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Droplet Breakup in Expansion-contraction Microchannels

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We investigate the influences of expansion-contraction microchannels on droplet breakup in capillary microfluidic devices. With variations in channel dimension, local shear stresses at the injection nozzle and focusing orifice vary, significantly impacting flow behavior including droplet breakup locations and breakup modes. We observe transition of droplet breakup location from focusing orifice to injection nozzle, and three distinct types of recently-reported tip-multi-breaking modes. By balancing local shear stresses and interfacial tension effects, we determine the critical condition for breakup location transition, and characterize the tip-multi-breaking mode quantitatively. In addition, we identify the mechanism responsible for the periodic oscillation of inner fluid tip in tip-multi-breaking mode. Our results offer fundamental understanding of two-phase flow behaviors in expansion-contraction microstructures, and would benefit droplet generation, manipulation and design of microfluidic devices.

In microfluidic channels, droplets are generated by injecting a liquid phase into another immiscible liquid. Drops break off from an orifice when shearing force and surface tension are balanced¹. Assuming constant shearing force from outer phase, droplets with uniform sizes are generated one by one in a dripping manner. Attributed to the precise control of flow, the monodisperse, and size-controlled droplets generated by droplet microfluidics are extensively used for wide applications, ranging from foods², pharmaceuticals^{3,4}, cosmetics⁵ to materials synthesis^{6–9}. These droplets can function as micro-reactors for chemical reactions^{10,11} and biological assays^{12–14} such as single-molecule polymerase chain reaction (PCR)¹⁵, or as carriers for active ingredients such as drugs^{16,17}, proteins¹⁸ and cells¹⁹. Typically, as micro-reactors or carriers, droplets are merged in diverging channels to initiate chemical reactions^{20,21}, or squeezed through narrow channels to probe the mechanical property of encapsulated protein networks^{18,22} and microcapsules^{23,24}, or split into several daughter droplets^{25,26} for different biological assays^{12,13}. Thus, channels with complex geometry are normally designed to facilitate manipulation of droplets, including mixing, splitting, diluting and fission^{20,21,25–27}. Controlling the dynamical behaviors of individual droplets in complex channels is thus crucial for droplet-based applications^{11–21}.

The geometry of microchannels, like expansions and contractions where flow velocity changes with the varying channel dimension, affects droplet behavior significantly. For example, the increase in flow velocity due to channel contraction increases the shearing force, leading to early breakup of the droplet from inner phase²⁴. An inappropriate design in channel geometry can cause undesired droplet behaviors. For instance, when droplets are used as drug carriers in small vessels, these droplets should not split until arriving at the targeted site²⁸. Moreover, as a large droplet is squeezed through a narrow channel, it may break into multiple daughter droplets in an uncontrolled manner^{29,30}. This uncontrolled breakup of primary droplet significantly jeopardizes the uniformity of final droplets. Consequently, it is highly desired to systematically investigate and quantify the conditions of droplet breakup in complex channels, especially where expansions and contractions in channel dimensions are involved.

When injected into an immiscible outer fluid, the inner liquid can break up in different modes, including geometry-controlled³¹, dripping^{7,32,33}, jetting⁷, tipstreaming^{34–36} and tip-multi-breaking³⁷. The variation in channel geometry results in the transition between breakup modes, and thus changes the size and size distribution of final droplets. For instance, the increase of droplet size and a transition from dripping to geometry-controlled mode are observed, when the distance between two capillaries in flow-focusing capillary devices is increased³⁸. Although quantified relationship between channel geometry and the size of final droplets is well studied for geometry-controlled, dripping, jetting and tipstreaming modes^{7,36,39}, it is yet to be established for the recently reported tip-multi-breaking mode³⁷, by which droplets are generated sequence by sequence with non-uniform size.

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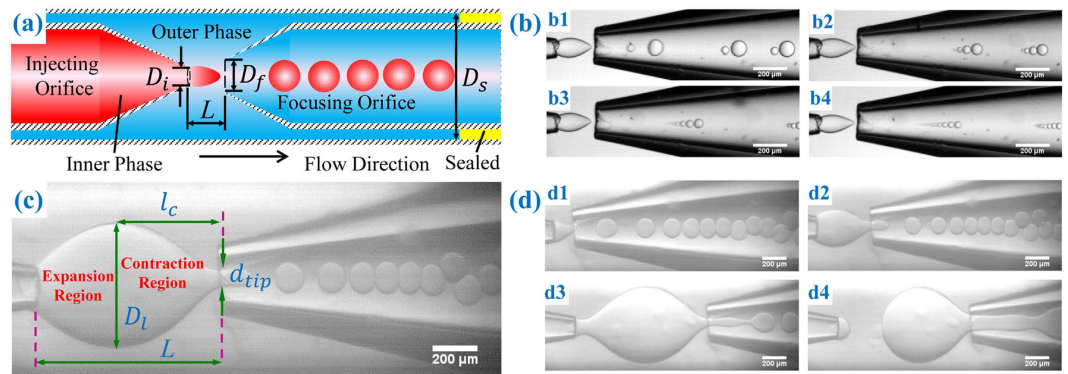


Figure 1. Device used and the influences of channel dimension on flow behaviors. (a) Schematic of the microcapillary device (not to scale). D_i , outer diameter of the injection nozzle; D_f , inner diameter of the focusing orifice; L , distance between two orifices; and D_s , inner dimension of the square capillary. Both inner and outer fluids flow from left to right, and the gutter between the cylindrical and square capillary at right hand-side is sealed during operation (yellow region) to enhance flow focusing, but open at rest, so as to flush out wastes. (b) Droplets produced sequence by sequence in tip-multi-breaking mode with descending size distribution. (b1) 2-droplet sequence, (b2) 4-droplet sequence, (b3) 6-droplet sequence and (b4) 8-droplet sequence. (c) Geometrical parameters characterizing the expansion-contraction microchannel. D_t , maximum diameter of the liquid tip; l_c , contraction distance; d_{tip} , diameter of inner liquid tip at the focusing orifice. l_c and $L - l_c$ represent the length of contraction and expansion region, respectively. (d) Variation of inner fluid tip shape with enlarging orifice distance L . $Q_{in} = 0.5 \text{ mL h}^{-1}$, $Q_{out} = 3.5 \text{ mL h}^{-1}$. $D_i = 186 \mu\text{m}$, $D_f = 210 \mu\text{m}$. (d1) Cone-shape tip with $L = 152.4 \mu\text{m}$, (d2) spindle-shape tip with $L = 444.1 \mu\text{m}$, and (d3,d4) droplet breakup at injection nozzle with $L = 1089.2 \mu\text{m}$. Time interval between (d3,d4) is 50 ms. Scale bars, 200 μm .

We investigate systematically the features of droplet breakup in capillary microfluidic devices with expansion-contraction configurations (Fig. 1a). The influences of varying channel geometry on flow behaviors are studied at the injection nozzle with diverging flow and at the focusing orifice with converging flow. Using local capillary numbers, we characterize the influences by conditions of droplet breakup, such as the location shift of droplet breakup and breakup mode transition. At the injection nozzle, we demonstrate that the local capillary number successfully predicts the condition for breakup location transition. At the focusing orifice, we highlight the transition of droplet breakup modes, and quantify both droplet size and number of droplets in the tip-multi-breaking mode³⁷ (Fig. 1b). We also examine the mechanism responsible for evolution of inner liquid tip and the oscillation period of the tip. Our understanding of droplet breakup behavior influenced by the channel geometry, offers valuable guidelines for designing microchannels to generate and manipulate droplets in a precisely controlled manner.

