



Ocean acidification reduces hardness and stiffness of the Portuguese oyster shell with impaired microstructure: a hierarchical analysis

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Abstract. The rapidly intensifying process of ocean acidification (OA) due to anthropogenic CO₂ is not only depleting carbonate ions necessary for calcification but also causing acidosis and disrupting internal pH homeostasis in several marine organisms. These negative consequences of OA on marine calcifiers, i.e. oyster species, have been very well documented in recent studies; however, the consequences of reduced or impaired calcification on the end-product, shells or skeletons, still remain one of the major research gaps. Shells produced by marine organisms under OA are expected to show signs of dissolution, disorganized microstructure and reduced mechanical properties. To bridge this knowledge gap and to test the above hypothesis, we investigated the effect of OA on juvenile shells of the commercially important oyster species, *Magallana angulata*, at ecologically and climatically relevant OA levels (using pH 8.1, 7.8, 7.5, 7.2). In lower pH conditions, a drop of shell hardness and stiffness was revealed by nanoindentation tests, while an evident porous internal microstructure was detected by scanning electron microscopy. Crystallographic orientation, on the other hand,

showed no significant difference with decreasing pH using electron back-scattered diffraction (EBSD). These results indicate the porous internal microstructure may be the cause of the reduction in shell hardness and stiffness. The overall decrease of shell density observed from micro-computed tomography analysis indicates the porous internal microstructure may run through the shell, thus inevitably limiting the effectiveness of the shell's defensive function. This study shows the potential deterioration of oyster shells induced by OA, especially in their early life stage. This knowledge is critical to estimate the survival and production of edible oysters in the future ocean.

1 Introduction

Edible oysters belonging to the genus *Magallana* have a complex life cycle, in which the free-swimming larvae attach onto a suitable hard substrate and then metamorphose into sessile juveniles within a few hours (Medaković et al., 1997;

Salvi and Mariottini, 2017). The oyster larval shell is primarily made of aragonite, a denser and mechanically stronger form of calcium carbonate (CaCO_3), compared to calcite which is a stable but mechanically brittle polymorphous CaCO_3 (Lawn and Wilshaw, 1993; Han et al., 1991). Upon metamorphosis, the fraction of calcite rapidly increases and becomes the main component in the juvenile and adult oyster shell (Medaković et al., 1997; Weiner and Addadi, 1997). The composition of the mineral and its organic matrix define a wide array of composites that relate to the mechanical strengths of the shell of each of these life stages (Lee et al., 2011). Early life stages of marine invertebrates, oysters included, are highly vulnerable to predators (Newell et al., 2007) and environmental stressors (Thomsen et al., 2015) when compared to the adult stages. Production of mechanically strong shells during larval and juvenile stages is essential to the post-larval phase because shell integrity and strength act as a protective barrier against shell-breaking and drilling predators.

Oceans currently absorb about a third of anthropogenic carbon dioxide (CO_2), which dissolves in seawater, forming carbonic acid, and increases the concentration of hydrogen ions, known as ocean acidification (OA). A study showed that the calcification rate of marine organisms, including oysters, is highly vulnerable to high carbon dioxide partial pressure ($p\text{CO}_2$; μatm) driven decreases in seawater pH (Feely et al., 2009; De Bodt et al., 2010). If the early life stages of edible oysters are vulnerable to near-future OA, then it could directly harm oyster survival and aquaculture production. Evidence of the negative effects of OA is, for example, the decline of spat production in oyster hatcheries on the west coast of the USA due to poorly calcified larval shells consequence of upwelled high $p\text{CO}_2$ waters (Barton et al., 2012). Previous studies on calcifying organisms suggest that OA not only reduces calcification rates, but also increases dissolution of formed shells in very high $p\text{CO}_2$ scenarios (Ries, 2011; Bednarsek et al., 2012). The decreased pH depletes carbonate ions necessary for CaCO_3 mineralization, as well as weakens marine organisms physiologically by causing acidosis and impairing internal pH homeostasis needed for optimal calcification (Dupont and Portner, 2013). Recently, an increasing number of studies have captured the importance of the mechanical properties of calcareous shells, the end product of calcification, under OA scenarios (Dickinson et al., 2012; Ivanina et al., 2013; Li et al., 2014; Fitzer et al., 2015; Collard et al., 2016; Tenniswood et al., 2016; Milano et al., 2016). For instance, it has been reported that the Pacific oyster, *Magallana gigas* (previously *Crassostrea gigas*), and the Eastern oyster, *Crassostrea virginica*, produced softer shells with reduced mechanical strength under OA conditions (Beniash et al., 2010; Dickinson et al., 2012). Despite these OA threats to oyster calcification process, studies are yet to demonstrate the structural organization of oyster shells under elevated CO_2 conditions. Importantly, modulating effects of

OA on the inherent relationship between shell structural and mechanical features is yet to be studied in detail.

This study is designed specifically to fill this gap in knowledge using the ecologically and economically important edible oyster *Magallana angulata* (previously *Crassostrea angulata*) also as a model species. Here, the quantitative relationship between microstructural and mechanical properties was examined using the newly formed juvenile oyster shells. Specifically, the effect of OA on this relationship was tested using three levels of environmentally and climatically relevant high CO_2 scenarios which induced decreased pH. As the calcitic foliated layer is the major shell structure for mechanical support in oysters (Lee et al., 2008), we examined its structural and mechanical properties by using a variety of characterization and imaging techniques such as scanning electron microscopy (SEM), electron back-scattered diffraction (EBSD) and nanoindentation tests. To further evaluate the overall shell integrity, we quantified shell density and shell density–volume ratio relationships using high-resolution micro-computed tomography scanning (micro-CT).

2 Materials and methods

2.1 Experimental design

Sexually matured adult oysters of the Portuguese oyster, *Magallana angulata*, were collected from the coastal area in Fujian, China (26°05′53.36″ N 119°47′45.81″ W), in the South China Sea on 29 July 2014. The adults were transported to the laboratory at the Swire Institute of Marine Science, University of Hong Kong. They were left to acclimatize in flow-through tanks in natural seawater at ambient conditions (31 salinity, 29 °C and $\text{pH}_{(\text{NBS})}$ 8.1) for a week. They were fed with a mixed algae diet (*Isochrysis galbana* and *Chaetoceros gracilis*). Sperm and eggs were obtained from more than 10 males and 10 females using the “strip spawning” method (Dineshram et al., 2013), and cultured under ambient conditions. A period of 24 h post-fertilization, embryos developed into D-shaped veliger larvae.

