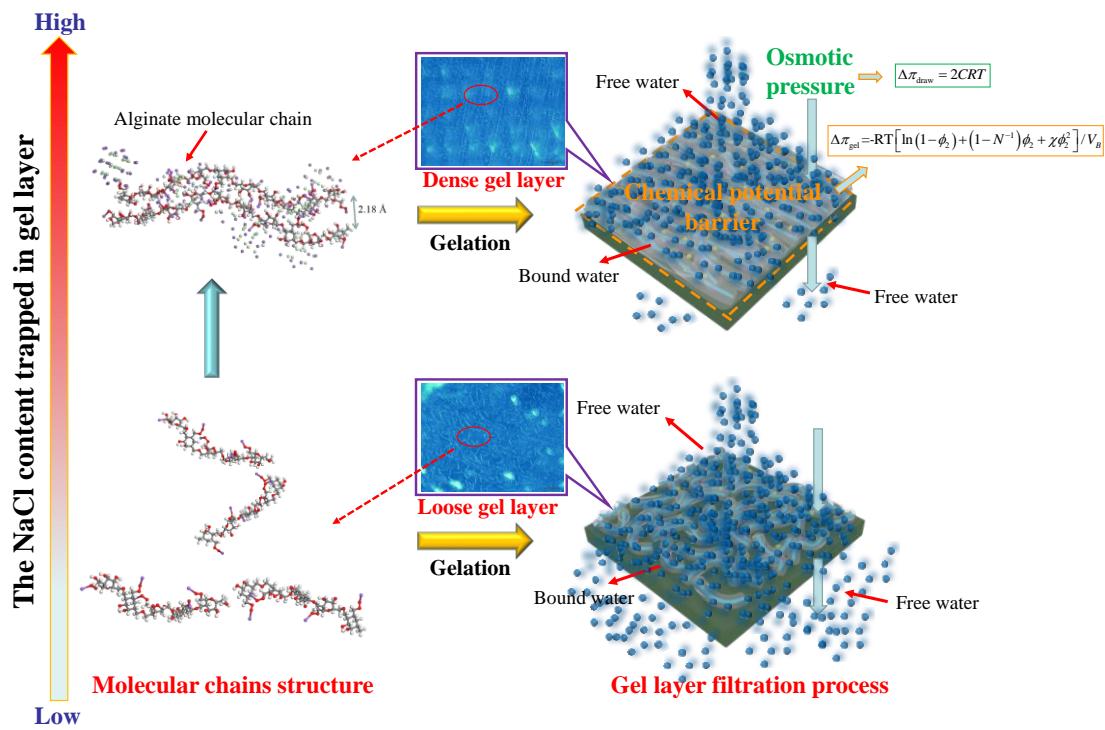


## Graphical Abstract



# 1 Novel molecular level insights into forward osmosis

## 2 membrane fouling affected by reverse diffusion of draw

### 3 solutions based on thermodynamic mechanisms

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13 **Abstract**

14 Forward osmosis membrane bioreactor (FOMBR) has a more intricate membrane  
15 fouling mechanism than MBR because of the **special** existence of the reverse diffusion of  
16 draw solution. The mechanisms of membrane fouling affected by reverse diffusion of draw  
17 solution and ion content in gel layer were investigated from thermodynamic perspective and  
18 molecular level in this study. Phase-contrast microscopy **non-invasively** observed that the  
19 molecular chain structure of the gel layer **containing** low content of reverse diffused solute  
20 (NaCl) was sparse, while the alginate molecular chains with high NaCl content were parallel  
21 and compact. Density functional theory (DFT) **further** simulated the formation mechanism of  
22 different alginate chain structures, that is, the reverse diffused solute shortened the distance  
23 between hydrogen bonds and reduced the interaction energy at the terminals of alginate  
24 chains. This proved that the NaCl trapped in alginate layer directly affected its structure.  
25 Heating experiments indicated that gel layer acted as a "chemical potential barrier" that  
26 prevented bound water from turning into free water. Accordingly, the "chemical potential  
27 barrier" described by Flory-Huggins theory was proposed, which was confirmed to account  
28 for the filtration of alginate gels affected by reverse diffusion of draw solution. To the best of  
29 our knowledge, this is the first membrane fouling study that explores the effect of reverse  
30 diffusion of draw solution on the "chemical potential barrier" of gel layer from perspective  
31 of molecular chain structure. This study investigated FOMBR fouling mechanism at  
32 molecular level and provided a new strategy for FOMBR system fouling quantification.

