

1 **Cocrystal Engineering of Itraconazole with Suberic acid via Rotary Evaporation and**
2 **Spray Drying**

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17 **ABSTRACT**

18 Cocrystallization represents an emerging approach to tackle the issues associated with
19 pharmaceutical product performance and processing, owing to its capability of modifying
20 a variety of physicochemical properties. In this study, we sought to modify the crystal form
21 of itraconazole (ITZ) with suberic acid (SUB) via rapid solvent removal methods, namely
22 rotary evaporation and spray drying. A phase pure ITZ-SUB cocrystal, which could not be
23 obtained by traditional cocrystallization methods, was successfully prepared by rotary
24 evaporation. The new cocrystal was confirmed by powder X-ray diffraction, differential
25 scanning calorimetry, and Fourier-transform infrared spectroscopy. Spray drying was
26 further employed for particle engineering of ITZ-SUB to achieve optimal pulmonary
27 delivery. By manipulating the critical processing parameters, inhalable ITZ-SUB
28 agglomerates with mass median aerodynamic diameter of $2.56 \pm 2.27 \mu\text{m}$ and fine particle
29 fraction of 64.10% w/w were reproducibly prepared. The inhalable powders contained
30 mainly co-amorphous ITZ-SUB, while a small portion of cocrystals still exists. Compared
31 with the raw ITZ, the intrinsic dissolution rate of ITZ-SUB cocrystal was ~39 times faster,
32 and a significantly larger fraction of ITZ-SUB agglomerates were dissolved after 180 min
33 of the test. Besides, both products remained stable after 1-month storage at 60 °C.

34

35 **Keywords:** Itraconazole, Cocrystal, Cocrystal agglomerate, Spray drying, Dry powder
36 inhaler formulation, Aerosol performance, Dissolution

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38

39 1. Introduction

40 Recent advances in cocrystal engineering have ignited mounting interests in the
41 development of next-generation formulations for improved drug delivery. Its promise in
42 tuning the physicochemical properties of problematic drugs, including solubility,
43 dissolution performance, hygroscopicity, and compressibility has been proven and well-
44 documented, thus offering vast opportunities for tailored therapy according to the clinical
45 needs of individual patients.^{1,2} Other merits pertaining to cocrystallization involve its high-
46 level applicability for a wide array of APIs, both weakly ionizable and neutral compounds,
47 which have either limited or no capacity for salt formation.^{3,4}

48 The successful pharmaceutical cocrystal fabrication lies in both prudent selection
49 of crystal formers and effective synthetic approaches. Kinetically driven processes, namely
50 rapid solvent removal via rotary evaporation and spray drying, have been shown to be
51 effective for the preparation of elusive cocrystals, which could not be obtained by
52 conventional methods, such as solid state grinding, slurring, and slow evaporation.^{5,6}
53 Despite the efficiency and ease-of-use of rotary evaporation in initial cocrystal screening
54 stage, its application in mass production is severely hindered since it is a batch process
55 with limited scalability. In contrast, spray drying which produces dry powder through rapid
56 drying of a liquid or slurry in an environment of hot air⁷, is a well-established processing
57 technology for particle engineering and industrial scale-up. Large batches of cocrystal
58 formulations with consistent quality can be continuously manufactured with high
59 production efficiency.⁷ Notably, it also allows the optimization of pharmaceutical
60 properties and aerosolization attributes (e.g., size, density, and morphology), particularly
61 for pulmonary drug administration, through modifying critical processing parameters, such
62 as feed solid concentration and gas atomization flow rate.⁸⁻¹⁰ Although spray drying holds
63 the promise for efficient development of inhalable cocrystal formulations, there are only a
64 few existing studies which probed into such emerging application.

65 Both the choice of formulation compositions and the design of particles with good
66 dispersibility are critical to ensure adequate lung deposition. The particle aerodynamic
67 diameter (d_A) is one of the most important parameters that impacts particle deposition into
68 the lung.¹³ It is defined as the diameter of a unit-density spherical particle, which gives the

69 same settling velocity as the non-spherical particle of interest, and as expressed in eqn. (1):

$$70 \quad d_A = d_V \sqrt{\frac{\rho}{\chi \rho_0}} \quad \text{Eqn. (1)}$$

71 where d_V is the equivalent volume diameter of the particle of interest, ρ is the particle
72 density, ρ_0 is the reference density ($1 \text{ g}\cdot\text{cm}^{-3}$), and χ is the dynamic shape factor. In general,
73 aerosols with d_A ranging from 1 to $5 \mu\text{m}$ are suitable for pulmonary deposition.¹⁴ Particles
74 with $d_A > 5 \mu\text{m}$ mostly deposit in the upper respiratory tract by inertial impaction, while
75 particles with $d_A < 1 \mu\text{m}$ tend to remain air-borne in the airways and are eventually exhaled
76 out during the normal breathing cycle. Nevertheless, particles with d_A ranging 1-3 μm
77 penetrate into the deep lung region better, thus they are recognized as the most optimal
78 fraction for delivery to the alveoli.¹⁵

79 Itraconazole (ITZ, Figure 1) is a synthetic triazole antifungal drug commercially
80 available in its amorphous form, Sporanox[®].¹⁶ The use of oral ITZ has been approved by
81 U.S. FDA since 1992 and is indicated for blastomycosis, histoplasmosis, aspergillosis and
82 onychomycosis.¹⁶ Of particular merit, it is strongly recommended by the Infectious
83 Diseases Society of America (IDSA) for the prophylaxis of invasive pulmonary
84 aspergillosis (IPA), a severe pulmonary infection of the *Aspergillus species* with high
85 morbidity and mortality rates (30-90%).^{17, 18} Through drug repositioning, it is also found
86 to be effective against several types of carcinoma, especially non-small-cell lung cancer.¹⁹⁻
87 ²¹ Nonetheless, as a Class II drug in the Biopharmaceutics Classification System (BCS)
88 that is practically insoluble in water, ITZ requires high oral dose (normally 200 mg daily)
89 to achieve adequate and sustained concentration in the lungs, causing unintended systemic
90 toxicities. Besides, the pharmacokinetic variability of ITZ may hamper its therapeutic
91 efficacy.²² Recent studies for improving the therapeutic outcomes of ITZ includes separate
92 efforts for developing inhalable ITZ powder formulations for targeted pulmonary
93 delivery,²³⁻²⁵ and for ameliorating its solubility through various crystal engineering
94 approaches, such as salt formation and cocrystallization.²⁶⁻²⁸

95 In the context of crystal engineering, Shevchenko et al. reported that cocrystals of
96 ITZ with C2–C7 aliphatic dicarboxylic acids were obtained from a combination of slow
97 evaporation using a 1:1 mixture of tetrahydrofuran with chloroform and solvent-assisted

98 milling using acetone. However, they did not evaluate the solubility and dissolution
99 performance of the resulting cocrystals. Interestingly, attempts of preparing ITZ cocrystals
100 with suberic acid, azelaic acid and sebacic acid (C8-C10) had failed via the same method,
101 despite their structural resemblance with other shorter-chain acids.²⁷ It is not uncommon
102 that cocrystallization with longer-chain acids usually fails or exhibits low efficiency via
103 conventional methods, where the carbon number 8 appeared to be the threshold of
104 successful cocrystallization.^{29, 30} We herein conjecture that such phenomenon may be
105 ascribed to the thermodynamic instability or the relatively high activation energy for
106 cocrystal formations. Both can be overcome by kinetic entrapment through fast
107 crystallization from a solution. Thus, the goals of this work were to: (1) prepare and
108 characterize a fast dissolving ITZ cocrystal with long-chain acid, i.e., suberic acid (SUB),
109 which could not be prepared previously by solvent assisted milling method, by rapid
110 solvent removal; and (2) investigate the potential of integrated crystal and particle
111 engineering through one-step spray drying process for continuous fabrication of a dry
112 powder inhalation (DPI) formulation of ITZ-SUB cocrystal. Different spray drying
113 attributes, including feed solid concentrations and gas atomization flow rates, were used to
114 prepare the ITZ-SUB powders with desired aerosolization performance for pulmonary
115 delivery.

116 **2. Experimental Section**

117 **2.1. Materials**

118 Itraconazole (ITZ) (purity $\geq 98.5\%$) was supplied from Yick Vic Chemicals &
119 Pharmaceuticals Ltd (Hong Kong, China). Suberic acid (SUB) was purchased from Alfa
120 Aesar (Ward Hill, MA, USA). Chloroform (CFM), ethanol (EtOH), tetrahydrofuran (THF)
121 and acetonitrile (ACN) of analytical grade were obtained from Merck KGaA (Darmstadt,
122 Germany). Water was purified through a Direct-Q water purifier (Water Corp., Milford,
123 MA). Ammonium acetate was purchased from Sigma Aldrich (St Louis, Mo, USA) and
124 analytical grade acetic acid was obtained from BDH Laboratory Supplies (Dorset,
125 England). Hydrochloric acid 37% was provided by RCI Labscan Ltd (Bangkok, Thailand).

126 **2.2. Preparation of Cocrystals by Rotary Evaporation**

127 Equimolar amounts (0.425 mmol) of ITZ (100 mg) and SUB (24.69 mg) were
128 dissolved in a 100 mL mixture of EtOH and CFM (4:1 v/v). Subsequently, the organic
129 solvent was removed using a rotary evaporator (Büchi, Germany) under a vacuum (90 mm
130 Hg) achieved by a vacuum pump (Millipore, USA) with the rotary flask rotating at a speed
131 of 40 rpm while being immersed in a water bath at 60 °C, resulting in an evaporation speed
132 of around 6.9mL/min. The resulting product was dried in an oven at 80 °C overnight to
133 remove residual solvent and gently grounded to a fine powder for further analysis. All
134 samples were stored in parafilm-wrapped plastic vials after drying in a double-zipped bag
135 to avoid significant moisture sorption.

136 **2.3. Determination of the Solubility of ITZ and SUB in Organic Solvents**

137 Excess amounts of ITZ and SUB were separately added in screw-capped test tubes
138 with 5 mL of different organic solvents and shaken for 72 hours. To make the
139 supersaturated suspension of ITZ and SUB in the mixture of EtOH and CFM (4:1 v/v), an
140 excess amount of ITZ was dissolved in 1 mL CFM initially, followed by the addition of 2
141 mL EtOH. Subsequently, an excess amount of SUB was poured in. Finally, another 2 mL
142 EtOH was added. Samples were withdrawn and filtered through 0.45 µm nylon syringe
143 filters (Membrane Solutions, Hong Kong), followed by dilution to appropriate
144 concentrations for HPLC assay (see Section 2.3).

145 **2.4. High Performance Liquid Chromatography (HPLC)**

146 The concentrations of ITZ and SUB in the solubility study were determined using
147 an HPLC equipped with a diode array detector (Agilent 1200 series, Agilent Technologies,
148 USA) and an Agilent Zorbax Eclipse Plus C18 column (5 µm, 250 mm× 4.6 mm). For ITZ,
149 the mobile phase employed was a mixture of 60% acetonitrile (ACN) and 40% 10 mM
150 ammonium acetate buffer (pH=5.7).³¹ For SUB, the mobile phase consisted of 90% ACN
151 and 10% ultrapure water. A 10 µL aliquot of each sample solution was injected and ran at
152 a flow rate of 1 mL/min at room temperature. The detection wavelength was 261 nm and
153 210 nm for ITZ and SUB, respectively.