Four environmentally and climatically relevant pH levels (the control: pH 8.1; the low treatments: pH 7.8, 7.5 and 7.2) were selected as proxies to investigate the effect of CO_2 -driven OA on oyster shells. According to IPCC projections, the average pH of oceans (currently pH 8.1) is expected to drop to pH 7.8 and 7.5 by the year 2100 and 2300, respectively (Feely et al., 2009). Lowest pH treatment was included in this study to understand the impact of extreme environmental conditions in the coastal habitats of *M. angulata* (pH 7.2), which manifest naturally fluctuating pH levels up to -0.8 units due to river runoff and microbial respiration (Duarte et al., 2013; Thiyagarajan and Ko, 2012). Treatment levels of pH were maintained by bubbling filtered natural seawater with air enriched with CO_2 at the required con-

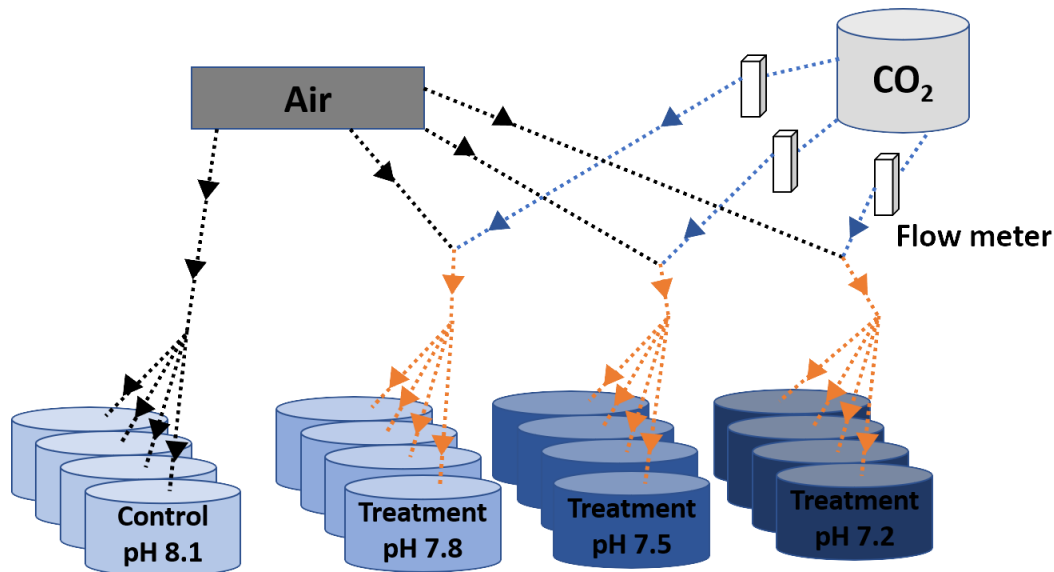


Figure 1. Scheme of the experimental system. Decreased pH conditions were obtained through bubbling CO₂-enriched air with the appropriate CO₂ concentration. Black and blue solid arrows indicate air and CO₂ flow respectively and orange arrows indicate CO₂-enriched air flow. The appropriate CO₂ concentrations in the injected CO₂-enriched air flow were controlled by using a dual variable area flow meter.

centrations using gas flow meters/controllers (Cole-Parmer, USA) (Fig. 1). Oyster larvae were raised from the D-shaped veliger stage to the juvenile stage under the four pH levels with four biologically independent replicates tanks for each treatment. D-shaped larvae (10 larvae mL⁻¹, 50 L replicate tanks, 1 μm FSW, 31 salinity, at 29 °C ± 2 °C) were reared until the pediveliger stage following methods described by Dineshram et al. (2013). After about 2 to 3 weeks, larvae attained competency for attachment and metamorphosis. Larvae were transferred from each 50 L replicate tank to 1 L replicate tanks containing plastic substrates coated with 7-day-old natural biofilms. Attachment and metamorphosis took place within 24 h. Attached oysters were reared in 1 L replicate tanks with the same pH level as before attachment for 35 days until collection. Larvae and juveniles were fed twice a day using a mixture of live *I. galbana* and *C. gracilis* (5–10 × 10⁶ cells mL⁻¹, 1 : 1 ratio). Seawater pH (NBS scale) and temperature were measured using a Mettler-Toledo (SG2) probe, and salinity was measured using a refractometer (ATAGO, S/Mill0E; Japan). The probe was calibrated using NIST buffers (pH = 4.01, 7.00, and 9.21; Mettler Toledo, GmbH Analytical CH8603 Schwerzenbach, Switzerland). In each culture, tanks levels of pH, temperature and salinity were measured daily. Daily measurements were firstly averaged within and among days per each replicate tank. Samples of seawater (50 mL) from each culture tank were collected every 4 days and poisoned with 10 μL of 250 mM mercuric chloride for total alkalinity (TA) analysis using the Alkalinity Titrator (AC-A2, Apollo SciTech's Inc., US). The TA measurement was standardized with a certified seawater reference material (Batch 106, Andrew G. Dickson, Scripps In-

stitution of Oceanography, USA). The carbonate system parameters, i.e. carbon dioxide partial pressure ($p\text{CO}_2$; μatm), carbonate ion concentration (CO_3^{2-} ; μmol kg⁻¹), calcite and aragonite saturation state (Ω_{Ca} , Ω_{Ar}), were calculated from pH, salinity, temperature and TA measured from each replicated tank ($n = 4$), using the CO2SYS software program (Pierrot et al., 2006) with equilibrium constants K_1 , K_2 and K_{SO_4} (Mehrbach et al., 1973; Dickson and Millero, 1987). The treatment level (mean ± SD; Table 1) was calculated using averages of the replicate culture tanks within each treatment ($n = 4$). On the 35th day post-metamorphosis, juveniles were collected and preserved in 75 % ethanol for the following analyses (Chan et al., 2012).

2.2 Shell microstructure analysis

The sessile juvenile oyster permanently cements the left valve of its shell to the substratum, whereas its right valve provides protection from predators and the environment. In this study, only the right valve was used in the shell analysis. The surface topography of the intact shell was examined under variable pressure at 30 kV using a scanning electron microscope (SEM; Hitachi S-3400N VP SEM, Hitachi, Japan). To examine sectional surface microstructures (MacDonald et al., 2010), shells were embedded in epoxy resin (Epoxy-Cure, Buehler) and sliced along the dorsal-ventral axis using a diamond trim saw blade. This allows for a more controlled comparison between the hinge region and the middle region of the shell. The hinge region (hereafter also referred to as “older shell”) is the part of the shell that is deposited first by the juvenile oyster, whereas the middle re-

Table 1. Seawater physico-chemical parameters in the experimental system.