Experiments

Capillary microfluidic devices were used to study the hydrodynamic behaviors of two-phase flows in microchannel with an expansion-contraction structure. The capillary microfluidic device was fabricated by aligning two tapered cylindrical glass capillaries inside a square capillary (inner dimension $D_s = 1050 \mu\text{m}$), as shown in Fig. 1a. We varied three geometrical parameters D_i , D_f and L systematically, which were outer diameter of the injection nozzle, inner diameter of the focusing orifice, and distance between the two orifices, respectively. We summarized the combinations of geometrical parameters in Table 1. Three dynamic dimensions (Fig. 1c), the maximum diameter of inner liquid tip D_t , inner tip diameter at the focusing orifice d_{tip} and the axial distance between the maximum inner tip and focusing orifice l_c , depend on both geometrical parameters D_i , D_f and L and the dynamic flow process (inner and outer flow rates Q_{in} and Q_{out} , respectively). The length of contraction, where flow converges, is represented by l_c ; while that of the expansion, where flow diverges, is represented by $L - l_c$ (Fig. 1c).

Both inner and outer fluids were injected into the microcapillary device from left to right, Fig. 1a. The flow rates for inner (Q_{in}) and outer (Q_{out}) phases were controlled by syringe pumps (Longer Pump). Q_{in} was experimentally confirmed to be constant with various orifice distance L (see “confirming constant inner flow rate” in supplementary information). The gutter between the cylindrical and square capillary at right hand side was sealed (yellow region in Fig. 1a) during operation, so as to enhance the flow contraction at the focusing orifice. The flow is visualized, monitored and recorded (images and videos) by a high-speed digital camera (MotionPro[®] X4, IDT, Taiwan, and Phantom M110) equipped with an inverted microscope (XD101, Nanjing Jiangnan Novel Optics Co. Ltd, and Nikon TS100). Captured images and videos were analyzed by ImageJ.

The fluid employed in experiments was water-in-oil two-phase flow, where the outer phase was silicone oil with fixed viscosity η_{out} , and inner phase were various glycerol-water mixtures with different values of viscosity η_{in} (see supplementary Table S1). The mixture of 70 wt.% glycerol and 30 wt.% distilled water was used as inner phase fluid for most of the experiments, except that of determining the condition for the transition of droplet breakup location as a function of viscosity ratio ξ (defined as $\xi = \eta_{in}/\eta_{out}$). The viscosity was measured by a viscometer (microVISC[™], RheoSense, Inc.). $\eta_{out} = 492.9 \pm 6.9 \text{ mPa s}$ for silicone oil, and $\eta_{in} = 19.07 \pm 0.12 \text{ mPa s}$ for

Case Number	D_i (μm)	D_f (μm)	L (μm)	Case Number	D_i (μm)	D_f (μm)	L (μm)
A	86	258	97–714	K	147	242	806
B	186	258	197–1100	L	147	242	267
C	203	258	119–1136	M	147	242	570
D	217	258	130–1284	N	147	242	150
E	186	150	126–1166	O	147	176	272
F	186	210	146–1094	P	73	242	270
G	186	353	155–1038	Q	208	242	265
H	265	155	1208–1654	L	147	197	409
I	305	155	163–1410				
J	305	258	192–1263				

Table 1. Eighteen cases tested with different device geometries. Cases A–J, with various combinations of D_i and D_f , are conducted to investigate the condition for the location shift of droplet breakup from focusing orifice to injection nozzle. For each case from A to J, the orifice distance L is variable, starting from an initial value comparable to the diameter of injection nozzle D_i , ending with the value when droplet breakup occurs at the injection nozzle. The range of L for each case from A to J can also be found in Fig. 3. Cases K–Q are used to characterize the influence of device geometry on tip-multi-breaking mode. Different combinations of D_i , D_f and L are studied. Case L is used to develop the correlation between the oscillation period T and the capillary number Ca_{focus} in tip-multi-breaking mode. The capillary number Ca_{focus} ranges from 0.15 to 0.45 in our experiments.

70 wt.% glycerol ($\xi = 0.039$). The interfacial tension was measured by a ring tensiometer (Surface Tensiometer 20, Cole-Parmer) to be $\gamma = 30.07 \text{ mN m}^{-1}$ without any surfactants.

Results and Discussion

Influences of channel dimension on flow behaviors. The influence of channel geometry is manifested in the two local shear stresses at the injection nozzle and focusing orifice, respectively. Qualitatively, small value of L renders shear stress strong at the focusing orifice, but weak at the injection nozzle. When fluid flows in a channel with short distance L , the sharp contrast in local stresses leads to a cone-shape tip (Fig. 1d1). Droplets are generated at the focusing orifice where channel converges. With increasing L , the decreased difference in local stresses results in a spindle-shape tip (Fig. 1d2). However, if L increases further, the inner fluid tip is likely to break up at the injection nozzle where channel diverges (Fig. 1d3,d4). Therefore, the breakup location shifts from the focusing orifice to injection nozzle. Meanwhile, the detached drop from the injection nozzle would break up again into multiple daughter droplets with non-uniform size as it is squeezed into the focusing orifice (Fig. 1d3,d4, see supplementary movie S1). To produce uniform droplets, breakup at injection nozzle should thus be avoided.

The variation in local shear stresses influences droplet breakup mode as well as droplet size distribution. As local shear stresses vary, the capillary number Ca , representing the ratio of shear stresses to surface tension, also changes. For instance, decrease of local capillary number at the focusing orifice by enlarging L leads to a transition from dipping to geometry-controlled mode, as observed by Benson *et al.*³⁸ and in the present work. Interestingly, we also found the variation of tip-multi-breaking mode with L (Fig. 2, see supplementary movie S2). Tip-multi-breaking mode was previously reported with descending size distribution in one droplet train³⁷ (Figs 1b and 2a). Here we find that the size distribution in one drop sequence can either keep constant for a while and then decrease (“constant-decreasing”, Fig. 2b), or increase first, then keep constant and finally decrease (“increasing-constant-decreasing”, Fig. 2c), depending on the orifice distance L .

Figure 3 maps various flow behaviors in Plane (L, l_c). The cone-shape and spindle-shape tips are separated by $l_c/L = 1$. Breakup at the focusing orifice lies above the boundary of $l_c = 0.5L$, while breakup at the injection nozzle is constrained by $l_c < 0.5L$. To understand this phase map, we introduce a ratio of the representative length for expansion $L - l_c$ to that for contraction l_c , $\chi = (L - l_c)/l_c$, which increases with L . The larger χ corresponds to stronger diverging flow but weaker converging flow. Initially, $\chi = 0$ for small values of L , indicating a converging flow at the focusing orifice without diverging. As such, shear stress at the focusing orifice is large enough to drag the inner tip into cone-shape, featured by $l_c = L$ (Fig. 3). As $\chi > 0$, the flow first diverges and then converges, so the tip of inner flow is spindle-shape, represented by $l_c < L$ in Fig. 3. In the range of $0 \leq \chi < 1$, droplet breakup occurs at the focusing orifice, and transitions of droplet breakup mode can be observed with an increase in L . When diverging flow magnifies further with the increase of L , droplet breaks up at the injection nozzle with $\chi > 1$. Therefore, the boundary for the transition of breakup location from focusing orifice to injection nozzle reads $\chi = 1$, which results in $l_c = 0.5L$ (Fig. 3).