154 **2.5. Differential Scanning Calorimetry (DSC)**

155 Thermograms were generated through a differential scanning calorimeter (Q1000,
156 TA, DE, USA), using nitrogen as the purge gas at a flow rate of 20 mL/min. Pure indium
157 was used prior to the analysis to calibrate the enthalpy and cell constant. Accurately
158 weighed samples (3–5 mg) were hermetically sealed in aluminum pans, with a pinhole-
159 vented lid if required, and heated from 30 °C to 250 °C at a scanning rate of 10 °C/min.
160 The heating rate of 2 °C/min was applied for constructing the temperature-composition
161 phase diagram while other conditions were unchanged.

162 **2.6. Powder X-Ray Diffraction (PXRD)**

163 The X-ray powder diffraction data were collected using a MiniFlex 600
164 diffractometer (Rigaku, Tokyo, Japan) equipped with Cu-K α radiation (40kV and 15mA).
165 Samples were uniformly packed in a glass capillary and scanned over the 2 θ interval of 3°
166 to 50° at 0.02° step size with scanning speed of 4° per minute.

167 **2.7. Fourier-Transform Infrared (FTIR) Spectroscopy**

168 The FTIR spectra were obtained with an FTIR spectrophotometer (ALPHA, Bruker,
169 Germany) in diffuse reflectance mode. A trace amount of the sample was gently mixed
170 with potassium bromide (KBr) using a marble pestle and mortar and then compressed into
171 a thin disc under two tons of force using Mini-Pellet Press (Specac Limited, UK). A total
172 of 16 scans were performed in the range of 4,000 cm⁻¹ to 500 cm⁻¹ at a resolution of 4 cm⁻¹
173 ¹ for each sample.

174 **2.8. Spray Drying**

175 Equimolar amounts (0.425 mmol) of ITZ (100 mg) and SUB (24.69 mg) were
176 dissolved in 100 mL mixture of EtOH and CFM (4:1 v/v). Subsequently, the solution was
177 spray-dried using a laboratory scale spray dryer with a high-performance cyclone in suction
178 mode and closed loop configuration (Mini Spray Dryer B-290, Dehumidifier B-296 and
179 Inert Loop B-295; Büchi Labortechnik, Flawil, Switzerland) with nitrogen as the drying
180 gas. Various solute concentrations and compressed gas atomization flow rate were tested
181 (Table 1), while other operating parameters were fixed: inlet temperature of 65 °C, rate of
182 aspiration at 100% (approx. 35 m³/h), solution feed rate of 1.5 mL/min. The outlet
183 temperature was approximately 50 °C.

184 **2.9. *In Vitro* Aerosol Performance**

185 The *in vitro* aerosol performance of spray-dried formulations was evaluated using
186 a Next Generation Impactor (NGI; Copley Scientific Limited, Nottingham, UK). Prior to
187 dispersion, a thin layer of silicon grease (Slipicone; DC Products, Waverley, VIC, Australia)
188 was sprayed onto the impactor stages to minimize particle bounce. Approximately 5 mg of
189 each powder was loaded in a Size 3 hydroxypropyl methylcellulose (HPMC) capsule
190 (Capsugel, West Ryde, NSW, Australia), which was then placed in a Breezhaler (Novartis
191 Pharmaceuticals, Hong Kong). Subsequently, the powders were dispersed at a flow rate of
192 90 mL/min for 2.7 s. After dispersion, the powders remained in the capsule, deposited at
193 the throat, as well as those on the different NGI stages were collected by rinsing with
194 analytical grade ACN, while the inhaler and the adaptor were washed with ultrapure water,
195 followed by dilution with ACN to a final ACN/water (60:40 v/v) solution. Then, the
196 solutions were filtered by 0.45 µm nylon syringe filters and assayed by HPLC. The
197 recovered dose was defined as the sum of powder mass assayed on all the parts in a single
198 run. The emitted fraction (EF) referred to the fraction of powder that exited the inhaler to
199 the total recovered dose. The fine particle fraction (FPF) was the mass fraction of the
200 particles < 5 µm with respect to the recovered dose. The mass median aerodynamic
201 diameter (MMAD) and the geometric standard deviation (GSD) were obtained by a linear
202 fit of cumulative mass vs. log cut-off diameter according to the British Pharmacopoeia
203 (2019).¹³

204 **2.10. Scanning Electron Microscopy (SEM)**

205 The particle morphology of the samples was observed by field emission scanning
206 electron microscopy (Hitachi S-4800 FEG, Hitachi, Tokyo, Japan). The powders were
207 sprinkled onto carbon adhesive tape mounted on SEM stubs and sputter coated with
208 approximately 11 nm gold-palladium alloy in two cycles to create a conductive layer and
209 avoid overheating.

210 **2.11. Dissolution Test**

211 The dissolution test was conducted in triplicates using the Copley Dissolution
212 Tester DIS8000 (Copley Scientific Limited, Nottingham, UK). For ITZ-SUB cocrystals
213 prepared by rotary evaporation, 100 mg ITZ powders and 124.7 mg ITZ-SUB cocrystal

214 powders with particle sizes ranging from 125-180 μm (VWR International, New York,
215 USA) were directly poured into 900 mL of 0.1N HCl solution, with a paddle rotated at 100
216 rpm, for a period of 180 min at 37 °C. At designated time points of 5, 15, 30, 45, 60, 90,
217 120 and 180 min, 5 mL of the dissolution medium was withdrawn and replaced with an
218 equal volume of fresh medium. The sample solution was filtered through 0.45 μm nylon
219 syringe filters and assayed for drug content by HPLC method as described in section 2.3.

220 To investigate the dissolution profile of spray dried ITZ-SUB powders on a DPI
221 respirable fraction, the powders with aerodynamic diameters $< 5 \mu\text{m}$ were collected using
222 a Fast Screening Impactor (FSI; MSP Corporation, Shoreview, USA).²² Briefly, 10 mg
223 spray dried ITZ-SUB powders were loaded in a Size 3 HPMC capsule and dispersed at 100
224 L/min flow rate for 2.4 s. After dispersion, powders with aerodynamic diameters $< 5 \mu\text{m}$
225 were deposited on a glass fiber filter (ADVANTEC; Toyo Roshi Kaisha, Ltd., Japan), then
226 transferred to a watch glass placed in the bottom of the vessel containing 400 mL 0.1 N
227 HCL with 0.3% sodium dodecyl sulfate (SDS),^{25, 32} with a paddle stirring at 75 rpm for a
228 period of 180 min at 37 °C. For raw ITZ powders, the amount equivalent to the ITZ mass
229 of the spray-dried ITZ-SUB powders obtained from FSI was directly weighted onto the
230 glass fiber filter attached to the watch glass. The sampling and assay procedures were the
231 same as the previous dissolution test.

232 **2.12. Dynamic Vapor Sorption (DVS)**

233 Water sorption isotherms of the samples were obtained using an automated vapor
234 sorption analyzer (Intrinsic DVS, Surface Measurement Systems Ltd., Allentown, PA) at
235 25 °C and a nitrogen flow of 50 mL/min. Each sample (6-10 mg) was first purged with dry
236 nitrogen until a constant weight was obtained. Then, the sample was exposed to a series of
237 relative humidities (RH) from 0% to 95% with a step size of 5% RH. The RH was changed
238 to the next target value once the equilibrium was reached, where either dm/dt was $\leq 0.002\%$
239 with a minimum equilibration time of 0.5 h or maximum equilibration time of 6 h at each
240 specific RH.

241 **2.13. Thermal Stress Study**

242 Raw ITZ, raw SUB, ITZ-SUB cocrystal and spray dried ITZ-SUB powders were

243 put in screw-capped glass bottles separately and then placed in 60 °C oven. After 1 month,
244 samples were collected for PXRD and DSC analysis.

245 **2.14. Statistical Analysis**

246 Two-sample *t*-test was employed for data analysis. A p-value less than 0.05 was
247 considered as statistically significant.

248

249 **3. Results and Discussion**

250 **3.1. Cocrystallization of Itraconazole with Suberic Acid**

251 In a previous study, the cocrystal formation potential of ITZ with a series of
252 structurally similar aliphatic dicarboxylic acids has been established.²⁶ ITZ cocrystals with
253 C2-C7 dicarboxylic acids could all be obtained by the means of either slow evaporation of
254 a solution in 1:1 THF and CFM mixture or solvent-assisted milling using acetone. However,
255 ITZ failed to cocrystallize with SUB and other longer chain acids. Hence, C7 was believed
256 to be the maximum carbon atom number of the aliphatic chain for successful
257 cocrystallization reaction between ITZ and a dicarboxylic acid.²⁷ Indeed, only
258 characteristic peaks of ITZ and SUB were found in the PXRD pattern of the samples
259 prepared from the slow evaporation method (Figure 2), indicating the failure of ITZ-SUB
260 cocrystal formation. This was attributed to either the inherent thermodynamically unstable
261 nature of ITZ-SUB or a lower molecular mobility of the longer aliphatic chain.³⁰
262 Encouraged by our earlier success,⁵ we sought to prepare this elusive ITZ-SUB cocrystal
263 using a rapid solvent removal method, i.e., rotary evaporation.

264 Although solutions of incongruently saturating systems can generate pure
265 cocrystals by rapid solvent evaporation, it has been suggested by ternary solubility phase
266 diagram that the solubility difference between the cocrystal formers should not be too large
267 in the chosen solvent system because this would significantly alter the propensity of
268 cocrystal formation.^{7, 11, 33} Among the tested three pure organic solvents, SUB is much
269 more soluble in THF and EtOH (2728.35 mmol/L and 498.81 mmol/L respectively) than
270 ITZ (37.84 mmol/L and 0.34 mmol/L respectively), while CFM solubility of ITZ (634.83
271 mmol/L) is much higher than SUB (6.20 mmol/L) (Table 2). While there is no single

272 solvent system that gives congruent solubility between ITZ and SUB, the EtOH and CFM
273 mixture in 4:1 v/v ratio did yield similar solubilities of ITZ and SUB. The solubility of ITZ
274 detected was 16.07 mmol/L and that of SUB was 16.35 mmol/L, which were not
275 significantly different ($p = 0.61$).

276 In this study, the cocrystal of ITZ and SUB in 1:1 stoichiometric ratio was
277 successfully obtained from a mixture of EtOH and CFM (4:1) using rotary evaporation. To
278 the best of our knowledge, this is the first 1:1 cocrystal of ITZ with a dicarboxylic acid but
279 without any solvent or water in the crystal lattice. The PXRD pattern of the prepared sample
280 exhibits a series of distinct diffraction peaks ($2\theta = 3.99^\circ, 6.19^\circ, 9.23^\circ, 12.96^\circ, 18.62^\circ$ and
281 21.71°) while characteristic peaks corresponding to ITZ ($2\theta = 14.50^\circ, 20.42^\circ$ and 23.54°)
282 and SUB ($2\theta = 9.9^\circ, 21.34^\circ$ and 24.77°) were absent (Figure 2). Those unique diffraction
283 peaks of the prepared sample suggested the formation of a new crystalline phase, likely the
284 ITZ-SUB cocrystal, whereas the absence of characteristic peaks of ITZ and SUB denotes
285 the absence of ITZ and SUB in the sample.