33 **Keywords**

34 FOMBR membrane fouling; reverse diffusion of draw solution; molecular chains  
35 observation; chemical potential barrier; density functional theory

36 **1. Introduction**

37 Stringent regulations and global water shortages place a great demand for efficient water  
38 treatment technologies [1-3]. For dealing to these intractable challenges, developing an efficient  
39 membrane bioreactor (MBR) technology is a realistic option [4]. FOMBR is viewed as a  
40 promising alternative of MBR since it demonstrates better removal efficiency and requires no  
41 external pressure [5-7]. Unfortunately, both MBR and FOMBR are subjected to membrane  
42 fouling, which increases the cleaning frequency and shortens the membrane life span [8-10].  
43 What is different between FOMBR fouling and MBR fouling is that reverse diffusion of draw  
44 solute exerts a seriously adverse impact on the fouling behavior and property of fouling layer  
45 [11-13]. Up to now, most of efforts have been focused to study the flux decline behavior, while  
46 limited attention has been paid to the effects of reverse diffusion of draw solution on the  
47 formation thermodynamic process and properties of fouling layer [11, 14]. Therefore, there is an  
48 imperative requirement for in-depth understanding of FOMBR fouling, especially the  
49 thermodynamic mechanisms of membrane fouling affected by reverse diffusion of draw solution  
50 [15].

51 Gel layer is generally considered as the principal cause to MBR fouling, and its specific  
52 filtration resistance (usually at level of  $10^{16}$ - $10^{17}$  m $\cdot$ kg $^{-1}$ ) can be a few orders of magnitude  
53 higher than that of cake layer even though the former has much thinner thickness [16-19]. In  
54 MBR, the formation of gel layer is a spontaneous process from the thermodynamic viewpoint  
55 because the macromolecular foulants (soluble microbial products (SMP) and extracellular  
56 polymeric substances (EPS)) tend to adsorb/mix as much water as possible to form a stable  
57 swelling gel layer [20-24]. Recent studies have shown that the gel layer formed by this process  
58 is non-porous, impervious and homogeneous. Thus, filtration through a gel layer is considered

59 to drag the bound water from gel layer to become free water in the permeate. It is required to  
60 overcome water chemical potential gap ( $\Delta\mu$ ) between permeate (high water chemical potential)  
61 and gel layer (low water chemical potential) when dragging bound water from gel layer to the  
62 permeate during filtration process [21, 25-27]. The osmotic pressure gap ( $\Delta\pi$ ) on both sides of  
63 FO membrane provides a driving force to overcome this water chemical potential gap ( $\Delta\mu$ ).  
64 While gel layer is equivalent to a “chemical potential barrier” which offsets a part of driving  
65 force, resulting in a great decrease in filtration efficiency [17, 25]. Moreover, the reverse  
66 diffused solute is partially trapped in gel layer. As a result, the ion content of gel layer in  
67 FOMBR is much higher than that in MBR, leading to significant changes in the formation  
68 process and properties of gel layer [28]. Singh et al. suggested that the interaction of gels with  
69 ion increased the viscosity of gel layers [29]. Derjaguin-Landau-Verwey-Overbeek (DLVO)  
70 theory has pointed out that the high ionic strength compresses the electric double layer and  
71 weakens the electrostatic repulsion force between gel molecules, thereby contributing to the  
72 formation of a dense gel layer [30-32]. FOMBR fouling involves a complex interaction between  
73 reverse diffused solute and foulants, which is far more complicated than pressure-driven MBR  
74 fouling. Therefore, further research on the interaction mechanism of the binary complex system  
75 composed of reverse diffused solute and foulants is of great significance to improving our  
76 insights into FOMBR fouling [5, 33].

77 The purpose of this study is to explore the underlying mechanism of FO membrane fouling  
78 affected by reverse diffusion of draw solution. Phase-contrast microscopy was used to observe  
79 the original molecular chain structure in fresh gel layer formed in draw solutions with different  
80 ionic strength. The molecular chain structure of gel layer with different content of reverse  
81 diffused solute were accurately predicted by the classical quantum mechanics method called

82 DFT. The **simulation** results might help further explain formation **mechanisms** of different  
83 molecular chain structures of gel layer. Finally, the membrane fouling of gel layer affected by  
84 reverse diffusion of draw solution was elucidated by combining molecular chain structure of gel  
85 layer with Flory-Huggins lattice theory from the perspective of water chemical potential. This  
86 research will not only provide insights into FO membrane fouling mechanism but also blaze a  
87 new way in the investigation into membrane fouling.