Treatments/parameter	Control	pH 7.8	pH 7.5	pH 7.2
pH	8.14 ± 0.04	7.88 ± 0.02	7.46 ± 0.01	7.23 ± 0.01
Temperature (°C)	27.04 ± 0.14	27.02 ± 0.08	24.35 ± 0.12	27.50 ± 0.08
Salinity (psu)	31 ± 0.5	31 ± 0.5	31 ± 0.5	31 ± 0.5
TA (µequiv kg ⁻¹)	2053.77 ± 46.51	2032.63 ± 25.60	2061.50 ± 4.56	2091.37 ± 39.37
pCO ₂ (µatm)*	352.93 ± 11.04	861.37 ± 130.34	1997.23 ± 124.42	4091.73 ± 447.85
CO ₃ ²⁻ (µmol kg ⁻¹)*	175.66 ± 24.96	97.92 ± 16.38	48.82 ± 6.07	26.59 ± 4.72
Ω _{Ca} *	4.59 ± 0.25	2.43 ± 0.41	1.21 ± 0.15	0.66 ± 0.12
Ω _{Ar} *	3.01 ± 0.18	1.59 ± 0.28	0.79 ± 0.10	0.43 ± 0.08

Data are mean ± SD of the replicate culture tanks ($n = 4$) for the seawater physico-chemical parameters measured or calculated during the duration of the experiment: pH (National Bureau of Standards scale), temperature (°C), salinity (psu), TA (µequiv kg⁻¹), carbon dioxide partial pressure ($p\text{CO}_2$; µatm), carbonate ion concentration (CO₃²⁻; µmol kg⁻¹), calcite saturation state (Ω_{Ca}), and aragonite saturation state (Ω_{Ar}). Values were first averaged within and among days per each of the replicate culture tanks. Afterwards, the treatment mean was computed. * Parameters were calculated using the CO2SYS software program (Pierrot et al., 2006) with equilibrium constants K_1 , K_2 and K_{SO_4} (Mehrbach et al., 1973; Dickson and Millero, 1987).

gion (hereafter also referred to as “younger shell”) is the part of the shell that is deposited more recently. The edge region (Galtsoff, 1964), formed most recently, was not included in this study because it is too thin and fragile to handle. The sectioned surfaces were polished for 2 to 5 min using grit papers (P320, P800, P1200, P2500, and P4000) and etched for 20 s using 1 % acetic acid, and then washed with distilled water and air-dried. The sectioned resin blocks were mounted on aluminium stubs using carbon adhesive tape with the polished side up. The area surrounding the specimen was painted with silver to reduce charge build-up, and the sectioned surfaces were sputter-coated with 50 nm thick gold–palladium alloy. The shell microstructure was examined under SEM with an accelerating voltage of 5 kV using a LEO 1530 Gemini FSEM (Zeiss, Germany). The cross-sectional porosity of foliated laminated structure was calculated using ImageJ software by standardizing and converting an SEM image to thresholding where the non-diffracted regions of SEM images were defined as pores. The pore area was then calculated by using the ImageJ “Analyse Particles” feature due to the divergence in the size of pores. The pores area was sized with a confidence area greater than 0.001 µm². Three to four specimens from each treatment were randomly selected and examined ($n = 3\text{--}4$). All data were tested for normality of residuals, normality and homogeneity of variance before conducting analysis of variance (ANOVA). The Student–Newman–Keuls test was used to compare the means following one-way ANOVA.

2.3 Shell crystallographic orientation analysis

Shell crystallographic orientation was analysed by electron back-scattered diffraction (EBSD). Shells were prepared according to the above method, minus etching. The shell surfaces were ultra-polished for 4 min using cloths with 1 and 0.3 µm Alpha alumina powders and for 2 min using colloidal silica. In order to investigate both larva aragonite and ju-

venile calcite composition, an area throughout the sectional surface of the older hinge regions was selected. The EBSD analyses were carried out under low vacuum mode (~ 50 Pa) with a beam voltage of 20 kV using an FEI Quanta 200F with the stage tilted at 70° to examine back-scattered Kikuchi patterns (Perez-Huerta and Cusack, 2009). Diffraction intensity, phase and crystallographic orientation maps were produced using the OIM Analysis 6.2 software. Data was partitioned through two clean-up procedures to display grains with a confidence index greater than 0.1. Pole figures were used to illustrate the spread of crystallographic orientation (Perez-Huerta and Cusack, 2009). The colours in the crystallographic orientation maps and pole figures were used to quantify the crystallographic orientation. Two randomly selected specimens were examined per treatment.

2.4 Shell mechanical properties analysis

After SEM and EBSD analysis, the resin blocks were re-polished for 5 min using grit papers (P2500 and P4000) and for another 5 min using cloth with colloidal silica to remove the gold–palladium coating and etched shell surface. The mechanical properties of the polished longitudinal cross sections were determined by measuring the hardness (H) and stiffness (E) using load and displacement sensing nanoindentation tests (Perez-Huerta et al., 2007). Hardness and stiffness of foliated layers were measured in the older hinge and younger middle regions of the specimens used in the SEM analysis. The nanoindentation tests were carried out from the interior to the exterior shell in these regions at ambient temperature with a Hysitron TriboIndenter TI 900 (TI 900, Hysitron, MN, USA) equipped with a Berkovich indenter (with a half-angle of 63.5°). Indentations were made in each specimen using a 6–11 indent-per-row pattern and a maximum load of 2000 µN with valid contact depth of 16 to 184 nm. The hardness and stiffness from each indentation were obtained from the loading–unloading curve us-

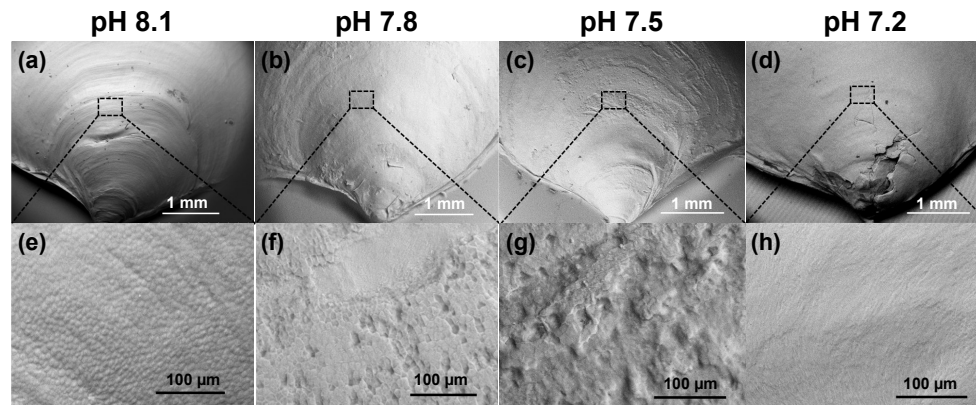


Figure 2. Scanning electron micrographs of 35-day-old juvenile *Magallana angulata* shells cultured at ambient or control pH 8.1 (a, e), treatment pH 7.8 (b, f), pH 7.5 (c, g) and pH 7.2 (d, h) were compared. Panels (a–d) show the low magnification tomography of the juvenile shells. Panels (e–h) show the enlarged view of the crystallite units (top view). (e) The prism units were arranged in compact prismatic structures at pH 8.1. (f) Prismatic arrangement was partially lost at pH 7.8. (g) A rough surface was observed, demonstrating a much lower level of organization at pH 7.5. (h) A smooth surface was observed with no prismatic arrangement due to dissolution by environmental seawater.