286 Regarding thermal property, the DSC thermogram of the ITZ-SUB cocrystal
287 showed a sharp endothermic peak at 135.7°C , which is lower than the melting points of
288 ITZ (167.7°C) and SUB (141.8°C) (Figure 3). Interestingly, the thermal curve of SUB
289 exhibited another smaller endothermic peak adjacent to the actual melting point, consistent
290 with the previous reports.^{34, 35} The enthalpy change of fusion (ΔH_f) of the cocrystal (89.05
291 kJ/mol) was significantly higher than those of ITZ (79.70 kJ/mol) and SUB (49.94 kJ/mol),
292 demonstrating a stronger crystal lattice upon the cocrystal formation. To further verify the
293 formation of a new cocrystal phase and confirm its stoichiometry, a temperature-
294 composition phase diagram of the ITZ-SUB system was constructed using binary mixtures
295 of SUB with varying ITZ mole fractions from 0 to 1, through DSC analysis (Figure 4). The
296 binary mixtures were generated by rotary evaporation which was the same method as the
297 cocrystal preparation. The phase diagram showed a typical shape of a binary cocrystal
298 system, where three local melting point maxima are separated by two eutectics. It revealed
299 a congruent melting temperature at 50:50 molar composition of either cocrystal former,
300 confirming a 1:1 stoichiometry. Two eutectic points were observed at 0.2 and 0.54 ITZ

301 mole fractions with melting temperatures at 121.5 °C and 130.2 °C, respectively (Figure
302 4).

303 Fourier-transform infrared spectroscopy was conducted to gain an insight into the
304 intermolecular interactions between ITZ and SUB. An outstanding observation when
305 comparing these FTIR spectra is that the C=O stretching vibration of SUB at 1695 cm⁻¹
306 wavelength dramatically shifted to a higher wavelength of 1711 cm⁻¹ in the ITZ-SUB
307 spectrum (Figure 5). This can happen if the -OH of the carboxyl group became involved in
308 intermolecular hydrogen bond formation, thus decreasing the intramolecular hydrogen
309 bonding strength between C=O and -OH.³⁶ In addition, the broad peak corresponding to
310 the O-H group in SUB, covering a wavelength range from 3300 cm⁻¹ to 3750 cm⁻¹, became
311 narrower at 3425 cm⁻¹ in the ITZ-SUB spectrum. Besides, ITZ-SUB cocrystal shared
312 highly similar PXRD pattern with the previously reported ITZ-succinic acid cocrystal in
313 the 2θ range of 3° to 10°. ²⁸ These results imply that the carboxyl groups of SUB and the
314 nitrogen atoms to the 1,2,4-triazole group of ITZ participated in the formation of hydrogen
315 bond, which is an interaction observed in the ITZ cocrystal with succinic acid.²⁸

316 Here, rapid rotary evaporation technique was again proven to be effective in the
317 fabrication of an elusive cocrystal. Suspending ITZ-SUB in water led to dissociation and
318 formation of ITZ. Thus, the formation of the ITZ-SUB cocrystal is likely realized by kinetic
319 entrapment as suggested by Ostwald's rule of stages³⁷, where thermodynamically unstable
320 phases appear first during the crystallization from solutions. However, the cocrystal
321 powders prepared by rotary evaporation technique were not suitable for pulmonary drug
322 delivery as the particle size was larger than 100 μm (Figure S1). Hence, spray drying was
323 employed to produce smaller ITZ-SUB particles.

324 **3.2.Spray Dried ITZ-SUB powders**

325 Having proven the feasibility of preparing the ITZ-SUB cocrystal by fast solvent
326 removal through rotary evaporation, spray drying was further probed for the feasibility of
327 producing inhalable ITZ-SUB dry powder formulations with suitable aerosol performance.
328 Spray drying is a versatile technique that allows continuous production of dry powders,
329 which can fit diverse purposes by adjusting the feed solution/suspension composition and
330 various process parameters. In this study, feed solute concentration and compressed gas

331 atomization flow rate were selected for process optimization because of their proven
332 impact on the particle size, where particle size generally increases with increasing solute
333 concentration and/or decreasing compressed gas atomization flow rate.^{8-10, 38}

334 Among the six tested spray-dried formulations (Table 1), SD3 appeared to be ideal
335 for pulmonary delivery as it exhibited the most desirable MMAD and GSD (2.56 ± 2.27
336 μm) as well as the highest FPF (64.1% w/w), which was much higher than that of most
337 commercial products (around 30% w/w).³⁹ As mentioned above, aerodynamic diameters
338 of 1-5 μm are widely accepted for drug delivery to the lung, but the range from 1 to 3 μm
339 is preferred for reaching the deep lung region.^{14, 15} This size range correlates well with good
340 clinical response to local treatment of lung diseases, such as IPA, and favors the penetration
341 of the drug into the capillary-rich alveolar region for systemic treatment. Additionally,
342 since SD3 can be delivered to deep lung without using a carrier, blend uniformity problems
343 are eliminated.⁴⁰ A carrier-free formulation also allows the delivery of high dose ITZ to the
344 lung since the carrier typically occupies 67.5% of the formulation mass.⁴¹

345 Both solute concentration of the feed solution and the compressed gas atomization
346 flow rate did affect the d_A of the resulted formulations. The primary particle size decreased
347 with decreasing solute concentration and increasing of the gas atomization flow rate
348 (Figure S5), as expected.^{8-10, 38} However, smaller primary particles are more likely to
349 aggregate. Therefore, a lower solute concentration and a higher gas flow rate did not
350 necessarily generate a formulation with smaller d_A upon aerosolization.

351 The NGI dispersion plot revealed that 68% of the SD3 powder deposited on Stages
352 3-5, where the aerodynamic diameters fell within 3.61-0.76 μm (Figure 6). However, SD3
353 powder consisted of agglomerates of smooth spherical particles with diameter around 1
354 μm (Figure 7). The particles deposited on the different stages of the NGI mainly differ in
355 the extent of agglomeration instead of primary particle size (Figure S7). Upon inhalation,
356 the spray-dried ITZ-SUB cohesive powder formed smaller agglomerates, favoring the
357 deposition in the lower lung region. The primary submicron particles may potentially
358 penetrate into the deep lung after the agglomerates have settled and de-agglomerated.
359 Besides, the electrostatic nature of these agglomerates may lead to a more efficient
360 deposition via the electrostatic attraction between the particle and the induced opposite

361 charge on the wall of the lung.⁴² Furthermore, in comparison with particles of the same
362 size, the agglomerates usually has a better bioavailability. This is due to the larger surface
363 area of its rough surface, thus enhancing dissolution rate, delaying lung clearance, and
364 improving systemic absorption.⁴³

365 It is worth noting that besides the most commonly occurring smooth spheres,¹⁰ there
366 are also a few rod-shaped particles present in the spray dried powders, indicating the
367 possibility of crystallized ITZ-SUB. The PXRD pattern of SD3 indicated low crystallinity,
368 but the characteristic peaks of ITZ-SUB at 6.19°, 9.23° and 21.71° were still observed
369 (Figure 2), while other spray dried powders were largely amorphous (Figure S2). The DSC
370 thermogram of all the spray dried samples illustrated the glass transition of the amorphous
371 content at around 50 °C, followed by the evaporation of the residual solvent and a broad
372 crystallization exotherm (Figure S3). Eventually, the recrystallized ITZ-SUB cocrystals
373 melted at 135.7 °C (Figure 3). The FTIR spectra of all the spray dried powders also
374 exhibited similar peaks with ITZ-SUB cocrystals (Figure 5 & S4). Hence, it can be
375 concluded that the spray dried powders contained mainly co-amorphous ITZ-SUB, but a
376 small portion of cocrystals still exists. Albeit the drug particles are generally required to be
377 crystalline due to the stability issue, the amorphous form is considered to be superior for
378 its rapid dissolution and absorption.⁴⁴

379 **3.3. Dissolution Profile**

380 As ITZ belongs to BCS Class II, its therapeutic efficacy is limited by its solubility.
381 It has been shown that the solubility of poorly-water soluble drugs can be improved by
382 cocrystallization, especially when using a water-soluble coformer.^{45, 46} However, direct
383 measurement of ITZ-SUB aqueous solubility was not possible as the cocrystal underwent
384 dissociation when suspended in an aqueous solution owing to the thermodynamically
385 unstable nature of the cocrystals. In the contrary, dissolution test is a useful alternative for
386 solubility measurement. Considering ITZ is commercially available as oral tablet and
387 capsule formulations, we have assessed the dissolution performance of both oral ITZ-SUB
388 cocrystal prepared by rotary evaporation for possible use in oral tablets or capsules and the
389 spray-dried inhalable ITZ-SUB cocrystal agglomerates.

390 The dissolution of crystalline ITZ-SUB prepared by rotary evaporation was tested
391 in a 0.1N HCl solution to stimulate the gastric environment. To minimize the size, size
392 distribution and morphology effects of the particles, only the 125- 180 μm fraction of both
393 raw ITZ and ITZ-SUB cocrystals was used for the test. The dissolution profile of the raw
394 ITZ plateaued at lower than 6%, while ITZ-SUB cocrystals followed first-order kinetics
395 with approximately 80% ITZ dissolved after 180 min (Figure 8). The intrinsic dissolution
396 rate (IDR) can be determined by the equation: $IDR = (dm/dt)_{max}/A$, where $(dm/dt)_{max}$ is the
397 slope of the initial linear region of the cumulative dissolution curve and A is the specific
398 surface area of the dissolution sample.⁴⁷ Although the exact specific surface area of the
399 samples was not measured in this study, the sieved ITZ-SUB cocrystal and raw ITZ were
400 within the same size range (125-180 μm) as well as exhibiting similar particle morphology
401 (Figure S8). Hence, the IDR ratio of ITZ-SUB to ITZ can be roughly estimated by the ratio
402 of the slope, which is 39.2, suggesting roughly 39-fold faster in the rate of dissolution of
403 the ITZ-SUB cocrystal than raw ITZ in 0.1 N HCl. Concerning the fraction of release,
404 significantly more ITZ-SUB cocrystal was dissolved than raw ITZ (66.8% and 2.1%,
405 respectively; $p = 8.2 \times 10^{-5}$) at 90 min, which was the end point of ITZ dissolution test
406 recommended by US FDA.⁴⁸ In addition, the apparent solubility, measured one day after
407 the dissolution test, of 97.1 $\mu\text{g}/\text{mL}$ for ITZ-SUB cocrystal was significantly higher than
408 0.6 $\mu\text{g}/\text{mL}$ for raw ITZ ($p = 5 \times 10^{-8}$).