Condition for the transition of droplet breakup location. To determine the condition for breakup location shifting from the focusing orifice to injection nozzle, we exploit the local capillary number at the injection nozzle in the form of $Ca_{inject} = F_s/F_\gamma$, with F_s and F_γ being the total shear forces and capillary forces, respectively. Firstly, we estimate F_s exerted on the inner fluid tip based on a modified Stokes’ drag^{32,40,41},

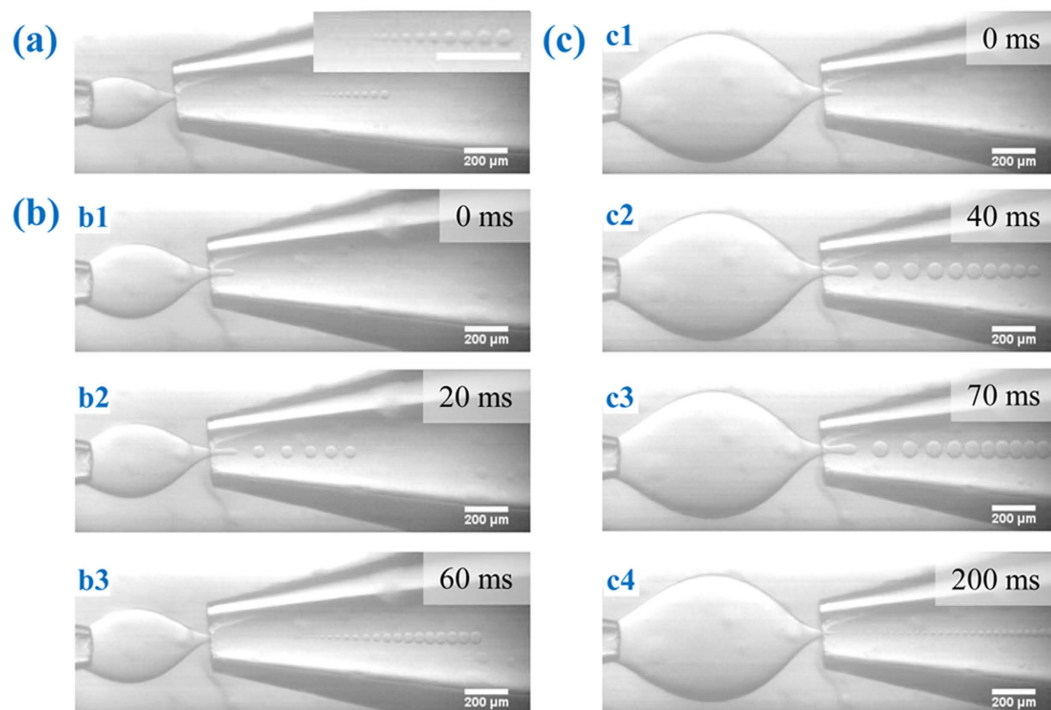


Figure 2. Effects of channel dimension on size distribution in tip-multi-breaking mode. All images are obtained with $D_i = 186 \mu\text{m}$, $D_f = 210 \mu\text{m}$, $Q_{in} = 0.02 \text{ mL h}^{-1}$ and $Q_{out} = 3.5 \text{ mL h}^{-1}$. (a) Droplet sequence with descending sizes in one droplet train. $L = 372.7 \mu\text{m}$. Inset: magnification of the droplet sequence. (b) Droplet size firstly keeps constant for dozens of milliseconds, and then decreases with time (“constant-decreasing”). $L = 537.5 \mu\text{m}$. (c) Droplet size increases first, then keeps constant for a while, and finally decreases with time (“increasing-constant-decreasing”). $L = 919.6 \mu\text{m}$. See supplementary movie S2 for details. Scale bars, $200 \mu\text{m}$.

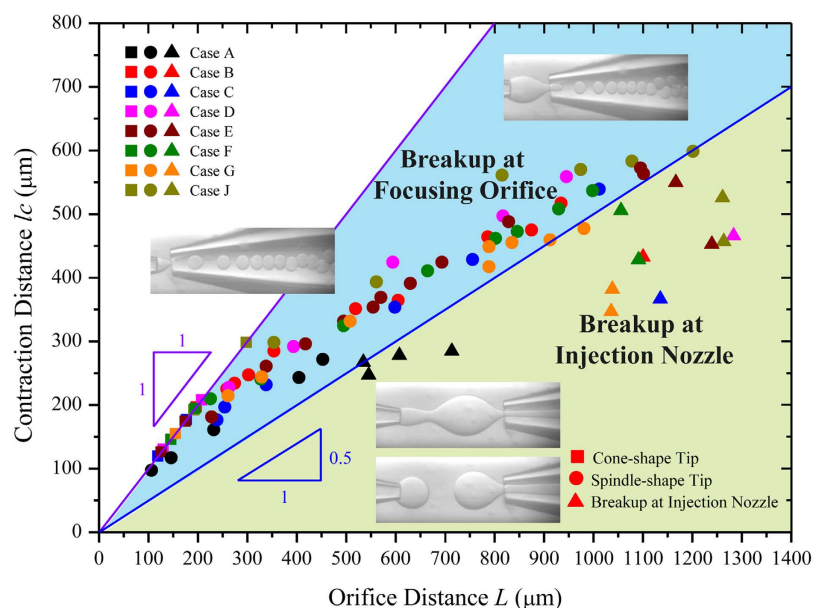


Figure 3. Phase diagram showing flow behaviors influenced by channel dimension. $l_c = L$ represents cone-shape tip, while the region of $l_c < L$ indicates the spindle-shape tip. $l_c = 0.5L$ marks the transition of droplet breakup location from focusing orifice to injection nozzle. Different symbol colors represent different cases (Table 1). Different symbol shapes distinguish flow behaviors, where square denotes cone-shape tip, circle is for spindle-shape tip, and triangle means breakup at injection nozzle. The phase diagram is obtained with $Q_{in} = 0.5 \text{ mL h}^{-1}$ and $Q_{out} = 3.5 \text{ mL h}^{-1}$. Insets: snapshots of cone-shape tip, spindle-shape tip and breakup at focusing orifice and injection nozzle, respectively.