ing the Oliver–Pharr model (Doerner and Nix, 1986; Oliver and Pharr, 1992). Five to six specimens of each treatment were randomly selected for nanoindentation tests (one to two specimens per replicate tanks). Measurements per replicate tanks were calculated by firstly averaging the values among indentations per specimen, and then among specimens per replicates. Afterwards, the effect of decreased pH on the hardness and stiffness of juvenile oyster shells was compared by three to four replicate measurements ($n = 3–4$). All data were tested for normality of residuals, normality, and homogeneity of variance before conducting ANOVA. The Student–Newman–Keuls test was used to compare the means following one-way ANOVA.

2.5 Shell density analysis

The three-dimensional shell density maps, the overall shell density and the density–volume ratio relationships were obtained using a high-resolution micro-CT scanning system (SkyScan 1076, Skyscan, Kontich, Belgium) with a spatial resolution of $9\ \mu\text{m}$. Individual shells were placed in a small plastic container held securely in the chamber of the micro-CT scanner. Shell densities and the corresponding volume ratios of partial density were calculated by relative comparison using standardized phantoms used for bone density measurement in the analytical software CT-Analyser v 1.14.4.1 (SkyScan) (Celenk and Celenk, 2012). The three-dimensional digital data were converted from ~ 1000 two-dimensional layers using reconstruction software CT-Volume v 2.2.1.0 (SkyScan). Three randomly selected specimens were used per treatment ($n = 3$). The volume ratio with partial density ranges of 0 to $0.5\ \text{g cm}^{-3}$, 0.5 to 1 and $> 1.5\ \text{g cm}^{-3}$, and density of the treatment groups were compared with the controls by following one-way ANOVA. For the datasets that did

not meet the requirement of variance homogeneity, i.e. the volume ratio with a partial density range of 1 to $1.5\ \text{g cm}^{-3}$, Kruskal–Wallis tests were used to compare the effect of pH on these shell properties. For all other datasets, the Student–Newman–Keuls test was used to compare the means by following one-way ANOVA. Otherwise, Dunn’s test was used after the Kruskal–Wallis test. Linear regressions (volume ratio (%) = $b \times \text{density (g cm}^{-3}\text{)} + a$) were utilized to determine the relationships between shell density and the corresponding volume ratio; a is the y intercept and b is the scaling exponent of consumption. To compare slopes of the resulting linear models, analysis of covariance (ANCOVA) was performed by using \log_{10} transformed volume ratio as the dependent variable, pH levels as the independent variable, and shell density range as covariates. All data met the homogeneity of variance and normality assumptions of parametric tests. ANCOVA were implemented in R 3.3.2 using the statistical package Linear and Nonlinear Mixed Effects Models (R Core Team, 2013).

3 Results

3.1 Shell surface and internal microstructure

As shown by the SEM, decreased pH altered both shell topography (Fig. 2) and internal microstructure (Fig. 3). Mineral dissolution was prominent on the outer surface layers of shells under decreased pH. The shells of juveniles raised at pH 7.8 (Fig. 2b, f) and pH 7.5 (Fig. 2c, g) showed signs of dissolution or physical damage when compared to the controls (pH 8.1) (Fig. 2a, e). At the lowest pH of 7.2 with undersaturated calcite conditions, the outer prismatic layer was completely absent at the older hinge and younger mid-