409 Hence, cocrystallization of ITZ with SUB remarkably improved the dissolution
410 performance and the apparent solubility of ITZ in an acidic medium. This substantial
411 amelioration can be attributed to the high aqueous solubility of SUB.⁴⁶ The dissolution rate
412 of dry powder inhaler formulations is also critical as undissolved particles will be cleared
413 by macrophage phagocytosis in the alveolar region and the mucociliary escalator in the
414 conducting airways.⁴⁴ Considering this, the dissolution protocol of dry powder inhaler
415 formulations must be different from that of conventional solid dosage forms as only
416 particles with MMAD $< 5\mu\text{m}$ could effectively enter the lower airways. Nevertheless, no
417 pharmacopoeia method or universally acceptable protocol has yet been developed for DPI
418 formulation. To this end, an FSI was employed in this study to collect particles with
419 MMAD $< 5\mu\text{m}$.²² Before the dissolution test, the FPF of spray-dried ITZ-SUB powders
420 using FSI, assayed by HPLC, was 64.6%. Here, we utilized a 0.1 N HCl with 0.3% SDS

421 as the medium in this dissolution test because previous studies have substantiated its close
422 correlation with the *in vivo* study that qualitatively compared different aerosol formulations
423 in the lung.^{24, 25, 49} The presence of SDS is expected to improve the dissolution performance
424 of the raw ITZ by improving wetting, provided no insoluble complex is formed with ITZ.⁵⁰
425 However, raw ITZ still dissolved slowly and only 50% was dissolved after 180 min. On
426 the contrary, the spray dried ITZ-SUB powders with $d_A < 5 \mu\text{m}$ dissolved very rapidly in
427 the initial 15 min and reached a plateau at 80% ITZ fraction of release after 30 min (Figure
428 9). At 180 min, the fraction of ITZ dissolved in ITZ-SUB was significantly higher than raw
429 ITZ ($p = 0.03$). Therefore, the spray dried ITZ-SUB cocrystals substantially enhanced both
430 the dissolution rate as well as the extent of ITZ dissolved, which can be explained by both
431 the amorphous nature and the large specific surface area of the agglomerate structure.
432 Consequently, the spray-dried ITZ-SUB holds promise for pulmonary administration of
433 ITZ.

434 **3.4. Stability Profile**

435 Stability is a key consideration in the development of pharmaceutical products.
436 Generally, relative humidity stress study and thermal stress study are the most common
437 approaches for stability test. As indicated by the DVS results, none of the crystalline
438 materials adsorbed a significant amount of water ($< 0.5\%$) at 95% RH (Figure 10).
439 Therefore, ITZ, SUB and ITZ-SUB cocrystal are all non-hygroscopic. However, both spray
440 dried ITZ and ITZ-SUB exhibited higher moisture sorption but the amounts were not
441 significant as the weight only changed less than 2.5%. Compared with spray dried ITZ
442 which was completely amorphous, spray dried ITZ-SUB powders adsorbed a less amount
443 of water. For the stability under thermal stress, ITZ-SUB prepared by both rotary
444 evaporation and spray drying did not dissociate into individual components at 60 °C for 1
445 month. The characteristic peaks of the cocrystal in the PXRD pattern were still evident and
446 the melting point remained unchanged according to the DSC thermogram (Figure 11).
447 While amorphous form of a chemical compound is generally considered less stable than its
448 crystalline counterpart, the spray dried ITZ-SUB powders did not undergo significant
449 crystallization after 1-month storage at 60 °C according to PXRD pattern (Figure 11d).
450 The residual solvent in the spray-dried ITZ-SUB was removed during storage as indicated

451 by the flat baseline before the melting point in its DSC thermogram. This suggests that
452 residual solvent can be easily removed from the spray-dried ITZ-SUB in commercial
453 manufacturing. Although a crystallization exotherm is absent, a melting event at 135.7 °C
454 corresponding to that of ITZ-SUB crystal is clearly observed. Thus, any crystallization
455 must have occurred over a wide temperature range so that only a broad peak can be
456 expected. It is worth to mention that ITZ in its branded product, Sporanox[®], also exists in
457 an amorphous state.¹⁷ Hence, the stability issue should not hinder the commercialization of
458 spray dried ITZ-SUB.

459

460 **4. Conclusion**

461 In this study, a phase pure ITZ-SUB 1:1 cocrystal was successfully prepared by
462 rapid solvent removal using rotary evaporation, and characterized by a variety of
463 techniques including PXRD, DSC, and FTIR. Despite the markedly improved dissolution
464 performance, the size of the resulting cocrystal restricted its application in the development
465 of inhalable formulations. To this end, spray drying was further employed to fabricate ITZ-
466 SUB agglomerates for superior pulmonary delivery. The spray dried ITZ-SUB powders
467 possessed more desirable size distribution, morphology, and dissolution rate for inhalation
468 in comparison to the parent drug.

469

470 **Acknowledgments**

471 We thank financial supports from The University of Hong Kong (Project number:
472 104004777 and 204600519). We also thank Ms. Roshni Dattani (School of Pharmacy,
473 University College London) for assistance with the dissolution study.

474

475 **Supporting Information**

476 PXRD patterns, DSC profiles, FTIR spectra, SEM images and NGI dispersion data
477 of spray dried ITZ-SUB, SEM images of spray dried ITZ-SUB powders (SD3) deposited
478 on the throat and different stages of the NGI, SEM images of ITZ and ITZ-SUB cocrystal.

479

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Table 1. Aerodynamic size distribution (MMAD, GSD and FPF) of spray-dried formulations under different solute concentrations and compressed gas atomization flow rates.

Formulation	Solute Concentration (mmol/L)	Gas flow (NI/h)	MMAD (μm)	GSD (μm)	FPF (% w/w)
SD1	1.14	742	4.22	3.03	33.71
SD2	1.14	601	4.04	2.75	42.27
SD3	3.41	742	2.56	2.27	64.10
SD4	3.41	601	3.69	2.75	42.49
SD5	5.68	742	5.05	2.92	32.98
SD6	5.68	601	4.49	2.87	37.2

Table 2. Solubilities (mean \pm standard deviation) of ITZ and SUB in selected organic solvents (n = 3).

Sample	Solubility (mmol/L) in:			
	THF	EtOH	CFM	EtOH/CFM (4:1 v/v)
ITZ	37.84 \pm 0.34	0.34 \pm 0.01	634.83 \pm 7.85	16.07 \pm 0.57
SUB	2728.35 \pm 33.32	498.81 \pm 9.83	6.20 \pm 0.32	16.35 \pm 0.66

Figure 1. Chemical structure of (1) ITZ and (2) SUB.

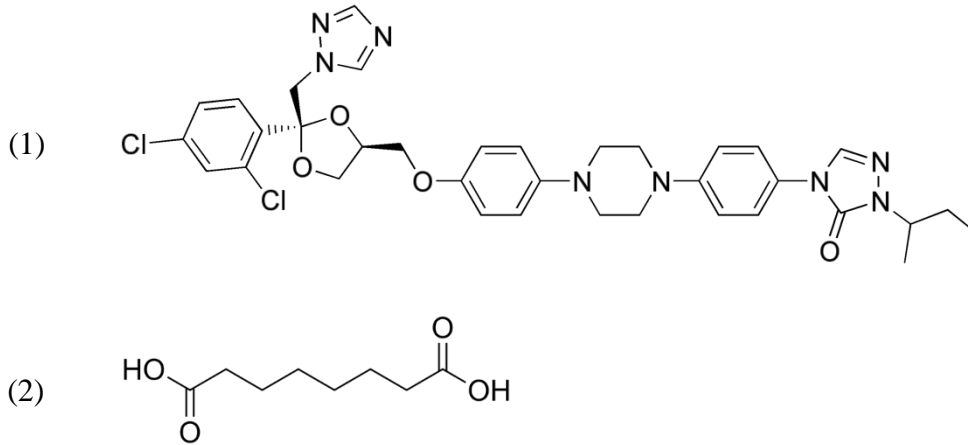


Figure 2. PXRD patterns of ITZ, SUB, Samples obtained from slow evaporation, ITZ-SUB cocrystals and spray dried ITZ-SUB (SD3). * denotes the characteristic peaks of ITZ-SUB cocrystal.

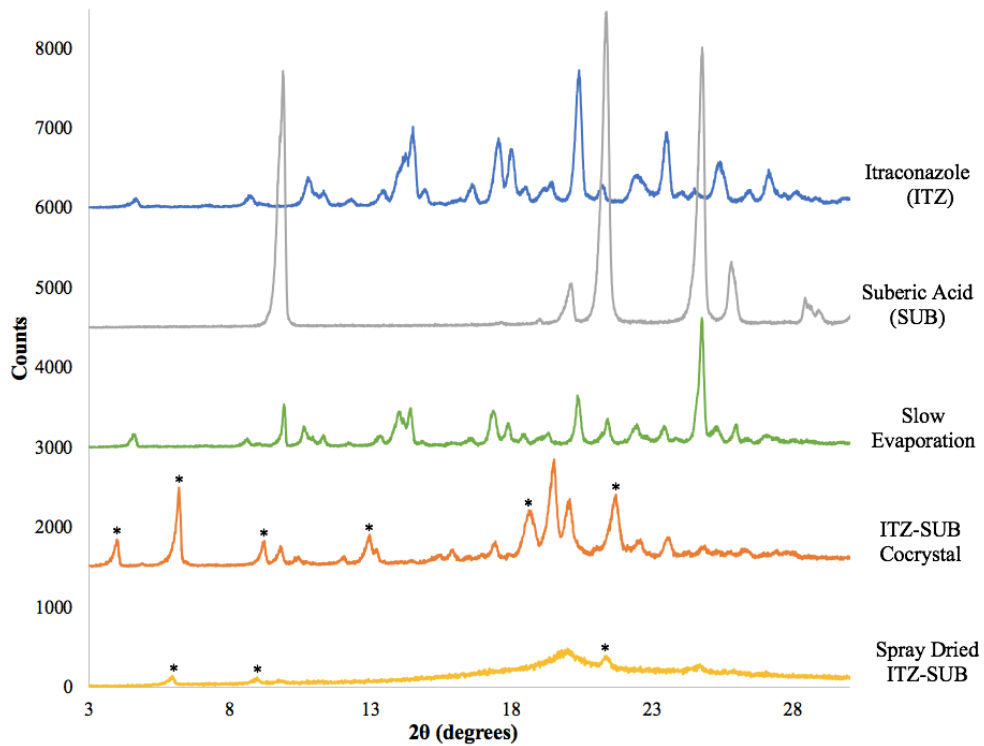


Figure 3. DSC profiles of ITZ, SUB, ITZ-SUB cocrystals and spray dried ITZ-SUB (SD3).

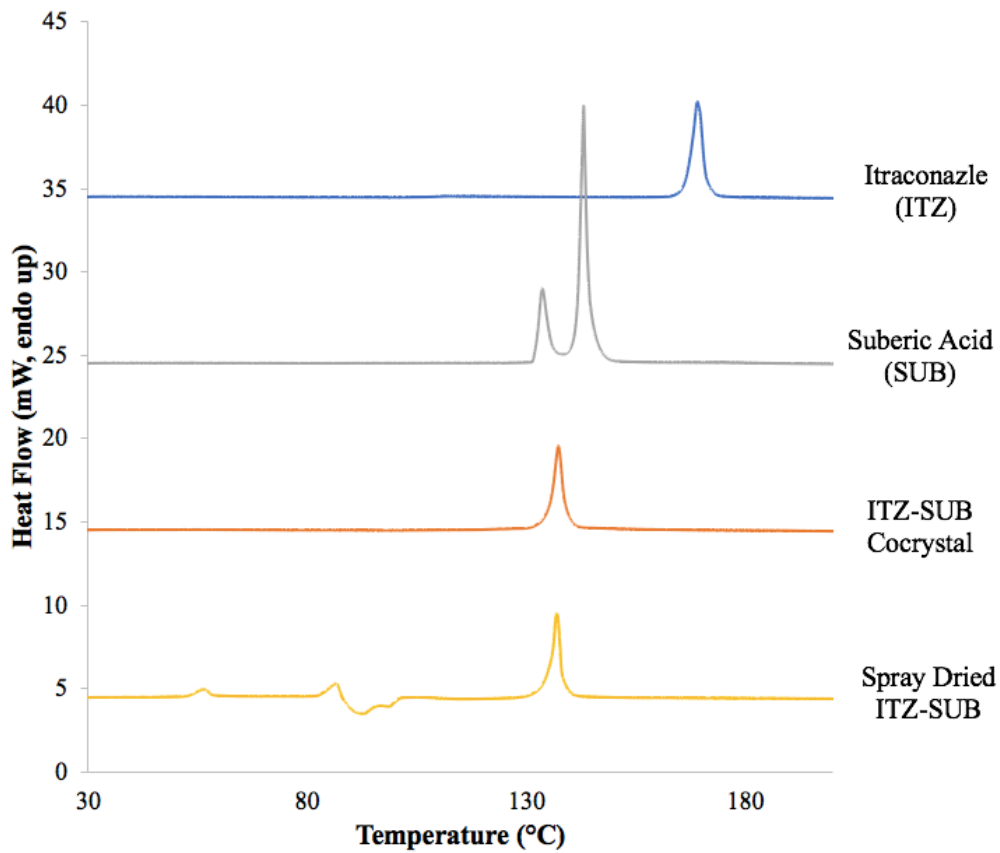


Figure 4. Melting point-composition phase diagram of ITZ-SUB cocrystal.