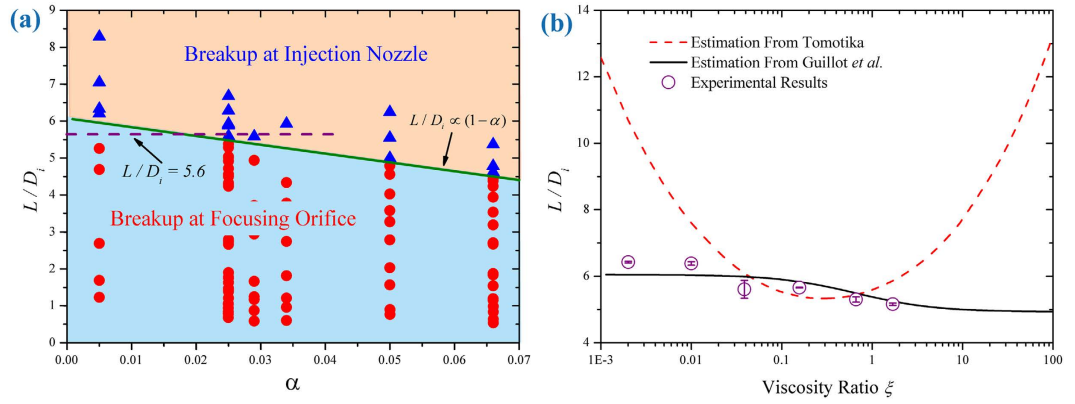


Figure 4. Condition for the transition of droplet breakup location. (a) Critical L/D_i for breakup at injection nozzle as a function of α . Solid line represents the condition of $L/D_i \propto (1 - \alpha)$ from Eq. (4) where $\alpha = \pi D_i^2 / 4D_s^2$, while dashed line denotes $L/D_i = 5.6$ as an estimate under the assumption of $1 - \alpha \approx 1$ with $\alpha \leq 0.04$. For the same α , several values of D_j are tested (see supplementary Fig. S2). Triangle represents breakup at injection nozzle, while circle denotes breakup at focusing orifice. Data are obtained with viscosity ratio $\xi = 0.039$. (b) Plot of L/D_i as a function of viscosity ratio ξ . The black solid curve is obtained from Eq. (7) based on the result of Guillot *et al.*⁴², while the red dashed curve is estimated from Tomotika⁴⁶ (see “determining the most unstable mode of a viscous jet” in supplementary information). Experimental results show better agreement with solid-curve estimation. Since the dashed-curve is taken under the circumstance of unbounded quasi-static flow, while solid-curve considers device confinement and flow rates, the difference in the two estimates reveals the significance of channel confinement and flow rates in affecting confined droplet breakup.

$$F_s = 2\pi u_{out} \eta_{out} \frac{1 + 1.5\xi}{1 + \xi} (D_c - D_i), \quad (1)$$

where $D_c = \sqrt[3]{LD_i^2}$ is the characteristic diameter of inner liquid tip, $\xi = \eta_{in} / \eta_{out}$ is viscosity ratio, and $u_{out} = Q_{out} / (D_s^2 - \pi/4D_i^2)$ is the mean velocity of outer fluid at the injection nozzle. Capillary forces are evaluated as^{32,41}

$$F_\gamma = \pi D_i \gamma. \quad (2)$$

Thus, the local capillary number at the injection nozzle is

$$Ca_{inject} = \frac{2Q_{out} \eta_{out}}{\gamma D_s^2} \frac{1 + 1.5\xi}{1 + \xi} \frac{\sqrt[3]{LD_i^2} - D_i}{D_i(1 - \alpha)}, \quad (3)$$

where $\alpha = \pi D_i^2 / 4D_s^2$ is the ratio of cross-sectional area of the injection nozzle to that of the square capillary. Droplet breakup at the injection nozzle occurs only when Ca_{inject} exceeds a critical value Ca_{cri} . Eq. (3) indicates that Ca_{inject} increases with increasing L , so Ca_{cri} implies a critical L for the transition to occur. The critical L depends on fluid properties, channel dimensions, and flow rates.

Now we determine the critical L triggering the breakup at the injection nozzle based on Eq. (3). The dynamic dimension D_l is found to be proportional to orifice distance L (see supplementary Fig. S1), thereby it is reasonable to estimate $D_l \propto L$. Assuming constant Q_{out} , η_{out} , η_{in} , γ and D_s in Eq. (3), we have $Ca_{inject} \propto (L - D_i) / D_i(1 - \alpha)$. Therefore, the condition for droplet breakup at the injection nozzle, $Ca_{inject} = Ca_{cri}$, leads to

$$L/D_i \propto (1 - \alpha). \quad (4)$$

Eq. (4) shows that the condition is represented as the ratio of orifice distance L to the outer diameter of the injection nozzle D_p , L/D_p , which decreases linearly with α . We confirm this relation experimentally as the solid line in Fig. 4a.

In subsequent analysis, we examine the analytic solution for condition L/D_p , which can be achieved by taking all the influencing parameters into consideration and making a quantitative estimation of D_j . Practically, α is much smaller than unity, (for example $D_i = 200 \mu\text{m}$ gives $\alpha \approx 0.028$), so we assume that α is small enough to assume $1 - \alpha \approx 1$ in the following analysis. According to Rayleigh-Plateau instability, the most unstable mode for the jet breakup gives $kR_0 = \sqrt{2}/2$ ^{42,43}, where $k = 2\pi/\lambda$ is the wave number with wavelength approximated as $\lambda = L$ in our case. R_0 is the unperturbed jet radius and estimated as $R_0 = (D_i + D_l)/4$, which is the average radius of the injection nozzle and the maximum tip. Replacing k and R_0 by D_p , D_l and L , we get,

$$\frac{L}{D_i} = \frac{\pi}{\sqrt{2}} + \frac{D_l}{D_i}. \quad (5)$$

Eq. (5) provides an accurate estimation of the relation between D_l and L compared with the previous simplified one $D_l \propto L$. Since breakup at the injection nozzle occurs when $Ca_{inject} = Ca_{cri}$, the following relation is achieved by rewriting Eq. (3) with the assumption of $1 - \alpha \approx 1$,

$$\sqrt[3]{\frac{L}{D_i} \left(\frac{D_l}{D_i} \right)^2} - 1 = \frac{Ca_{cri} \gamma D_s^2}{2Q_{out} \eta_{out}} \frac{1 + \xi}{1 + 1.5\xi}. \quad (6)$$

Solving Eqs. (5) and (6) together, L/D_i is finally obtained in the following form

$$\frac{L}{D_i} = \frac{\sqrt{2}\pi}{3} + \frac{a}{6} + \frac{\pi^2}{3a}, \quad (7)$$

with $a = (108b - 2\pi^3\sqrt{2} + 12\sqrt{81b^2 - 3b\pi^3\sqrt{2}})^{1/3}$, and $b = \left(\frac{Ca_{cri} \gamma D_s^2}{2Q_{out} \eta_{out}} \frac{1 + \xi}{1 + 1.5\xi} + 1 \right)^3$.

Eq. (7) is achieved under two assumptions: $1 - \alpha \approx 1$, and neglecting the influence of focusing orifice (D_f). Our experimental results validate these assumptions. For experimental cases with $\alpha \leq 0.04$ (guaranteeing $1 - \alpha \approx 1$) but different D_f values, the critical condition L/D_i basically keeps constant as $L/D_i = 5.6$ (dashed line in Fig. 4a, also see supplementary Fig. S2), independent of α . However, a determination of Ca_{cri} is necessary to predict L/D_i theoretically.

As presented by Erb *et al.*³², a Ca_{cri} value of 0.1 is accurate enough to predict the condition for droplet breakup over a wide range of viscosity ratios. We thus plot L/D_i as a function of ξ by using $Ca_{cri} = 0.1$. The other parameters involved are $Q_{out} = 3.5 \text{ mL h}^{-1}$, $\eta_{out} = 492.875 \text{ mPa s}$, $\gamma = 30.07 \text{ mN m}^{-1}$, $D_s = 1050 \text{ }\mu\text{m}$, respectively. As shown in Fig. 4b, the prediction (solid line) agrees well with experimental results when viscosity ratio varies over three orders of magnitude.

