermatogenesis in the rat testis following adjuvant-induced infertility. *Int J Androl* **35**, 86–101 (2012).
- Li, N. *et al.* Connexin 43 reboots meiosis and reseals blood-testis barrier following toxicant-mediated aspermatogenesis and barrier disruption. *FASEB J* **30**, 1436–1452 (2016).
- Cheng, C. Y. & Mruk, D. D. Cell junction dynamics in the testis: Sertoli-germ cell interactions and male contraceptive development. *Physiol Rev* **82**, 825–874 (2002).
- Russell, L. D. & Peterson, R. N. Sertoli cell junctions: morphological and functional correlates. *Int Rev Cytol* **94**, 177–211 (1985).
- Vogl, A. W., Vaid, K. S. & Guttman, J. A. The Sertoli cell cytoskeleton. *Adv Exp Med Biol* **636**, 186–211 (2008).
- Wan, H. T., Mruk, D. D., Wong, C. K. C. & Cheng, C. Y. Targeting testis-specific proteins to inhibit spermatogenesis - lesson from endocrine disrupting chemicals. *Expert Opin Ther Targets* **17**, 839–855 (2013).
- Wan, H. T., Mruk, D. D., Wong, C. K. C. & Cheng, C. Y. Perfluorooctanesulfonate (PFOS) perturbs male rat Sertoli cell blood-testis barrier function by affecting F-actin organization via p-FAK-Tyr⁴⁰⁷ - an *in vitro* study. *Endocrinology* **155**, 249–262 (2014).
- Liu, C. & Zhang, N. Nanoparticles in gene therapy principles, prospects, and challenges. *Prog Mol Biol Transl Sci* **104**, 509–562 (2011).
- Mangraviti, A. *et al.* Polymeric nanoparticles for nonviral gene therapy extend brain tumor survival *in vivo*. *ASC Nano* **9**, 1236–1249 (2015).
- Kozielski, K. L., Rui, Y. & Green, J. J. Non-viral nucleic acid containing nanoparticles as cancer therapeutics. *Expert Opin Drug Deliv* (*in press*; PMID: 27248202) (2016).
- Mruk, D. D. & Cheng, C. Y. An *in vitro* system to study Sertoli cell blood-testis barrier dynamics. *Methods Mol Biol* **763**, 237–252 (2011).
- Orth, J. M. Proliferation of Sertoli cells in fetal and postnatal rats: A quantitative autoradiographic study. *Anat Rec* **203**, 485–492 (1982).
- Mather, J. P. *et al.* The hormonal and cellular control of Sertoli cell secretion. *Journal of steroid biochemistry* **19**, 41–51 (1983).
- Lui, W. Y., Lee, W. M. & Cheng, C. Y. Transforming growth factor- β 3 regulates the dynamics of Sertoli cell tight junctions via the p38 mitogen-activated protein kinase pathway. *Biol Reprod* **68**, 1597–1612 (2003).