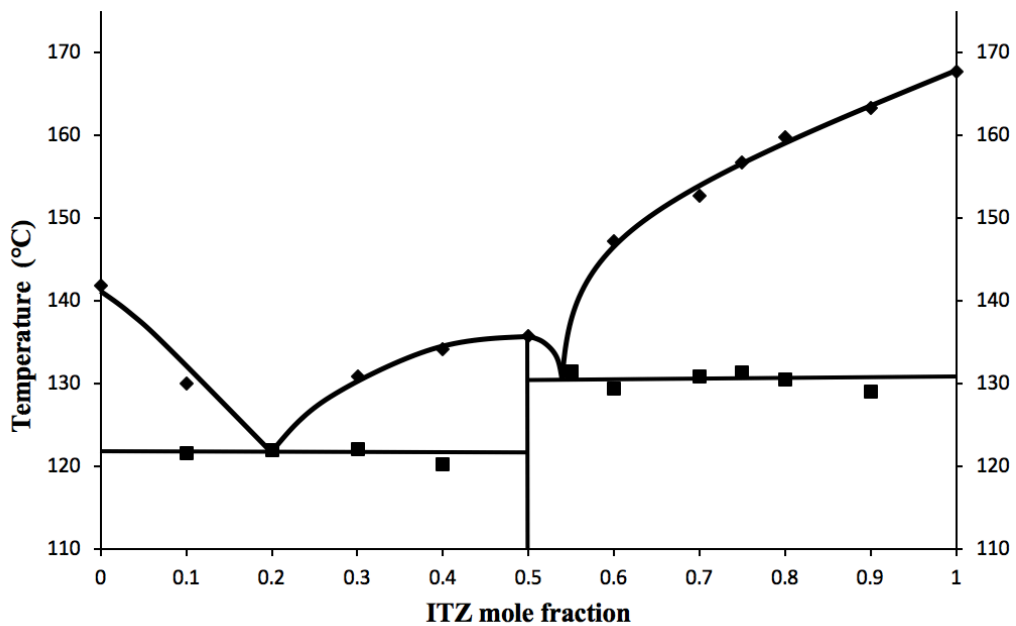


Figure 5. FTIR spectra of ITZ, SUB, ITZ-SUB cocrystal and spray dried ITZ-SUB (SD3).

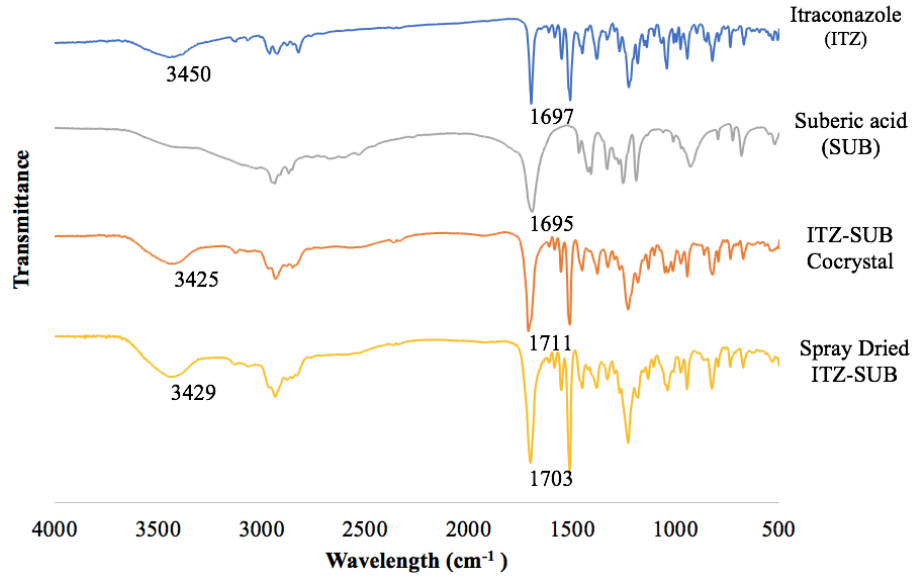


Figure 6. NGI dispersion data of spray dried ITZ-SUB (SD3, n=3). S1–S7 presents impactor stages 1–7, followed by the corresponding upper aerodynamic cut-off diameter in parentheses. MOC is the micro-orifice collector in the NGI.

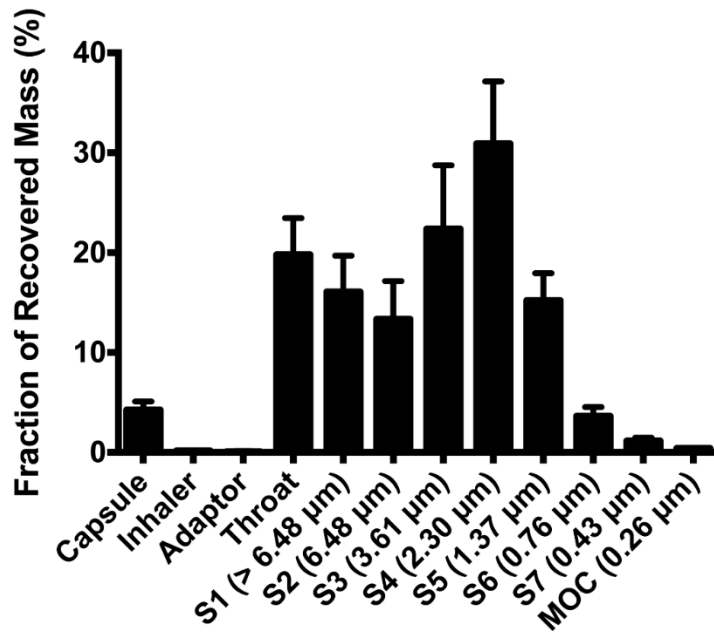


Figure 7. SEM images of spray dried ITZ-SUB (SD3). (a) at 500× magnification; (b) at 5000 × magnification.

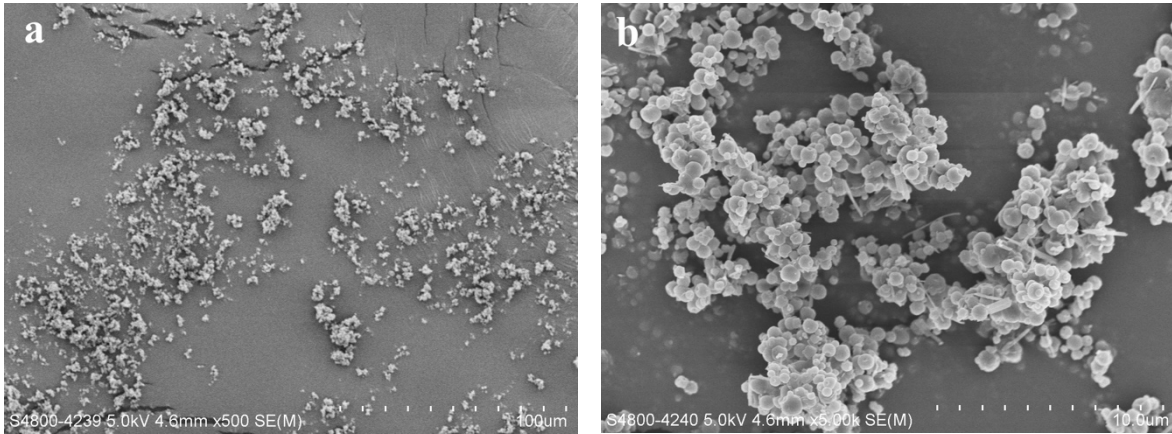


Figure 8. Dissolution profiles of ITZ and ITZ-SUB cocrystal with size ranged 125-180 μm (n=3).

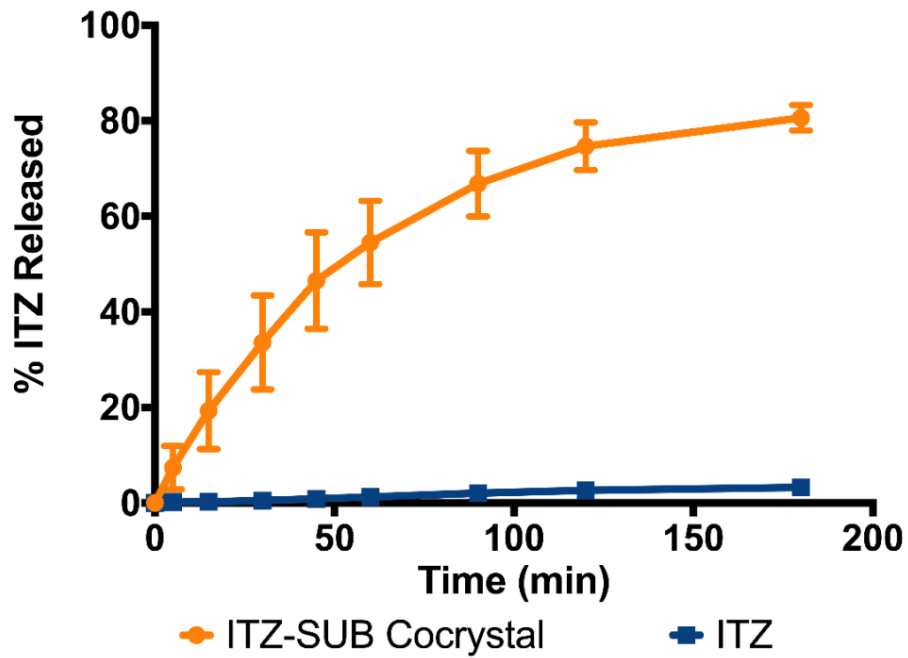


Figure 9. Dissolution profiles of ITZ and spray dried ITZ-SUB powders (SD3) with aerodynamic diameters <math> < 5 \mu\text{m}</math> (n=3).