Characteristics of tip-multi-breaking mode. Having determined the condition for droplet breakup at the injection nozzle, we now turn our attention to breakup at the focusing orifice. According to Taylor⁴⁴, we define outer phase capillary number as $Ca_{focus} = \eta_{out} \varepsilon_{out} D_l / \gamma$ locally at the focusing orifice, where $\varepsilon_{out} = \Delta u_{out} / l_c$ is the strain rate represented by the velocity gradient along the flow direction. Δu_{out} is estimated as the difference between the average flow velocity at the focusing orifice, $4Q_{out} / \pi D_f^2$, and that at the maximum tip diameter, $4Q_{out} / (4D_s^2 - \pi D_l^2)$, which yields Ca_{focus} in the following form

$$Ca_{focus} = \frac{4\eta_{out} Q_{out} D_l}{\gamma l_c} \left(\frac{1}{\pi D_f^2} - \frac{1}{4D_s^2 - \pi D_l^2} \right). \quad (8)$$

As contraction length l_c increases, capillary number Ca_{focus} decreases, indicating a weaker external shear stress. Since increasing distance L results in larger contraction length l_c (Fig. 3), capillary number Ca_{focus} decreases with L . Eq. (8) thus accounts for the transition from dripping to geometry-controlled mode by increasing L . Next, we use Ca_{focus} to quantify the droplet breakup in tip-multi-breaking mode at the focusing orifice.

Droplet sizes vary with local capillary number. For dripping, a scaling law⁴⁵ suggests $R/h \sim Ca^{-1}$, with Ca being capillary number, R being droplet radius, and h being channel dimension. However, for tip-multi-breaking mode, such a scaling cannot hold, because droplet sizes are not uniform even for the same capillary number (Fig. 2). In this case, droplet size should be normalized by a dynamic length scale instead of a static one. In tip-multi-breaking mode, force balance between shear stress and surface tension gives $\eta_{out} u_{out} / r_{tip} \sim \gamma / R_{drop}$, where R_{drop} and r_{tip} are radii of the droplet and tip neck, respectively. If the characteristic velocity u_{out} is approximated as $u_{out} \sim \varepsilon_{out} D_l$ by considering geometry parameters, then the force balance gives $\eta_{out} (\varepsilon_{out} D_l) / d_{tip} \sim \gamma / D_{drop}$ with R_{drop} and r_{tip} replaced by D_{drop} and d_{tip} . Finally, we arrive at a scaling law,

$$D_{drop} / d_{tip} \sim Ca_{focus}^{-1}. \quad (9)$$

Eq. (9) provides a scaling for tip-multi-breaking in a form similar to $R/h \sim Ca^{-1}$, but different in that droplet diameter D_{drop} is scaled by a dynamic length d_{tip} , rather than the static channel dimension h . This scaling is confirmed to agree with the experimental data very well, as shown in Fig. 5a. For a droplet sequence with polydisperse droplets (Fig. 2), D_{drop} / d_{tip} equals constant due to the same capillary number Ca_{focus} . So the non-uniformity of the droplets is interpreted by the change of inner tip diameter d_{tip} . For example, descending size distribution of tip-multi-breaking mode in Fig. 2a is the result of d_{tip} thinning monotonically with time during the formation of one droplet sequence. Likewise, different size distributions in Fig. 2b,c are attributed to the different ways in which d_{tip} alters with time.

Apart from influencing droplet size distribution in tip-multi-breaking mode, capillary number Ca_{focus} affects the number of droplets in the sequence as well. Here, we focus only on the droplet sequence with descending size distribution, as shown in Fig. 2a. Previously, we found that droplet number n qualitatively increases with capillary

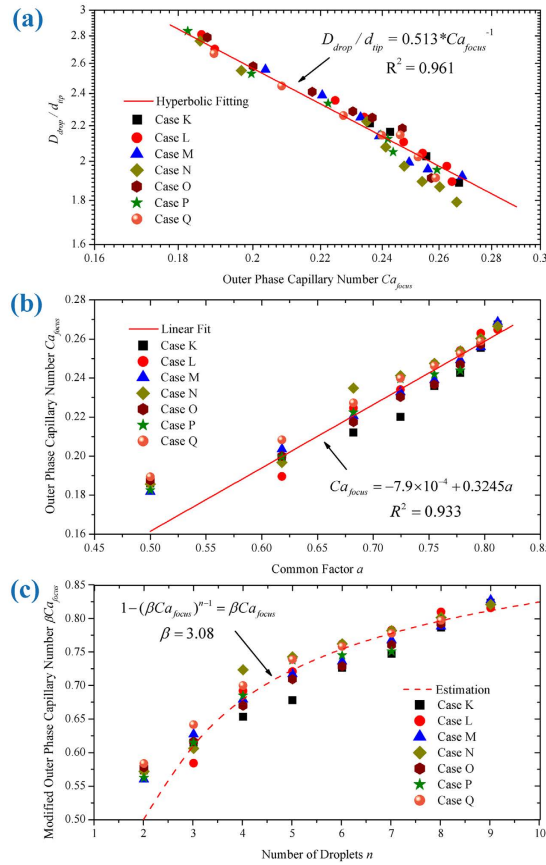


Figure 5. Characteristics of tip-multi-breaking mode. (a) Log-log plot of droplet size as a function of local capillary number Ca_{focus} . Experimental data collapse onto a fitted line of $D_{drop}/d_{tip} = 0.513Ca_{focus}^{-1}$. (b) Outer phase capillary number Ca_{focus} showing a linear relation to common ratio a . (c) Plot of capillary number Ca_{focus} versus droplet number n . Dashed line shows the estimation of $1 - (\beta Ca_{focus})^{n-1} \approx \beta Ca_{focus}$ ($\beta = 3.08$), which gives a fair prediction when $n > 2$, and the larger the value of n is, the smaller the deviation becomes. Different symbols denote different cases (Table 1).

number Ca_{focus} ³⁷. Now, to obtain a quantitative relationship between n and Ca_{focus} , it is necessary to summarize here some fundamental results from ref. 37. First, the individual droplet size in one droplet sequence constitutes a geometrical progression, with common factor being a . Second, common factor a and droplet number n are related as

$$1 - a^{n-1} \approx a. \tag{10}$$

If a and Ca_{focus} are related, then the relation between n and Ca_{focus} would finally be obtained based on Eq. (10). Experimentally, common ratio a is found to increase linearly with Ca_{focus} , $Ca_{focus} \approx 0.3245a$ (Fig. 5b). Then, by assuming $a = \beta Ca_{focus}$ (β is a constant), we get

$$1 - (\beta Ca_{focus})^{n-1} \approx \beta Ca_{focus}. \tag{11}$$

When constant β fits as $\beta = 1/0.3245 = 3.08$, experimental data basically collapse around the prediction given by Eq. (11) (Fig. 5c). The discrepancy between the prediction and experimental data gets smaller and smaller when n grows. In fact, Ca_{focus} varies smoothly, while n are discrete natural numbers ($n > 1$). So for every single n , there should be a narrow range rather than only one value of Ca_{focus} , as displayed in Fig. 5c. Based on Eq. (11) droplet train with prescribed droplet number can be tuned on-demand by varying the matching capillary number, for instance by changing outer phase flow rate. These droplet sequences may have potential applications in materials science, for example, in designing new barcode emulsions and particles with multiple cores of different sizes and numbers.