33. Li, J. C. H., Lee, W. M., Mruk, D. D. & Cheng, C. Y. Regulation of Sertoli cell myotubularin (rMTM) expression by germ cells *in vitro*. *J Androl* **22**, 266–277 (2001).
34. Lee, N. P. Y., Mruk, D. D., Conway, A. M. & Cheng, C. Y. Zyxin, axin, and Wiskott-Aldrich syndrome protein are adaptors that link the cadherin/catenin protein complex to the cytoskeleton at adherens junctions in the seminiferous epithelium of the rat testis. *J Androl* **25**, 200–215 (2004).
35. Galdieri, M., Ziparo, E., Palombi, F., Russo, M. A. & Stefanini, M. Pure Sertoli cell cultures: a new model for the study of somatic-germ cell interactions. *J Androl* **2**, 249–254 (1981).
36. Wright, W., Zabludoff, S., Erickson-Lawrence, M. & Karzai, A. Germ cell-Sertoli cell interactions. Studies of cyclic protein-2 in the seminiferous tubule. *Annals of the New York Academy of Sciences* **564**, 173–185 (1989).
37. Li, J. C., Mruk, D. D. & Cheng, C. Y. The inter-Sertoli tight junction permeability barrier is regulated by the interplay of protein phosphatases and kinases: an *in vitro* study. *J Androl* **22**, 847–856 (2001).
38. Nicholls, P. K. *et al.* Activin signaling regulates Sertoli cell differentiation and function. *Endocrinology* **153**, 6065–6077, doi: 10.1210/en.2012-1821 (2012).
39. Sluka, P., O'Donnell, L., Bartles, J. R. & Stanton, P. G. FSH regulates the formation of adherens junctions and ectoplasmic specializations between rat Sertoli cells *in vitro* and *in vivo*. *J Endocr* **189**, 381–395 (2006).
40. Du, M. *et al.* A novel subcellular machine contributes to basal junction remodeling in the seminiferous epithelium. *Biol Reprod* **88**, 60 (2013).
41. Kaitu'u-Lino, T. J., Sluka, P., Foo, C. F. & Stanton, P. G. Claudin-11 expression and localisation is regulated by androgens in rat Sertoli cells *in vitro*. *Reproduction* **133**, 1169–1179 (2007).
42. Chen, J. *et al.* Cryptorchidism-induced CFTR down-regulation results in disruption of testicular tight junctions through up-regulation of NF- κ B/COX-2/PGE₂. *Hum Reprod* **27**, 2585–2597 (2012).
43. Lee, N. P., Mruk, D. D., Lee, W. M. & Cheng, C. Y. Is the cadherin/catenin complex a functional unit of cell-cell-actin-based adherens junctions (AJ) in the rat testis? *Biol Reprod* **68**, 489–508 (2003).
44. Li, N. *et al.* Actin-bundling protein plastin 3 is a regulator of ectoplasmic specialization dynamics during spermatogenesis in the rat testis. *FASEB J* **29**, 3788–3805 (2015).
45. Mruk, D. D. & Cheng, C. Y. Enhanced chemiluminescence (ECL) for routine immunoblotting. An inexpensive alternative to commercially available kits. *Spermatogenesis* **1**, 121–122 (2011).
46. Mok, K. W., Mruk, D. D., Lee, W. M. & Cheng, C. Y. Rictor/mTORC2 regulates blood-testis barrier dynamics via its effects on gap junction communications and actin filament network. *FASEB J* **27**, 1137–1152 (2013).
47. Easton, A. S. Regulation of permeability across the blood-brain barrier. *Adv Exp Med Biol* **763**, 1–19 (2012).
48. Cheng, C. Y. & Mruk, D. D. The blood-testis barrier and its implication in male contraception. *Pharmacol Rev* **64**, 16–64 (2012).
49. Mruk, D. D. & Cheng, C. Y. The mammalian blood-testis barrier: Its biology and regulation. *Endocr Rev* **36**, 564–591, doi: 10.1210/er.2014-1101 (2015).
50. Pelletier, R. M. The blood-testis barrier: the junctional permeability, the proteins and the lipids. *Prog Histochem Cytochem* **46**, 49–127 (2011).
51. Cheng, C. Y. & Mruk, D. D. A local autocrine axis in the testes that regulates spermatogenesis. *Nature Rev Endocrinol* **6**, 380–395 (2010).
52. Janeki, A., Jakubowiak, A. & Steinberger, A. Effect of cadmium chloride on transepithelial electrical resistance of Sertoli cell monolayers in two-compartment cultures - a new model for toxicological investigations of the "blood-testis" barrier *in vitro*. *Toxicol Appl Pharmacol* **112**, 51–57 (1992).