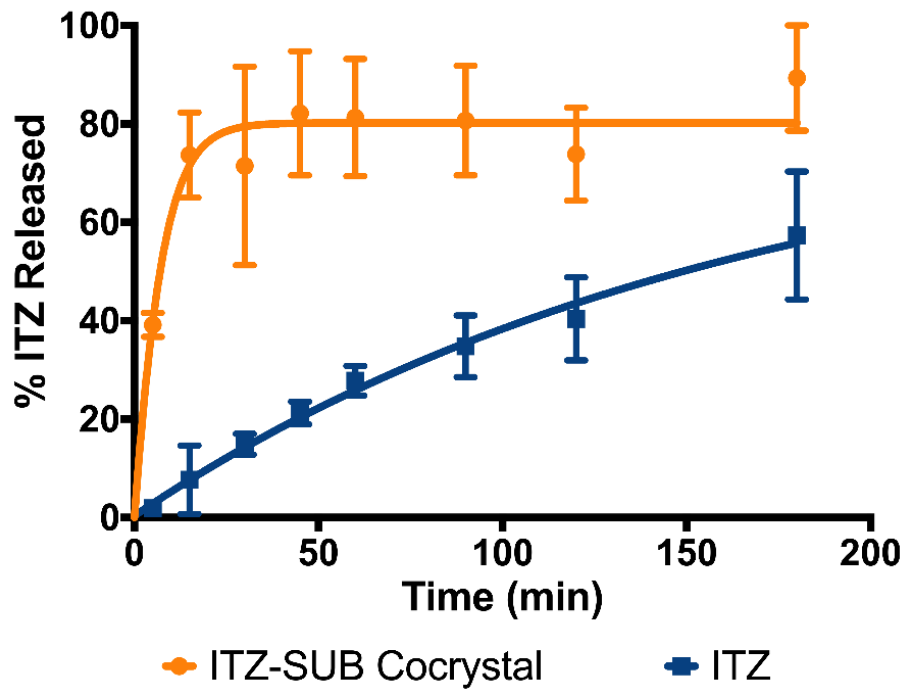


Figure 10. Water sorption isotherms at 25 °C of ITZ, SUB, ITZ-SUB cocrysal, SD ITZ and SD ITZ-SUB.

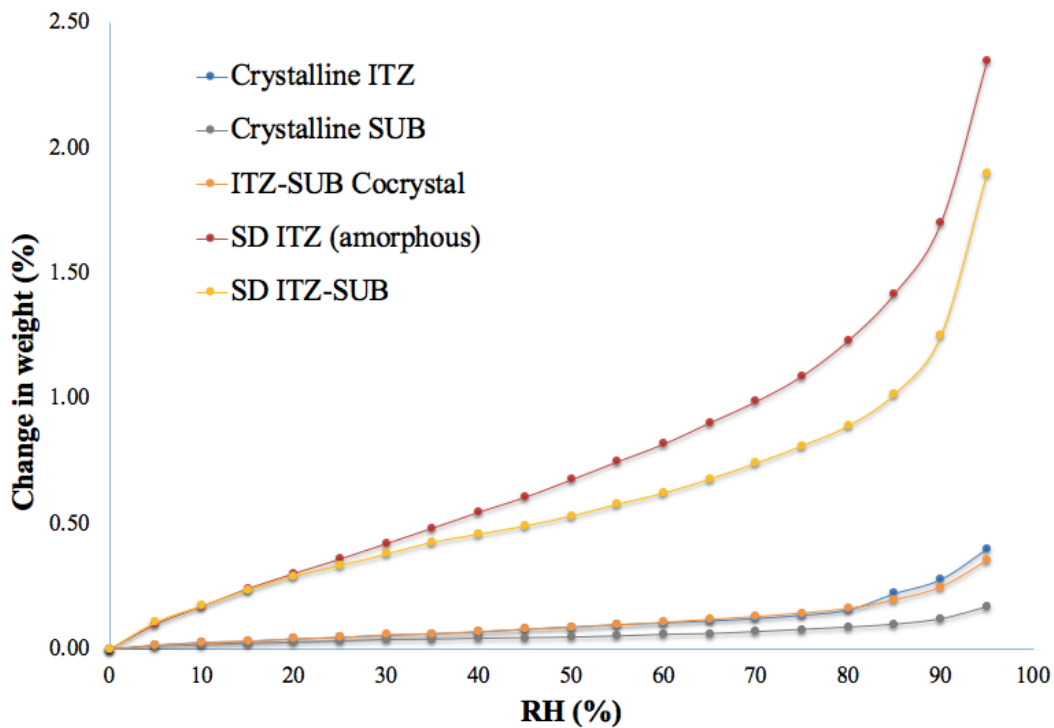
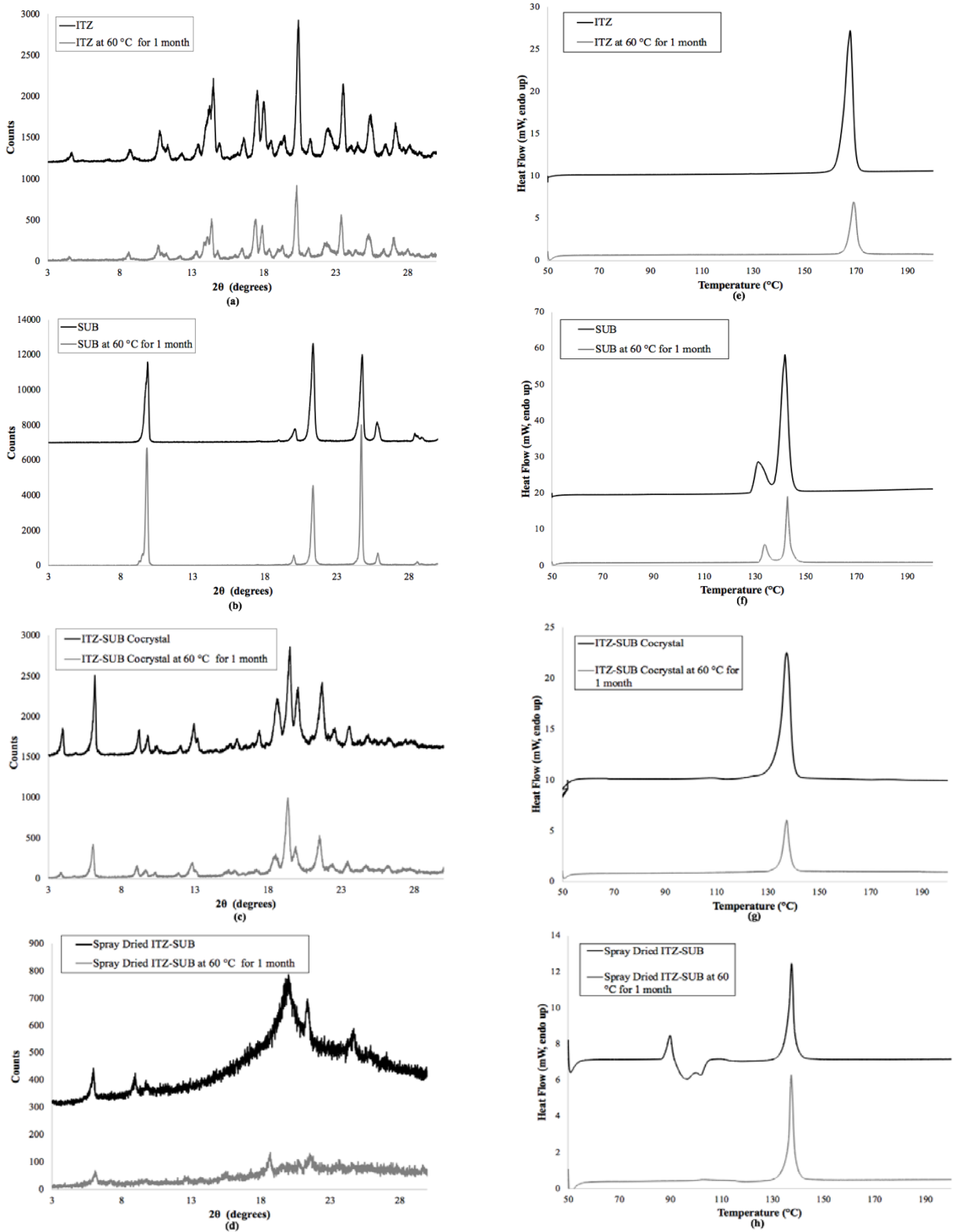


Figure 11. Stability profiles of a) ITZ, b) SUB, c) ITZ-SUB cocrystals and d) spray dried ITZ-SUB using PXRD and e) ITZ, f) SUB, g) ITZ-SUB cocrystals and h) spray dried ITZ-SUB using DSC analysis.



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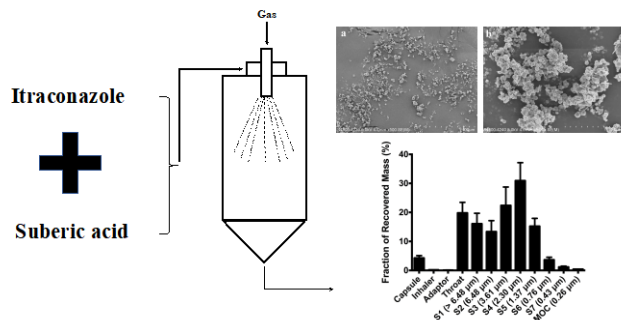
2

3 **Cocrystal Engineering of Itraconazole with Suberic acid via Rotary Evaporation and**
4 **Spray Drying**

5 Jingwen Weng^{1,#}, Si Nga Wong^{1,#}, Xiaoyan Xu¹, Bianfei Xuan¹, Chenguang Wang²,
6 Ruipeng Chen³, Changquan Calvin Sun², Richard Lakerveld², Philip Chi Lip Kwok³, Shing
7 Fung Chow^{1,*}

8

9 TOC graphic:



10

11

12 Synopsis:

13 Using spray drying, we have successfully prepared 1:1 itraconazole-suberic acid
14 agglomerates exhibiting improved dissolution behavior and excellent aerosol performance
15 for deep lung delivery.

16

Supporting Information

Cocrystal Engineering of Itraconazole with Suberic acid via Rotary Evaporation and Spray Drying

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Equal contribution

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Figure S1. SEM image of ITZ-SUB cocrystal.

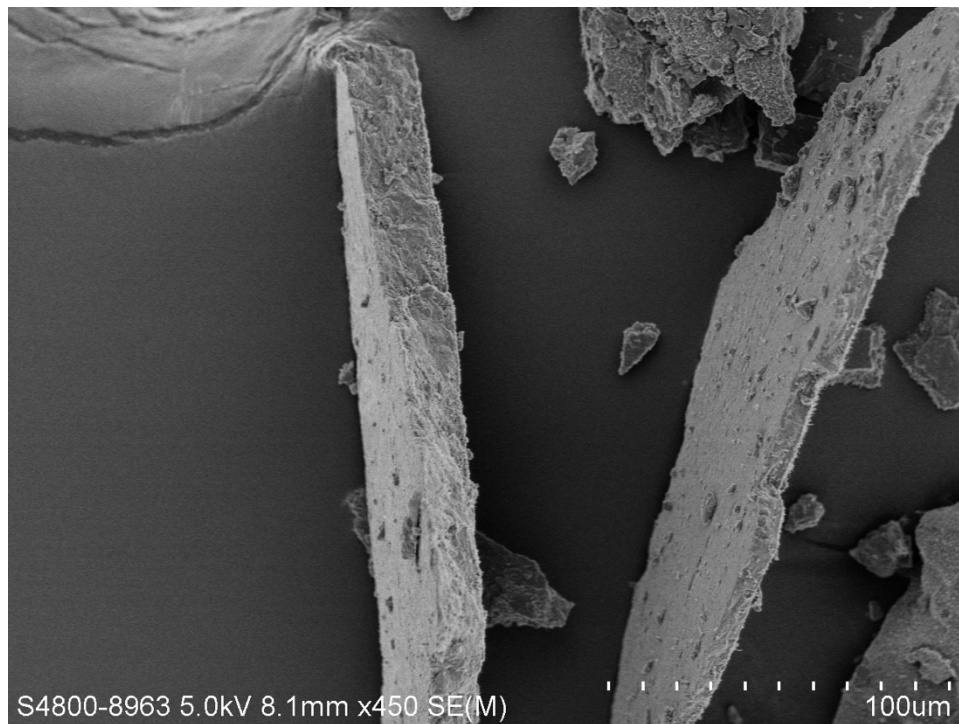


Figure S2. PXRD patterns of spray dried ITZ-SUB (SD1-6).