Oscillation of the inner liquid tip. The periodic oscillation of the tip features tip-multi-breaking mode. We show the evolution of d_{tip} for three types of tip behaviors, descending, constant-decreasing, and increasing-constant-decreasing in Fig. 6a–c, respectively, from experimental data. To quantify the variation of outer-fluid viscous stress during the d_{tip} evolution, we define its local capillary number Ca_{tip} by $Ca_{tip} = 4\eta_{out}Q_{out}/$

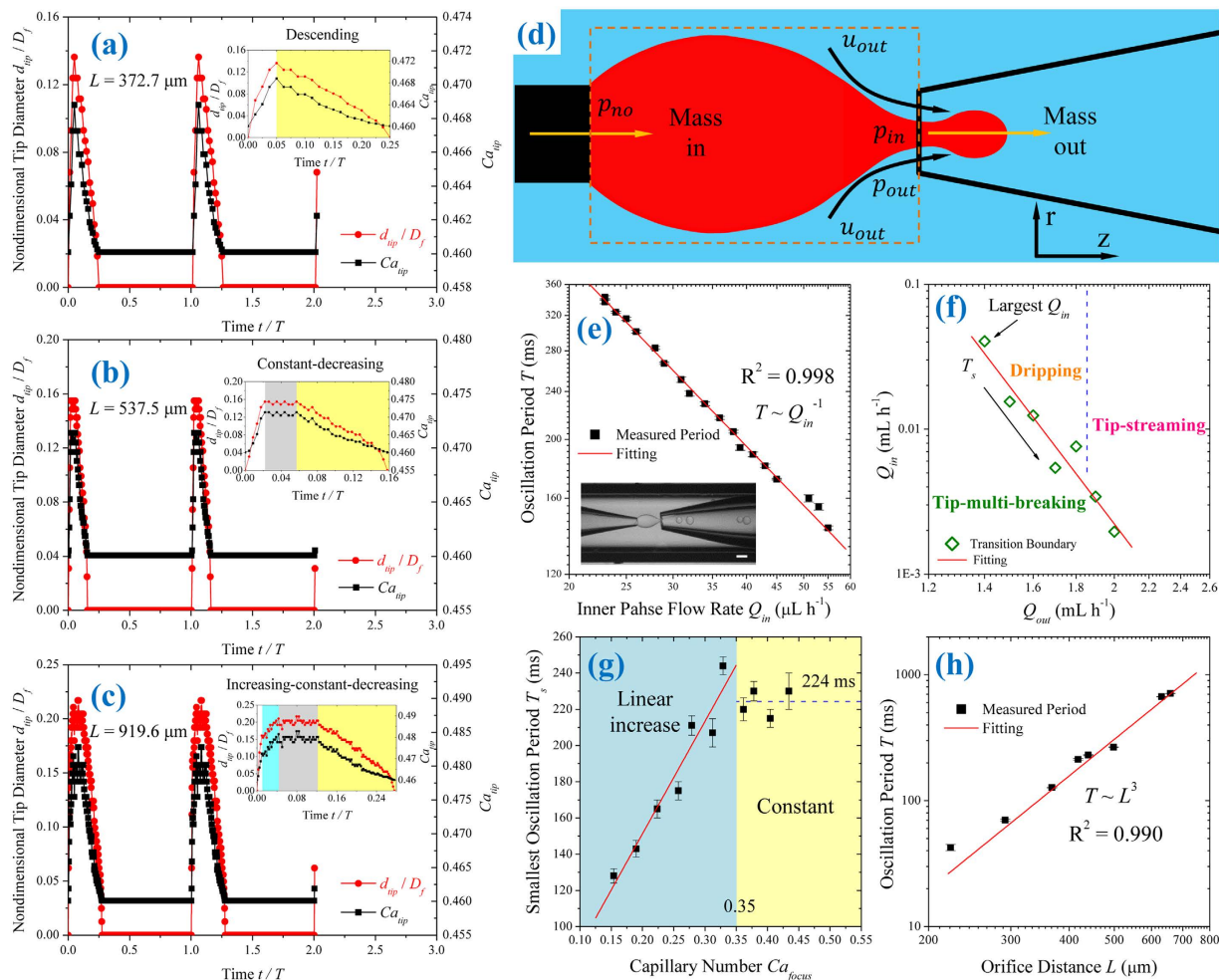


Figure 6. Oscillation of inner liquid tip. (a–c) Evolutions of nondimensional tip diameter d_{tip}/D_f (red) and capillary number Ca_{tip} (black) at the focusing orifice in tip-multi-breaking mode. $D_f = 210 \mu\text{m}$. (a) Descending behavior of d_{tip}/D_f and Ca_{tip} measured from Fig. 2a with $L = 372.7 \mu\text{m}$, and period $T = 160$ ms. (b) Constant-decreasing behavior of d_{tip}/D_f and Ca_{tip} measured from Fig. 2b with $L = 537.5 \mu\text{m}$, and period $T = 456$ ms. (c) Increasing-constant-decreasing behavior of d_{tip}/D_f and Ca_{tip} measured from Fig. 2c with $L = 919.6 \mu\text{m}$, and period $T = 834$ ms. The insets in (a–c) display the magnifications of tip evolution and temporal variation of Ca_{tip} . Since no droplet is generated during the initial period of increasing tip diameter (white area), we focus on the later stages of tip-diameter increasing (cyan area), constant tip-diameter (gray area) and tip-diameter decreasing (yellow area) where droplets are generated in (a–c). (d) Schematic of the control volume of the inner liquid tip confined by the left injection nozzle and right focusing orifice. p_{in} and p_{out} are pressures at the focusing orifice for inner and outer phases, respectively, while p_{no} is the inner fluid pressure at the injection nozzle. (e) Log-log plot of oscillation period T versus inner fluid flow rate Q_{in} , with $Q_{out} = 1.5 \text{ mL h}^{-1}$. Inset: snapshot of a droplet-sequence generation. Scale bar, $200 \mu\text{m}$. (f) Transition boundary of tip-multi-breaking mode in Q_{out} - Q_{in} plane; data adapted from ref. 37. (g) Minimum oscillation period T_s versus capillary number Ca_{focus} . (h) Log-log plot of oscillation period T versus Orifice distance L for fixed inner and outer flow rates. $Q_{in} = 4.5 \mu\text{L h}^{-1}$, $Q_{out} = 3 \text{ mL h}^{-1}$. The device used in (e,g,h) is case L with geometrical dimension shown in Table 1.

$[\pi(D_f^2 - d_{tip}^2)\gamma]$ ($\eta_{out} = 492.9 \text{ mPa s}$, $Q_{out} = 3.5 \text{ mL h}^{-1}$, $D_f = 210 \mu\text{m}$ and $\gamma = 30.07 \text{ mN m}^{-1}$ in our experiments) at the focusing orifice, and plot the temporal variation of Ca_{tip} in Fig. 6a–c. Due to the penetration of inner tip into the focusing orifice (see supplementary movie S2), a d_{tip} -increasing stage occurs at the very beginning of its evolution (white areas in the insets in Fig. 6a–c). No droplet is generated at this initial tip-growing stage because of the low viscous shear from the outer fluid. Afterwards, the droplet-generation takes place as the shear stress is large enough (cyan, gray and yellow areas in the insets in Fig. 6a–c). During the time period of droplet generation, the droplet size, d_{tip} and Ca_{tip} all vary in the form of descending (Fig. 6a), constant-decreasing (Fig. 6b), or increasing-constant-decreasing (Fig. 6c) as L increases.