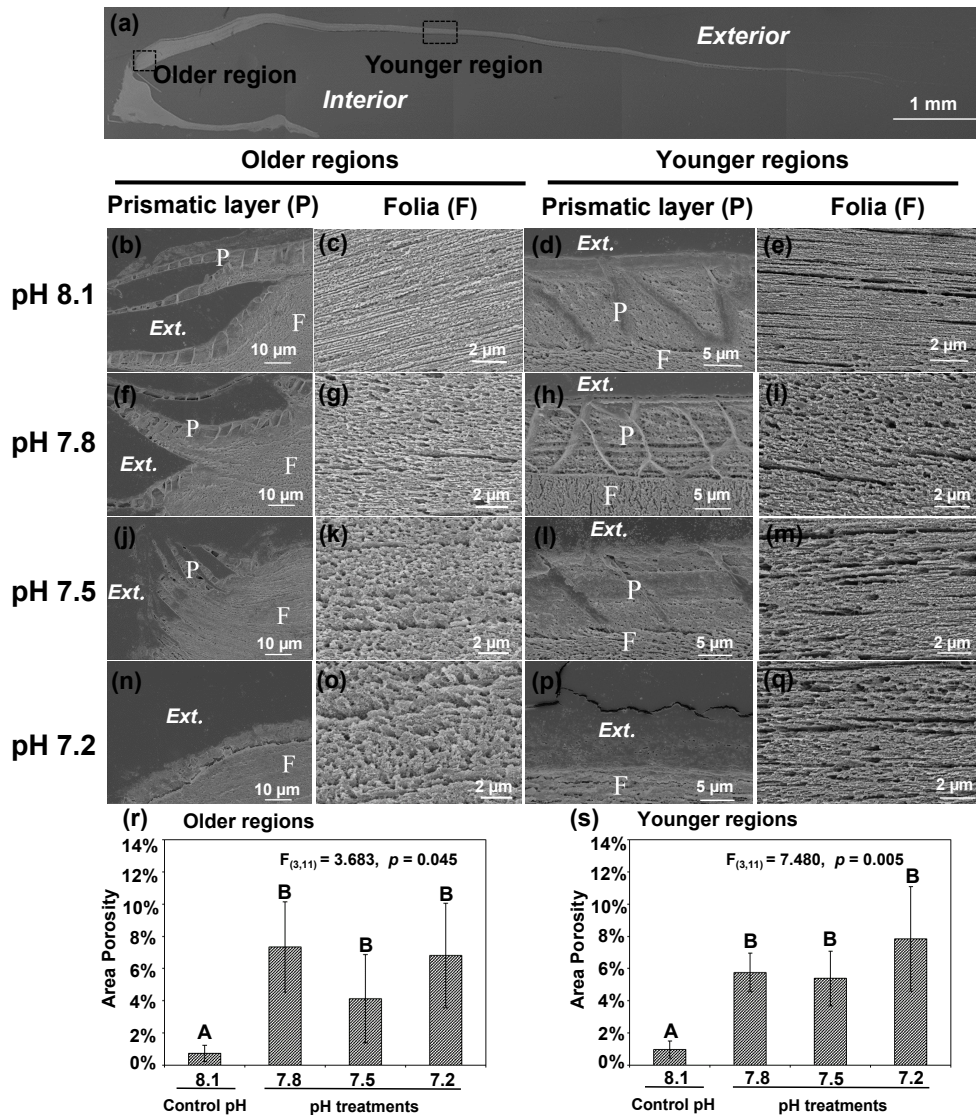


Figure 3. Microstructures were observed in the cross-sectional shell surfaces of 35-day-old juvenile *Magallana angulata*. Scanning electron micrographs were taken near the older hinge region (b, c, f, g, j, k, n, o) and the younger middle region (d, e, h, i, l, m, p and q). Panel (a) shows the scanning electron micrograph of the full shell cross-sectional surface. Second row: the prismatic layer (b, d) and tightly packed foliated structure (c, e) at pH 8.1. Third row: the prismatic layer (f, h) and the foliated structure with more and bigger pores (g, i) at pH 7.8 compared with at pH 8.1. Fourth row: the incomplete prismatic layer (j, l) and more porous foliated structure (k, m) at pH 7.5 compared with at pH 8.1. Fifth row: the prismatic layer was not detectable (n, p) with porous foliated structure (o, q) at pH 7.2. The porosity of foliated layers at the older (r) and younger regions (s) of the shell reared under control and low pH treatments. The mean values are presented in the bar chart (mean \pm SD, $n = 3$ –4). Annotations: P is the prismatic layer; F is the foliated layer.

dle regions of the shell (Fig. 2d, h). Though the overall calcitic foliated laminae's alignment was retained, those in the shells of controls (pH 8.1) were compactly arranged and well-ordered with minimal gaps between layers (Fig. 3c, e). In contrast, the foliated layers in shells under all three decreased pH treatments presented a more porous alignment in that the foliated laminae were less tightly packed and irregularly arranged (Fig. 3g, i, k, m, o and q). The area porosity of foliated layers increased significantly with decreasing pH

levels, regardless of older and younger shell (older region: $F_{(3,11)} = 3.683, p = 0.045$; younger region: $F_{(3,11)} = 7.480, p = 0.005$; Fig. 3r, s).

3.2 Crystallographic orientation

Electron back-scattered diffraction (EBSD) intensity mapping analysis showed diffraction patterns for both calcite and aragonite crystals of older hinge regions in the juvenile shells (Fig. 4). The crystallographic orientation maps

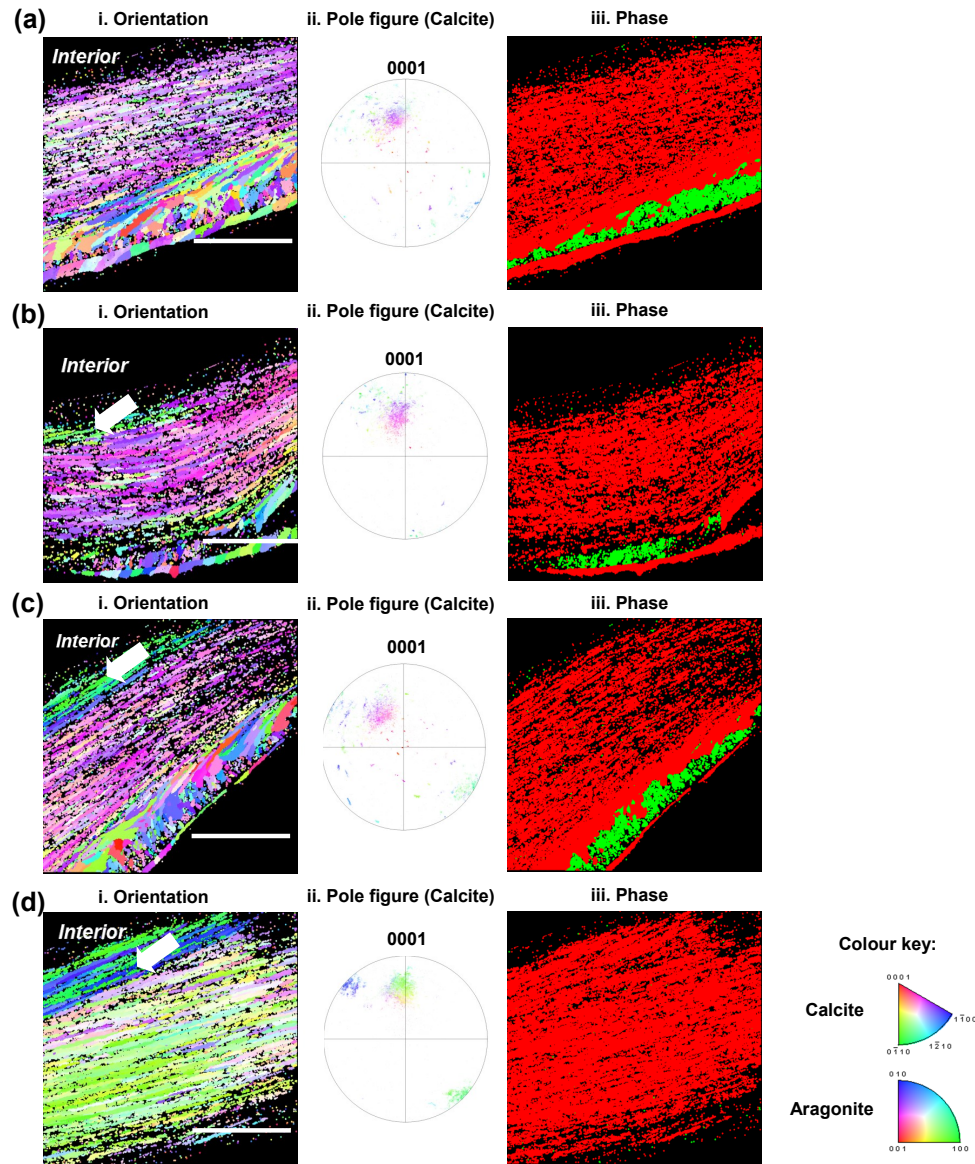


Figure 4. Electron back-scattered diffraction analyses of shells grown for 35 days at control pH 8.1 (a), treatment pH 7.8 (b), pH 7.5 (c) and pH 7.2 (d). Crystallographic orientation maps (left column) of calcite crystals in reference to the {0001} plane and aragonite crystals in reference to the {001} plane. Crystallographic planes of calcite are colour-coded according to the normal crystallographic direction shown in the colour key (Perez-Huerta and Cusack, 2009). Pole figures for calcite (centre column) corresponding to the crystallographic orientation maps with the same colour key. The right column shows the phase maps of calcite exhibited in red and aragonite in green. The white arrow denotes a change in colour of 5 to 10 marginal foliated laminates. Scale bar = 45 μm .