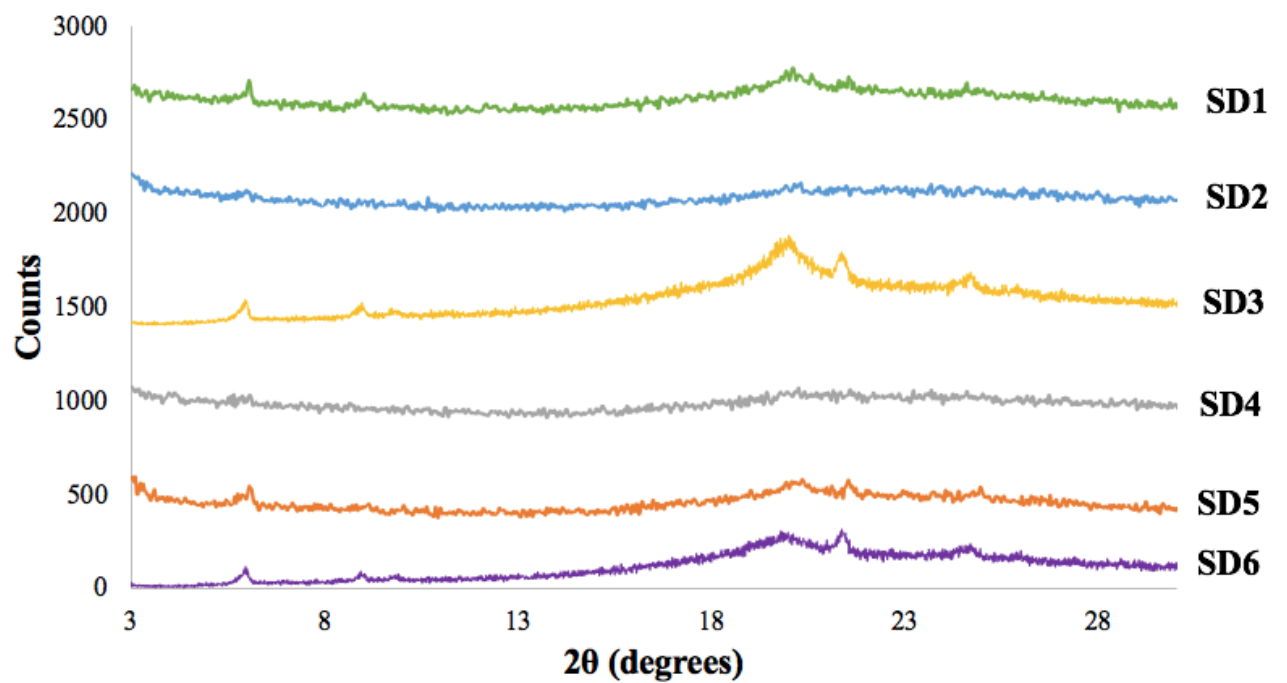


Figure S3. DSC profiles of spray dried ITZ-SUB (SD1-6).

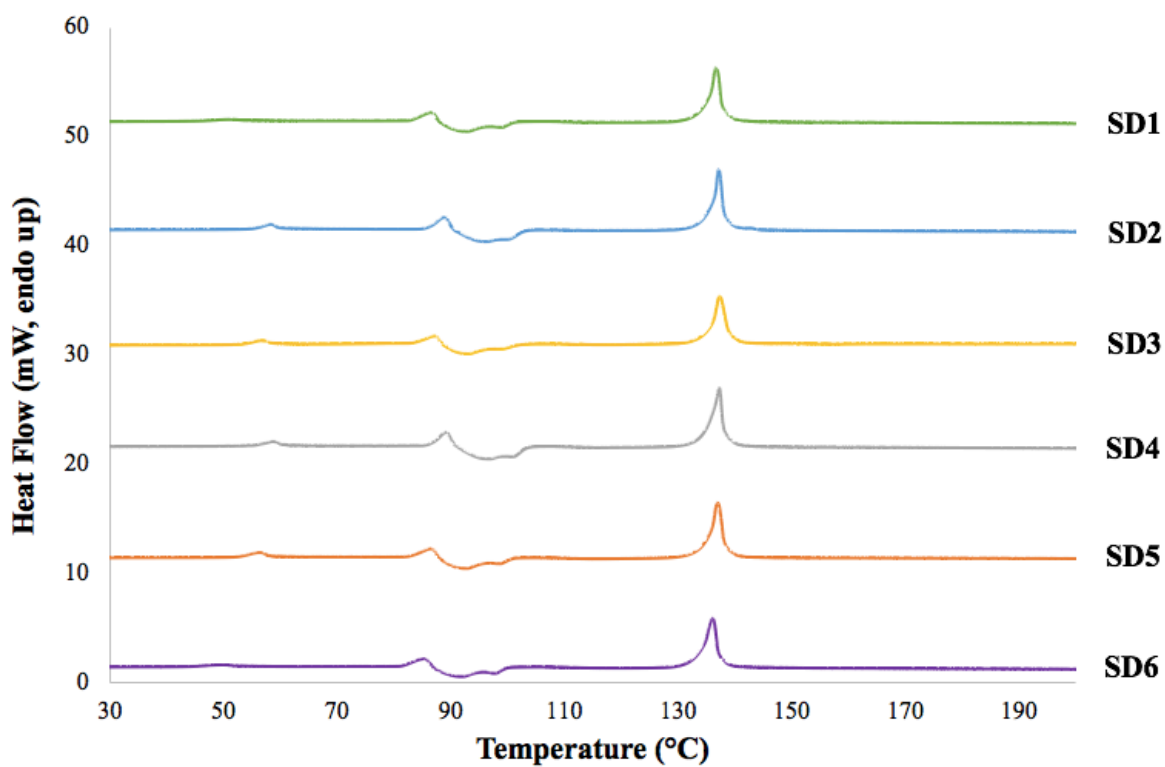


Figure S4. FTIR spectra of spray dried ITZ-SUB (SD1-6).

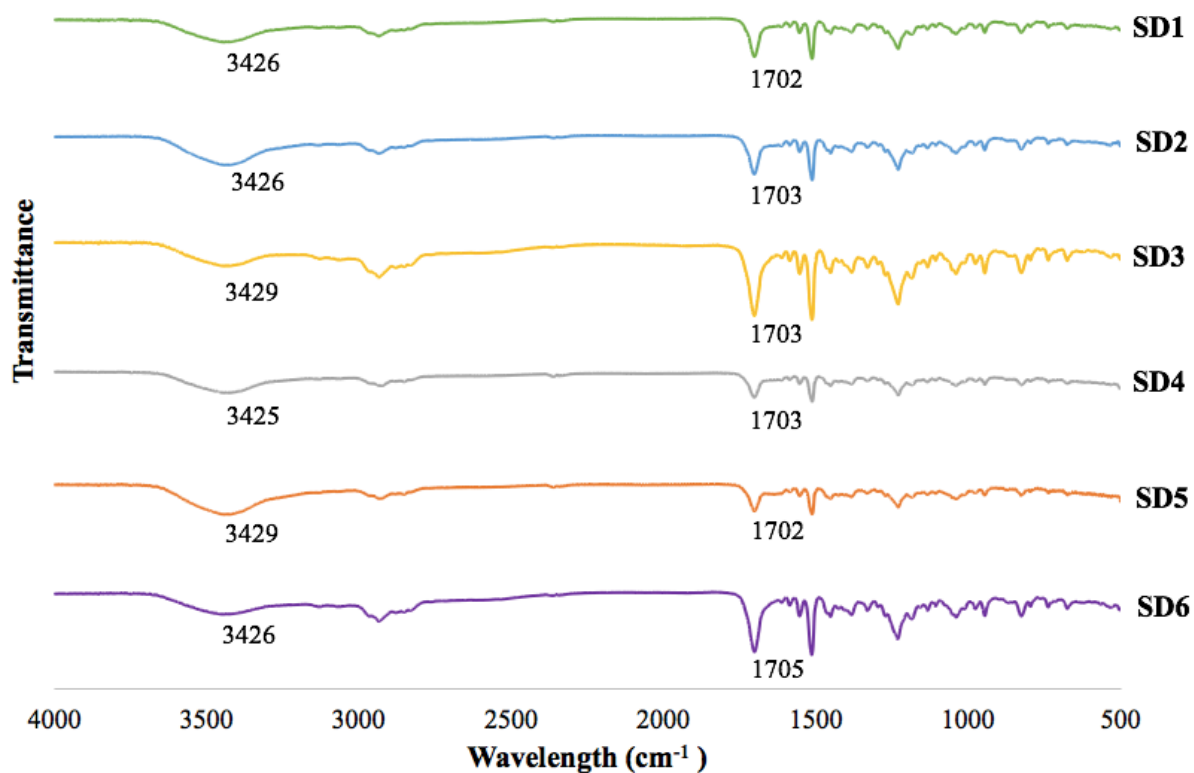


Figure S5. SEM images of spray dried ITZ-SUB formulations at 5k \times magnification.