Although the flow rates of both inner and outer fluids, the density-weighted area average of local flow field, are kept constant by syringe pumps, the local flow field can be unsteady around the focusing orifice, which induces the oscillation of the inner-fluid tip (Fig. 6a–c). To isolate the mechanism responsible for the tip oscillation, consider the normal stresses balance across the liquid-liquid interface⁴⁰ at the focusing orifice (Fig. 6d):

$$p_{in} - p_{out} + 2\eta_{out} \frac{\partial u_{r,out}}{\partial r} - 2\eta_{in} \frac{\partial u_{r,in}}{\partial r} = \gamma\kappa, \quad (12)$$

where p is pressure, $\eta_{out} \partial u_{r,out} / \partial r$ and $\eta_{in} \partial u_{r,in} / \partial r$ are, respectively, the outer and inner viscous stresses normal to the interface, κ is twice the mean curvature of the interface, estimated as $2/d_{tip}$ at the focusing orifice. Because the viscosity ratio is much smaller than unity ($\xi = 0.039 \ll 1$), $\eta_{in} \partial u_{r,in} / \partial r$ is negligible compared with $\eta_{out} \partial u_{r,out} / \partial r$. At fixed flow condition (Q_{in} , Q_{out} and L are constant for every single case in Fig. 6a–c), p_{out} and $\eta_{out} \partial u_{r,out} / \partial r$ can be assumed to be invariant during the tip thinning. Therefore, according to Eq. (12), p_{in} increases with the shrinkage of d_{tip} , for which κ is increased. When p_{in} is sufficiently large to compete with the pressure at the nozzle p_{no} (Fig. 6d), the inner tip is pushed upstream out of the focusing orifice. After recoiling, the tip is inflated by the inner fluid flow again and penetrates into the focusing orifice once it is large enough. The variation in inner pressure p_{in} is thus responsible for the tip oscillation. Further studies are needed to quantify this force analysis by experimentally measuring local pressure and flow fields inside microchannels, which is beyond our current capability of experiments.

The three distinct types of tip oscillation (Fig. 6a–c) correspond to different values of L . Eq. (12) accounts for this L -dependent behavior of d_{tip} . Q_{in} and Q_{out} are held constant in Fig. 6a–c, so that p_{in} and p_{out} can be assumed as constant at the maxima of d_{tip} . With enlarging L , the reduction in $\eta_{out} \partial u_{r,out} / \partial r$ leads to a decrease in the mean curvature κ . Consequently, as $\kappa = 2/d_{tip}$, the maxima of d_{tip} increases with L , as confirmed experimentally in Fig. 6a–c. For the largest L in Fig. 6c, the tip needs the longest time to fully develop into its maximal diameter d_{tip} after penetrating into the focusing orifice. An increasing stage of d_{tip} is therefore identified (cyan area in the inset in Fig. 6c). Afterwards, d_{tip} is temporarily stabilized (gray area) because of the transient mass balance of mass in and out from the tip (Fig. 6d), followed by the necking thinning (yellow area) due to inner fluid drainage (see supplementary Fig. S3 for experimental confirmation). Due to different time required for the tip to be fully developed, intermediate L in Fig. 6b holds d_{tip} constant for a while before tip thinning (inset in Fig. 6b), whereas the case with the smallest L in Fig. 6a has d_{tip} diminishing immediately once the tip diameter approaches the peak (inset in Fig. 6a).

We now show the variation of tip oscillation period T with inner flow rate Q_{in} , outer capillary number Ca_{focus} , and orifice distance L . In determining the relationship between T and Q_{in} , Q_{out} and L are fixed as constant. Since droplet sequence is fixed by Ca_{focus} , in this case, the volume V_s of one droplet sequence is invariant when Q_{in} varies. Thus, mass conservation, $Q_{in} = V_s/T$, suggests that T is inversely proportional to Q_{in} , verified by experiments in Fig. 6e. As Q_{out} changes, there is a smallest oscillation period T_s corresponding to the largest Q_{in} that enables the occurrence of tip-multi-breaking mode (Fig. 6f), as found in ref. 37. We show variation of T_s with Ca_{focus} in Fig. 6g where T_s is experimentally measured on the transition boundary between the tip-multi-breaking mode and the others shown in Fig. 6f. It shows that, for capillary number below 0.35, T_s increases linearly with Ca_{focus} . For Ca_{focus} above 0.35, T_s is however essentially independent of capillary number, and fluctuates around 224 ms in our experiments. This is due to the volume reduction of the tip as Ca_{focus} increases (see supplementary Fig. S4 for details). With both Q_{in} and Q_{out} fixed, an increase in L increases the tip volume V_t (dashed box in Fig. 6d) because $V_t \sim LD_t^2 \sim L^3$. As the volume V_s of one droplet sequence is proportional to the tip volume V_t , $V_s \sim V_t \sim L^3$; the mass conservation $Q_{in} = V_s/T$ leads to $T \sim V_s \sim L^3$ when Q_{in} is kept constant. This is experimentally confirmed in Fig. 6h.

Concluding Remarks

In conclusion, we have systematically studied the influence of expansion-contraction geometry on droplet breakup in capillary microfluidic devices, which we separate into two parts: at the injection nozzle where flow diverges, and at the focusing orifice where flow converges. We demonstrate that the variation of expansion-contraction dimension, tuned by orifice distance L , affects two local shear stresses at the focusing and injection orifices, and thus significantly influences the flow behavior. These influences include changes of droplet breakup location and breakup mode. At the injection nozzle, we derive a condition of critical L for breakup location transition by balancing local shear and capillary forces. Similarly, at the focusing orifice, we determine the local capillary number as a ratio of shear stresses to capillary effects, and quantify its relation to the size and number of droplets in tip-multi-breaking mode. The force balance on the liquid-liquid interface at the focusing orifice provides physical insight into the dynamical behavior of the tip oscillation. We have also experimentally examined the variation of the tip oscillation period with inner fluid flow, outer phase capillary number and orifice distance. Beyond the capillary devices used in this work, we expect our results to be also applicable to other two-phase microsystems involving expansion-contraction structures. Such fundamental understanding of droplet breakup in microfluidics involving expansion-contraction geometries would be useful in droplet generation, manipulation, and microfluidic device design.

References

1. Baroud, C. N., Gallaire, F. & Dangla, R. Dynamics of microfluidic droplets. *Lab Chip* **10**, 2032–2045 (2010).
2. Augustin, M. A. & Hemar, Y. Nano- and micro-structured assemblies for encapsulation of food ingredients. *Chem. Soc. Rev.* **38**, 902–912 (2009).
3. Whitesides, G. M. The origins and the future of microfluidics. *Nature* **442**, 368–373 (2006).
4. Haber, C. Microfluidics in commercial applications; an industry perspective. *Lab Chip* **6**, 1118–1121 (2006).
5. Patravale, V. & Mandawgade, S. Novel cosmetic delivery systems: an application update. *Int. J. Cosmetic Sci.* **30**, 19–33 (2008).
6. Kim, J. H. *et al.* Droplet microfluidics for producing functional microparticles. *Langmuir* **30**, 1473–1488 (2013).
7. Nunes, J., Tsai, S., Wan, J. & Stone, H. Dripping and jetting in microfluidic multiphase flows applied to particle and fibre synthesis. *J. Phys. D: Appl. Phys.* **46**, 114002 (2013).