(Fig. 4 left column) showed changes in crystallographic orientation from the control (pH 8.1) to low pH conditions (pH 7.8, 7.5 and 7.2) as represented by colour change corresponding to the colour key (Fitzer et al., 2014). The spread of data points in pole figures (Fig. 4 centre column) highlighted the variation in crystallographic orientation between the juvenile oysters under the low pH and the control conditions. Though the foliated layers of shells under decreased pH showed colour variations within a limited area (~ 5 –10

foliated laminates) close to the interior, the majority of calcite crystal units showed uniform orientation, the same as those in the control (Fig. 4 left and centre columns). It was confirmed by the pole figures that the preferred crystallographic orientation of foliated layers was identical, resulting in the extent of the variation in crystal orientation of 40° regardless of pH treatments, corresponding to the colours in the orientation maps (Fig. 4 left column). But notably, there was an absence of aragonite in the shells formed under pH 7.2

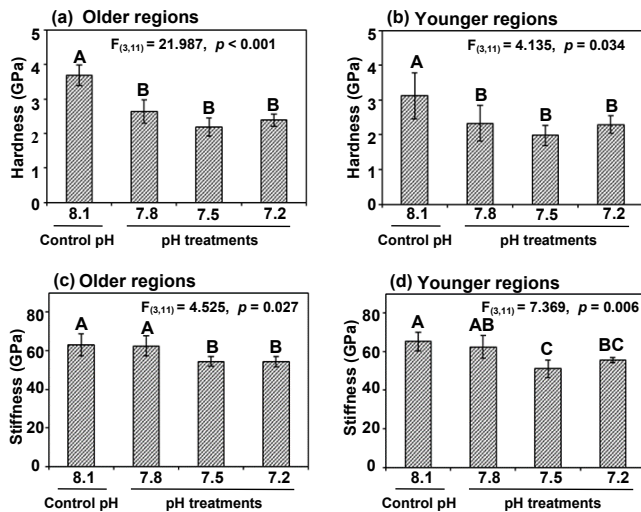


Figure 5. Shell mechanical properties in terms of hardness (a, b) and stiffness (c, d) with longer and shorter exposures in older hinge regions (a, c) and younger middle regions (b, d) in cross-sectional shell surfaces of *Magallana angulata* were compared. Data of mechanical properties are presented as mean \pm SD of three to four replicates ($n = 3$ or 4).

(Fig. 4 right column). Although the aragonitic crystals are not present in the most extreme treatment (pH 7.2), the overall crystallographic orientation of the calcitic fraction did not change between treatments.

3.3 Shell hardness and stiffness

Shell hardness was significantly reduced as treatment pH decreased, relative to control (older region: $F_{(3,11)} = 21.987$, $p < 0.001$; younger region: $F_{(3,11)} = 4.135$, $p = 0.034$). Similarly, shells at pH 7.5 and 7.2 had reduced stiffness compared to the controls (Fig. 5c, d) (older region: $F_{(3,11)} = 4.525$, $p = 0.027$; younger region: $F_{(3,11)} = 7.369$, $p = 0.006$). The reduced mechanical features due to decreased pH were observed in both the older hinge regions and younger middle regions (Fig. 5).

3.4 Shell density

Three-dimensional shell density maps (Fig. 6a–d), the overall shell density and the relationship of shell density–volume ratio by micro-computed tomography (micro-CT) showed an overall decrease of shell density with decreasing pH (Fig. 6e) ($F_{(3,8)} = 5.318$, $p = 0.026$). A similar decrease is visible in the linear regressions (volume ratio (%)) = $b \times$ density (g cm^{-3}) + a) in Fig. 6f. Volume ratios were decreased with the increased shell density in all pH treatments (ANCOVA; shell density, $F_{(1,263)} = 1253.14$, $p < 0.001$). There was an interaction between pH and shell density (ANCOVA; pH \times density, $F_{(3,263)} = 4.994$, $p = 0.002$), indicating that the effect of pH on the density–volume ratio relationship

was different. The lower scaling of consumptions at pH 7.8 (mean exponent -0.063), pH 7.5 (mean exponent -0.065) and pH 7.2 (mean exponent -0.062) versus the control pH level of 8.1 (mean exponent -0.052) indicates that the volume ratio of denser shell was reduced with decreased pH while the volume ratio of less dense shell was increased correspondingly (Fig. 6f). The three-dimensional shell density map (Fig. 6a–d) reinforces the effect of decreased pH on the density–volume ratio relationships. In the controls, shells were produced with denser minerals compared to shells in decreased pH (Fig. 6a). Shells at pH 7.8, pH 7.5 and pH 7.2 had larger proportions of lower shell density regions or “pores” (Fig. 6b–d). These pores were observed in the three-dimensional density maps as density values below the detection threshold (Fig. 6a–d). Classifying the shell volumes into four density categories, i.e. < 0.5 , 0.5 – 1 , 1 – 1.5 and $> 1.5 \text{ g cm}^{-3}$, showed that the proportions of high ($> 1.5 \text{ g cm}^{-3}$) and low ($< 0.5 \text{ g cm}^{-3}$) shell density areas were significantly different between pH treatments (Fig. 6a–d). The volume ratios of high density areas were significantly reduced in each pH treatment (pH 7.8, pH 7.5 and pH 7.2) when compared to the controls (pH 8.1) ($F_{(3,8)} = 4.856$, $p = 0.033$). Meanwhile, the volume ratios of low density areas ($< 0.5 \text{ g cm}^{-3}$) significantly increased in each of the lower pH treatments (pH 7.8, pH 7.5 and pH 7.2) compared to the controls ($F_{(3,8)} = 6.945$, $p = 0.013$). There were no significant differences in the volume ratios of the middle densities (0.5 – 1 g cm^{-3} : $\chi^2_{(2)} = 5.615$, $p = 0.132$; 1 – 1.5 g cm^{-3} : $F_{(3,8)} = 3.713$, $p = 0.061$) among treatments (Fig. 6a–d).