53. Okanlawon, A. & Dym, M. Effect of chloroquine on the formation of tight junctions in cultured immature rat Sertoli cells. *J Androl* **17**, 249–255 (1996).
54. Nicholls, P. K., Harrison, C. A., Gilchrist, R. B., Farnworth, P. G. & Stanton, P. G. Growth differentiation factor 9 is a germ cell regulator of Sertoli cell function. *Endocrinology* **150**, 2481–2490 (2009).
55. Qiu, L. *et al.* Sertoli cell is a potential target for perfluorooctane sulfonate-induced reproductive dysfunction in male mice. *Toxicol Sci* **135**, 229–240 (2013).
56. Musil, L. S., Cunningham, B. A., Edelman, G. M. & Goodenough, D. A. Differential phosphorylation of the gap junction protein connexin 43 in junctional communication-competent and -deficient cell lines. *J Cell Biol* **111**, 2077–2088 (1990).
57. Sosinsky, G. E. *et al.* The C-terminus of connexin43 adopts different conformations in the Golgi and gap junction as detected with structure-specific antibodies. *Biochem J* **408**, 375–385, doi: 10.1042/BJ20070550 (2007).
58. Yan, H. H. N., Mruk, D. D., Wong, E. W. P., Lee, W. M. & Cheng, C. Y. An autocrine axis in the testis that coordinates spermiation and blood-testis barrier restructuring during spermatogenesis. *Proc Natl Acad Sci USA* **105**, 8950–8955 (2008).
59. Evans, W. & Martin, P. Gap junctions: structure and function (Review). *Mol Membr Biol* **19**, 121–136 (2002).
60. Cheng, C. Y. *et al.* Regulation of blood-testis barrier dynamics by desmosome, gap junction, hemidesmosome and polarity proteins: An unexpected turn of events. *Spermatogenesis* **1**, 105–115 (2011).
61. Zhang, S. S. & Shaw, R. M. Multilayered regulation of cardiac ion channels. *Biochim Biophys Acta* **1833**, 876–885, doi: 10.1016/j.bbamcr.2012.10.020 (2013).
62. Nielsen, M. S. *et al.* Gap junctions. *Comprehensive Physiology* **2**, 1981–2035, doi: 10.1002/cphy.c110051 (2012).
63. Upham, B. L. Role of integrative signaling through gap junctions in toxicology. *Current protocols in toxicology/editorial board, Mahin D. Maines Chapter 2, Unit2*, **18**, doi: 10.1002/0471140856.tx0218s47 (2011).
64. Citi, S., Guerrero, D., Spadaro, D. & Shah, J. Epithelial junctions and Rho family GTPases: the zonular signalosome. *Small GTPases* **5**, 1–15, doi: 10.4161/21541248.2014.973760 (2014).
65. Clauss, S. B., Walker, D. L., Kirby, M. L., Schimmel, D. & Lo, C. W. Patterning of coronary arteries in wildtype and connexin43 knockout mice. *Dev Dyn* **235**, 2786–2794 (2006).
66. Reaume, A. G. *et al.* Cardiac malformation in neonatal mice lacking connexin43. *Science* **267**, 1831–1834 (1995).
67. Li, W. E. *et al.* An essential role for connexin43 gap junctions in mouse coronary artery development. *Development* **129**, 2031–2042 (2002).
68. Walker, D. L., Vacha, S. J., Kirby, M. L. & Lo, C. W. Connexin43 deficiency causes dysregulation of coronary vasculogenesis. *Dev Biol* **284**, 479–498 (2005).
69. Francis, R. *et al.* Connexin43 modulates cell polarity and directional cell migration by regulating microtubule dynamics. *PLoS One* **6**, e26379, doi: 10.1371/journal.pone.0026379 (2011).

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Author Contributions

C.Y.C. designed research; N.L., H.C. and C.Y.C. performed research; D.D.M., C.K.C.W., W.M.L. and C.Y.C. contributed new reagents/analytic tools; N.L. and C.Y.C. analyzed data; N.L. and C.Y.C. performed the *in vivo* animal experiments; N.L. and C.Y.C. prepared all figures. C.Y.C. wrote the paper; D.D.M. and C.Y.C. revised the manuscript. All authors reviewed the manuscript.

Additional Information

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