8. Qiu, Y. *et al.* Microfluidic-based fabrication, characterization and magnetic functionalization of microparticles with novel internal anisotropic structure. *Sci. Rep.* **5**, doi: 10.1038/srep13060 (2015).
9. Wang, W., Zhang, M.-J. & Chu, L.-Y. Functional polymeric microparticles engineered from controllable microfluidic emulsions. *Acc. Chem. Res.* **47**, 373–384 (2013).
10. Demello, A. J. Control and detection of chemical reactions in microfluidic systems. *Nature* **442**, 394–402 (2006).
11. Song, H., Chen, D. L. & Ismagilov, R. F. Reactions in droplets in microfluidic channels. *Angew. Chem. Int. Ed.* **45**, 7336–7356 (2006).
12. Dittrich, P. S. & Manz, A. Lab-on-a-chip: microfluidics in drug discovery. *Nat. Rev. Drug Discov.* **5**, 210–218 (2006).
13. Kim, S. H., Shim, J. W. & Yang, S. M. Microfluidic multicolor encoding of microspheres with nanoscopic surface complexity for multiplex immunoassays. *Angew. Chem. Int. Ed.* **50**, 1171–1174 (2011).
14. Yuan, J., Zhao, X., Wang, X. & Gu, Z. Image decoding of photonic crystal beads array in the microfluidic chip for multiplex assays. *Sci. Rep.* **4**, doi: 10.1038/srep06755 (2014).
15. Nakano, M. *et al.* Single-molecule PCR using water-in-oil emulsion. *J. Biotechnol.* **102**, 117–124 (2003).
16. Khan, I. U. *et al.* Microfluidic conceived pH sensitive core-shell particles for dual drug delivery. *Int. J. Pharm.* **478**, 78–87 (2014).
17. Zhao, C.-X. Multiphase flow microfluidics for the production of single or multiple emulsions for drug delivery. *Adv. Drug Deliver. Rev.* **65**, 1420–1446 (2013).
18. Evans, H. M. *et al.* In situ formation, manipulation, and imaging of droplet-encapsulated fibrin networks. *Lab Chip* **9**, 1933–1941 (2009).
19. Chan, H. F. *et al.* Rapid formation of multicellular spheroids in double-emulsion droplets with controllable microenvironment. *Sci. Rep.* **3**, doi: 10.1038/srep03462 (2013).
20. Zhang, Y. X. & Wang, L. Q. *Microfluidics: Fabrication, Droplets, Bubbles and Nanofluids Synthesis*. (Springer, 2011).
21. Tan, Y.-C., Fisher, J. S., Lee, A. I., Cristini, V. & Lee, A. P. Design of microfluidic channel geometries for the control of droplet volume, chemical concentration, and sorting. *Lab Chip* **4**, 292–298 (2004).
22. Seemann, R., Brinkmann, M., Pfohl, T. & Herminghaus, S. Droplet based microfluidics. *Rep. Prog. Phys.* **75**, 016601 (2012).
23. Kong, T. T., Wang, L. Q., Wyss, H. M. & Shum, H. C. Capillary micromechanics for core-shell particles. *Soft Matter* **10**, 3271–3276 (2014).
24. Abbaspourrad, A., Carroll, N. J., Kim, S. H. & Weitz, D. A. Polymer microcapsules with programmable active release. *J. Am. Chem. Soc.* **135**, 7744–7750 (2013).
25. Link, D., Anna, S. L., Weitz, D. & Stone, H. Geometrically mediated breakup of drops in microfluidic devices. *Phys. Rev. Lett.* **92**, 054503 (2004).
26. Zhang, Y. X. & Wang, L. Q. Nanoliter-droplet breakup in confined T-shaped junctions. *Curr. Nanosci.* **7**, 471–479 (2011).
27. Khoshmanesh, K. *et al.* A multi-functional bubble-based microfluidic system. *Sci. Rep.* **5**, doi: 10.1038/srep09942 (2015).
28. Chen, H., Li, J., Shum, H. C., Stone, H. A. & Weitz, D. A. Breakup of double emulsions in constrictions. *Soft Matter* **7**, 2345–2347 (2011).
29. Bordoloi, A. D. & Longmire, E. K. Drop motion through a confining orifice. *J. Fluid Mech.* **759**, 520–545 (2014).
30. Choi, J.-H., Lee, S.-K., Lim, J.-M., Yang, S.-M. & Yi, G.-R. Designed pneumatic valve actuators for controlled droplet breakup and generation. *Lab Chip* **10**, 456–461 (2010).
31. De Menech, M., Garstecki, P., Jousse, F. & Stone, H. Transition from squeezing to dripping in a microfluidic T-shaped junction. *J. Fluid Mech.* **595**, 141–161 (2008).
32. Erb, R. M., Obrist, D., Chen, P. W., Studer, J. & Studart, A. R. Predicting sizes of droplets made by microfluidic flow-induced dripping. *Soft Matter* **7**, 8757–8761 (2011).
33. Kong, T. T., Liu, Z., Song, Y., Wang, L. Q. & Shum, H. C. Engineering polymeric composite particles by emulsion-templating: thermodynamics versus kinetics. *Soft Matter* **9**, 9780–9784 (2013).
34. Jeong, W.-C. *et al.* Controlled generation of submicron emulsion droplets via highly stable tip-streaming mode in microfluidic devices. *Lab Chip* **12**, 1446–1453 (2012).
35. Anna, S. L. & Mayer, H. C. Microscale tipstreaming in a microfluidic flow focusing device. *Phys. Fluids* **18**, 121512 (2006).
36. Lee, W., Walker, L. M. & Anna, S. L. Role of geometry and fluid properties in droplet and thread formation processes in planar flow focusing. *Phys. Fluids* **21**, 032103 (2009).
37. Zhu, P. A., Kong, T. T., Kang, Z. X., Tian, X. W. & Wang, L. Q. Tip-multi-breaking in capillary microfluidic devices. *Sci. Rep.* **5**, doi: 10.1038/srep11102 (2015).
38. Benson, B. R., Stone, H. A. & Prud'homme, R. K. An “off-the-shelf” capillary microfluidic device that enables tuning of the droplet breakup regime at constant flow rates. *Lab Chip* **13**, 4507–4511 (2013).
39. Christopher, G. & Anna, S. Microfluidic methods for generating continuous droplet streams. *J. Phys. D: Appl. Phys.* **40**, R319 (2007).
40. Batchelor, G. *An Introduction to Fluid Dynamics*. (Cambridge University Press, 1967).
41. Umbanhowar, P., Prasad, V. & Weitz, D. Monodisperse emulsion generation via drop break off in a coflowing stream. *Langmuir* **16**, 347–351 (2000).
42. Guillot, P., Colin, A., Utada, A. S. & Ajdari, A. Stability of a jet in confined pressure-driven biphasic flows at low Reynolds numbers. *Phys. Rev. Lett.* **99**, 104502 (2007).
43. Eggers, J. & Villermaux, E. Physics of liquid jets. *Rep. Prog. Phys.* **71**, 036601 (2008).
44. Taylor, G. The formation of emulsions in definable fields of flow. *Proc. R. Soc. Lond. A* **146**, 501–523 (1934).
45. Thorsen, T., Roberts, R. W., Arnold, F. H. & Quake, S. R. Dynamic pattern formation in a vesicle-generating microfluidic device. *Phys. Rev. Lett.* **86**, 4163 (2001).
46. Tomotika, S. On the instability of a cylindrical thread of a viscous liquid surrounded by another viscous fluid. *Proc. R. Soc. Lond. A* **150**, 322–337 (1935).

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Author Contributions

P.Z., T.K. and L.W. designed the project. P.Z., L.L., X.T. and Z.K. performed the experiments. P.Z. analyzed the data. P.Z., T.K. and L.W. wrote the manuscript. L.W. supervised the study. All authors commented on the paper.

Additional Information

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