4 Discussion

This study provided new compelling information of structure–property relationships in calcareous shells of commercially important oyster species at different spatial scales and under a variety of environmentally and climatically relevant OA scenarios (i.e. different levels of decreased pH via $p\text{CO}_2$ increase). The revealed structural information and subsequent analysis of mechanical features in this study provided an important experimental basis for developing models to forecast the impact of ocean acidification on marine calcifying organisms. The rate of calcification of many marine organisms is expected to be significantly reduced in near-future oceans with a reduced pH of 7.8 due to OA (Ries, 2011; Bednarek et al., 2012; Duquette et al., 2017; Chatzinikolaou et al., 2017). This study observed the same calcification trend in *Magallana angulata* because OA not only depletes carbonate ions necessary for CaCO_3 mineralization, but it also metabolically weakens marine organisms through altered physiological processes, i.e. acidosis (Dupont and Portner, 2013). Importantly, this study provides strong evidence to support the argument that shells produced by oysters under OA suffer from dissolution with disorganized or impaired crystal orientation and microstructures, and reduced mechanical proper-

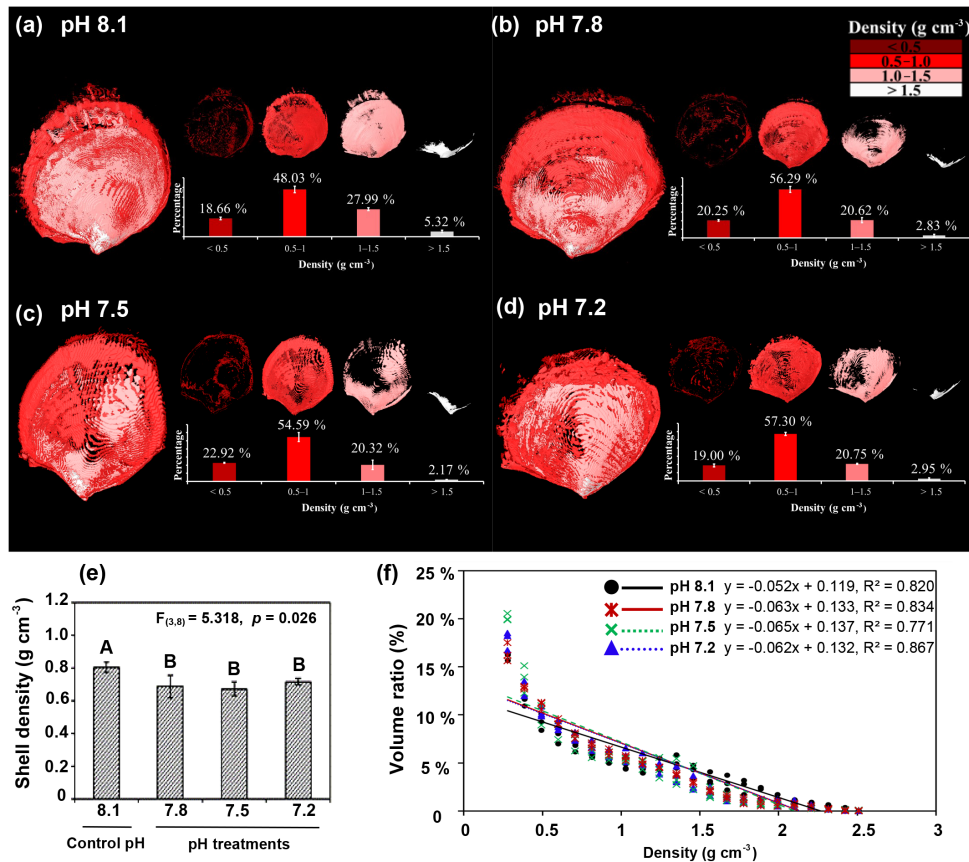


Figure 6. Effects of low pH on the shell density map (a–d), overall density (e), and shell density–volume ratio relationships (f) for the four experimental pH treatment groups were examined by micro-CT of shells of *Magallana angulata*. Three-dimensional reconstructions represent the density distribution of the shells produced in ambient or control pH 8.1 (a), treatment pH 7.8 (b), pH 7.5 (c) and pH 7.2 (d). The volume ratios of density categories of <math>< 0.5</math>, 0.5–1.0, 1.0–1.5, and > 1.5 \pm SD of three replicates ($n = 3$). (f) shell density–volume ratio relationships for the four experimental pH treatment groups of *C. angulata*. Regression lines for the three low pH treatments closely overlap and are partly obscured.

ties. The possible mechanisms and consequences underlying such negative effects of decreased pH on mechanics of shell structure are discussed in the following sections.

4.1 Effect of ocean acidification on shell mechanical features: a hierarchical analysis

In any given biologically formed material, mechanical properties at the macroscale generally depend on the composition of material components and materials’ microstructural features (Rodríguez-Navarro et al., 2002; Meng et al., 2018). In this study, oyster shell material is composed of two calcium carbonate polymorphs, calcite and aragonite. Oysters begin their life (larvae) with aragonite-based shells, and these are completely replaced by calcite in adult shells, though juvenile shells may retain a tiny portion of aragonite (Weiss et al., 2002). Calcite is relatively less soluble in seawater compared to aragonite regardless of environmental pH. Calcite is a relatively less soluble form of CaCO_3 in conditions with de-

creased pH when compared to aragonite (Lippmann, 1973). This chemical feature of calcite may have made it feasible for the juvenile oysters to successfully mineralize and retain a laminated calcareous structure, even under undersaturated CaCO_3 saturation levels, e.g. decreased pH 7.4 (Fig. 3).

Like the previously described oyster shell microstructure (Dauphin and Denis, 2000; MacDonald et al., 2010), the materials used in this study are composed of structurally organized layers. The bulk of the microstructure is characterized by foliated layers of crystal units organized in lamellae. In order to understand the modulating effect of environmental pH on the relationship between the shell structural and mechanical features, we have quantified the space or pore size between laminated layers within the folia. The decreased pH significantly increased size and quantity of the pores in the folia layer. The presence of such laminated folia with pores or gaps was an obvious consequence of decreased pH. However, the larvae were still capable of producing a new foliated layer under these treatments whilst at undersaturation (at

pH 7.2). Although the juvenile shells show signs of physical dissolution, the EBSD and porosity data suggest that the microstructure growth is impaired initially. This microstructural impairment was observed even under the near-future level of decreased pH 7.8, where the porosity was increased 10-fold (Fig. 3r). On the other hand, the preferred orientation of crystal units within the folia layer showed no difference in all low pH treatments, with the *c* axis of calcite units approximately perpendicular to the outer and inner shell surface. Thus, the significantly reduced hardness and stiffness of the foliated layer under decreased pH might be due to the impaired microstructure with significantly higher pore size and numbers.

Furthermore, we measured the impacts of decreased pH on whole shell density and thus on pores or gaps in foliated layers using micro-CT analysis. Notably, higher density shell volume reduces with decreasing pH. This result supports our finding on the effect of decreased pH on microscale structure and mechanical features in the folia. Calcite shell materials are brittle in nature, like egg shells or ceramics; therefore their resistance to deformation (or breaking force) is largely dependent on the stiffness parameter of the shell (Lawn and Wilshaw, 1993). Here, we found that both hardness and stiffness of the folia layer reduce with decreasing pH, which may have triggered shell fracture under simulated external attack. A folia layer with lower stiffness and hardness resulting from a porous laminated microstructure is expected to be highly vulnerable to predatory attack, even though the preferred orientation remains unaffected (Kemeny and Cook, 1986). In addition, the overall decrease of shell density detected by micro-CT analysis indicates the porous internal microstructure may occur throughout the juvenile shell. In other words, the juvenile oyster shell with impaired microstructural features is more prone to predator attack under the near-future level of decreased pH due to OA processes.