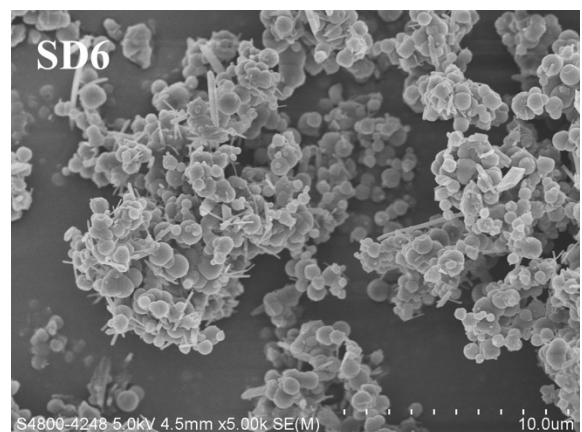
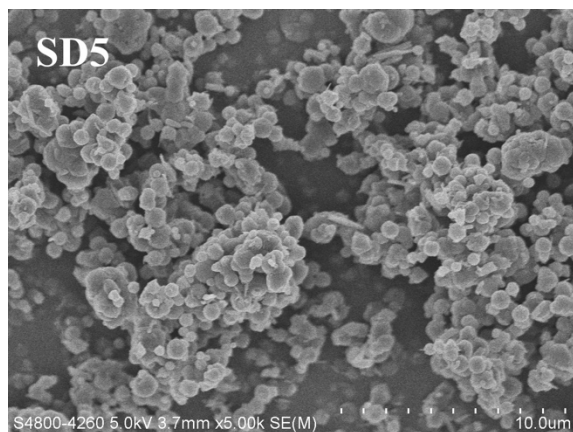
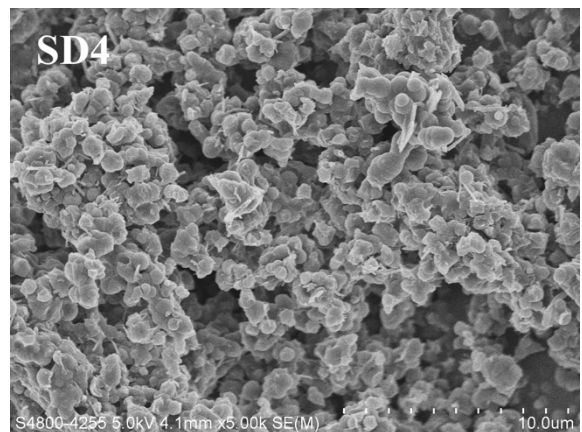
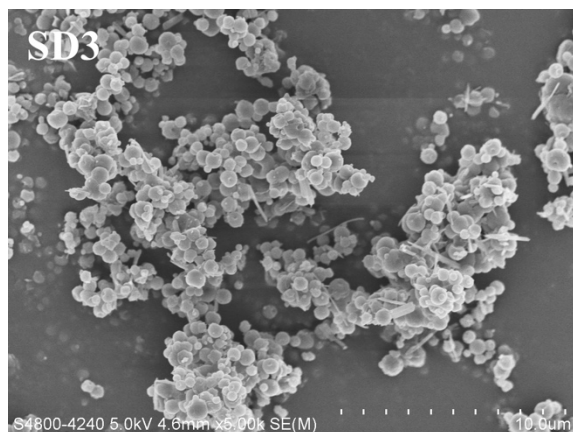
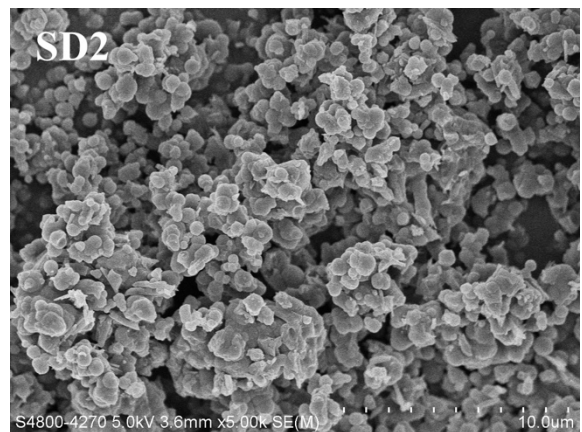
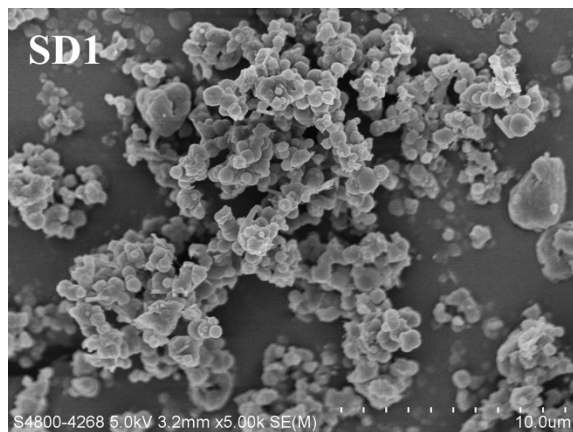


Figure S6. NGI dispersion data of spray dried ITZ-SUB.

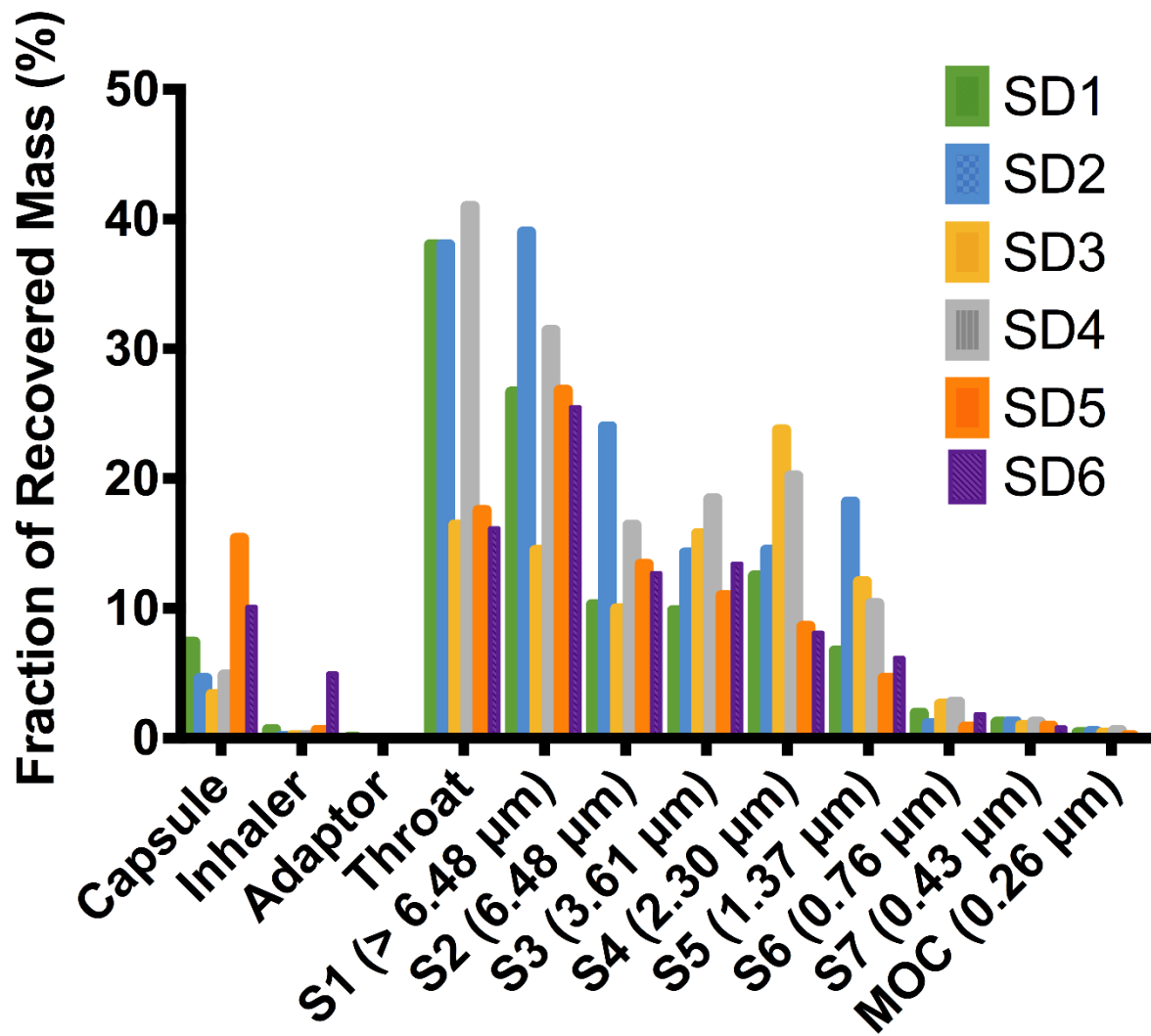


Figure S7. SEM images of spray dried ITZ-SUB powders (SD3) deposited on the throat and different stages of the NGI at 5k \times magnification.

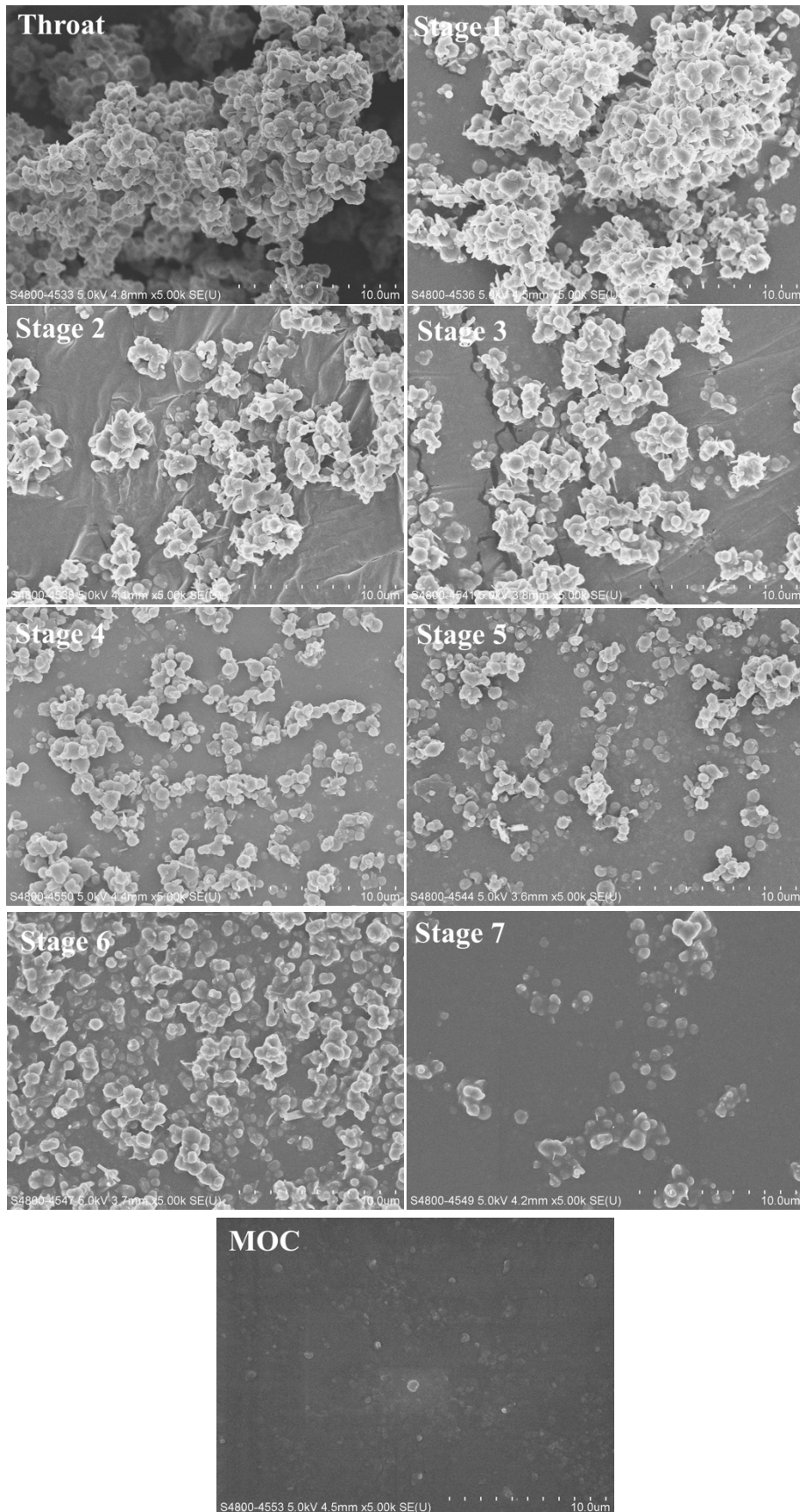


Figure S8. SEM images of a) ITZ and b) ITZ-SUB cocrystal.