4.2 Effect of ocean acidification on shell microstructure and crystallography

The outermost prismatic layers of the older hinge and younger middle regions had completely disappeared when juvenile oysters were exposed to the extreme scenarios (pH of 7.2 and calcite undersaturation $\Omega_{\text{cal}} \approx 0.66$) (Figs. 2h and 3n, p). Undersaturated waters, with regards to calcite ($\Omega_{\text{cal}} < 1$), result in the dissolution of calcitic materials (Bednarsek et al., 2012; Lippmann, 1973). Similar impacts were observed in *Argopecten irradians* (pH 7.8 and pH 7.5) (Talmage and Gobler, 2010), *Mercenaria mercenaria* (pH 7.7) (Dickinson et al., 2013) and *Saccostrea glomerata* (pH 7.8 and pH 7.6) (Watson et al., 2009).

The juvenile oysters exposed to decreased pH exhibited a porous microstructure in foliated layers (Fig. 3). Firstly, this may be due to the decreased calcification rate resulted from the metabolic depression and/or energy shortage in the decreased pH conditions (Gobler and Talmage, 2014; Lannig et al., 2010). Secondly, it could be due to the dissolution of

newly formed minerals of the inner surface in the lower pH conditions (Melzner et al., 2011). Marine invertebrates' calcification has highly controlled mechanisms and remains to be explored by further studies (Krause-Nehring et al., 2011). Animals are capable of actively increasing the site of calcification by pumping protons out of the calcification site, thereby enabling calcium carbonate precipitation (Ries et al., 2010; Toyofuku et al., 2017). Supersaturated calcite conditions of oysters were found restricted to the shell edge including the outer mantle and the first intracellular nucleation site (Mount et al., 2004). Undersaturated calcite conditions may be maintained elsewhere in contact with the inner shell surface (Addadi et al., 2006; Thomsen et al., 2010). Therefore, in low pH conditions due to OA, these inner areas of newly formed minerals, which are precipitated as structural building blocks for the prismatic and foliated layers, may still be prone to dissolution. When the shell dissolution rate is faster than the mineralization rate, organisms tend to produce thinner and lighter (less dense) shells, resulting in an impaired shell microstructure. This may explain the multiple negative effects of reduced pH in our results, including porous and less dense foliated layers. Similarly, mussel shells grown in lower pH conditions (pH 7.65) showed inner shell surface dissolution (Melzner et al., 2011) and an impaired shell microstructure (Hahn et al., 2012), which is consistent with the results in this study. The crystallography of marine shells is the other important proxy for environmental stressors (Milano et al., 2017). The crystallographic orientation maps of foliated layers showed a preferred crystallographic orientation with $\sim 40^\circ$ of variation regardless of pH treatments, which concurs with the results of field samples (Checa et al., 2018). Compared to calcite, aragonite represents a small fraction of the oyster shells and is more soluble under decreased pH conditions (Fitzer et al., 2014), which could explain the absence of aragonite in the older hinge regions at pH 7.2 (Fig. 4 right column) is observed in this study. A similar absence of aragonite was also reported in mussel shells in high $p\text{CO}_2$ (1000 μatm) conditions (Fitzer et al., 2014). Nevertheless, aragonite dissolution may be very relevant for oyster shells during early life stages but not so much for adult shells. Therefore, it plays an insignificant role in shell mechanical properties of the calcite-predominant adult shells and thus the adult oyster survival.

4.3 Ecological implications and conclusion

Although previous studies showed that early larval life stages of several edible oyster species were relatively physiologically tolerant of near-future OA conditions (Dineshram et al., 2013; Ko et al., 2013, 2014; Thiyagarajan and Ko, 2012), this study shows that they are still vulnerable due to the effects decreased pH has on shell characteristics, like porosity, hardness and stiffness. A similar negative impact of OA on shell mechanical properties was reported in various marine calcifiers. For example, the pearl oyster, *Pinctada fucata*,

produced a 25.9% weaker shell after exposure to seawater at pH 7.8 (Welladsen et al., 2010). Decreasing shell mechanical properties in decreased pH conditions were also observed in *Mytilus californianus* (Gaylord et al., 2011), which produced 20% weaker shells under pH 7.8, *Mercenaria mercenaria* (Dickinson et al., 2013; Ivanina et al., 2013), which produced approximately 60% softer shells under pH 7.77 (16 salinity) and *Hydroides elegans* (Li et al., 2014), which produced 80% softer tubes under pH 7.8. However, the effects of increased carbon dioxide partial pressure ($p\text{CO}_2$) on shell mechanical properties are species-specific. Near-future conditions (pH 7.8) did not affect shell hardness in the sea urchin *Paracentrotus lividus* (Collard et al., 2016) or in the barnacle *Amphibalanus amphitrite* (McDonald et al., 2009). Furthermore, juvenile oysters of *C. gigas* significantly increased their shell strength and size as a compensatory adaptive response to low pH conditions (i.e. pH 7.8) (Wright et al., 2014), and the blue mussel, *Mytilus edulis*, produced a stiffer and harder calcite layer under increased $p\text{CO}_2$ conditions (i.e. pH 7.3) (Fitzer et al., 2015).

The long-term survival strategy of oysters with mechanically softer shells is yet to be studied. However, as shown in a recent study (Sanford et al., 2014), it appears that weaker shell structures will result in compromised defence abilities. Moreover, results from a recent study suggest that oysters with reduced and impaired calcification mechanisms have lower repair capabilities (Coleman et al., 2014). This hierarchical study revealed that the OA conditions may cause a deterioration of oyster shells and thus pose a serious threat to oyster survival and the health of coastal oyster reef structures in the near-future ocean. This biological effect of OA on shell structures and mechanical features should be incorporated in coastal oceanographic biophysical models to accurately project the survival of oysters in near-future coastal oceans which could be used for commercial shellfisheries to plan for sustainable growth under climate-change-induced acidification.

Data availability. All data related to the paper have been displayed in the text.

Author contributions. YM and TV conceived and designed the study. ZG and HY performed the nanoindentation test and contributed materials and analysis equipment. SCF and MC performed the EBSD analyses and contributed analysis equipment. YM and AU performed the CO_2 propagation experiment. YM and CL performed the SEM imaging. KWKY contributed materials and analysis equipment for micro-CT scanning. YM analysed the data and drafted the manuscript. VBSC and TV contributed to the revisions.

Competing interests. The authors declare that they have no conflict of interest.

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