88 **2. Materials and methods**

89 **2.1. Samples preparation**

90 It is generally accepted that fouling layer is a mixture of many compounds, and the fouling  
91 behavior of real fouling layer could be satisfactorily simulated by a model foulant when  
92 studying the mechanism of membrane fouling [27, 34-36]. In previous studies, sodium alginate  
93 (SA) is frequently used as a model compound for gel layer in membrane fouling study due to its  
94 availability and high ability to form a gel layer, and sludge floc suspensions were most  
95 commonly used to form a cake layer [20, 37-39]. The alginate solutions (1.0 g/L) were prepared  
96 by dissolving 2.0 g SA powder into 2000 mL deionized (DI) water with continuous magnetic  
97 stirring for 2 h until the SA powder completely dispersed. The sludge floc suspensions (1.0 g/L)  
98 were obtained by centrifuging the active sludge at 5000 g for 5 min and sediment was diluted  
99 with a buffer solution (2 mM K<sub>3</sub>PO<sub>4</sub>, 4 mM KH<sub>2</sub>PO<sub>4</sub> and 10 mM KCl) at pH 7 [38]. The active  
100 sludge obtained at stable operation stage of a lab-scale FOMBR. The alginate solutions and the  
101 sludge floc suspensions samples were used for fouling tests.

102 **2.2. Batch fouling tests**

103 Batch FO fouling tests were conducted to compare the fouling behavior between **model gel**

104 foulants (alginate solutions) and sludge floc suspensions affected by reverse diffusion of draw  
105 solution. As shown in Fig. 1, an external FOMBR was used to explore the membrane fouling  
106 behaviors affected by reverse diffusion of draw solution in this paper. The membrane module is  
107 placed outside the bioreactor, and the model gel foulants and sludge floc suspensions flow  
108 through the FO membrane in a circulating manner. To study the effect of reverse diffusion of  
109 draw solution on FO membrane fouling, same volume (500mL) of foulants (prepared according  
110 to [section 2.1](#)) were filtered in draw solution with different ionic strength (1, 2, 3, 4 mol/L NaCl  
111 solutions). By plotting the relationship between flux and permeate volume instead of the flux  
112 versus time [\[40\]](#). Moreover, to eliminate the continuous dilution of draw solution caused by the  
113 penetration of water during the filtration process, baseline experiments were carried out before  
114 batch fouling tests.

115 The weight of permeate was recorded in real-time with a balance connected to a computer,  
116 and each fouling test was conducted 3 times. The working temperature maintained at about 25 °C.  
117 According to literature, the membrane flux ( $J$ ,  $\text{L}/(\text{m}^2 \cdot \text{h})$ , LMH) could be calculated by [\[41\]](#):

$$118 \quad J = \frac{\Delta V}{A_m \Delta t} \quad (1)$$

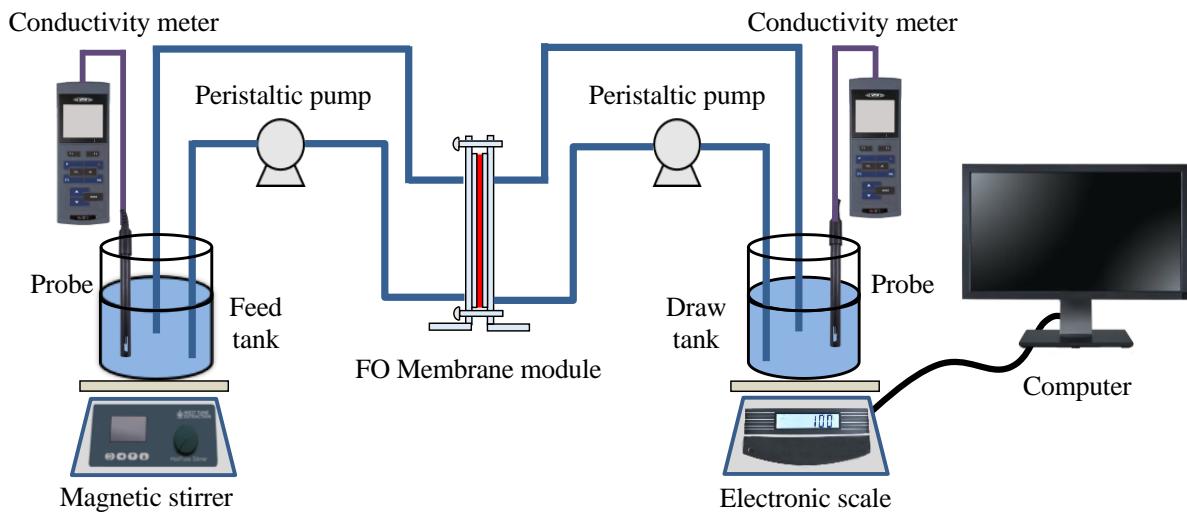
119 where  $\Delta V$  is the permeate volume (L),  $\Delta t$  is the time interval (h), and  $A_m$  is the FO  
120 membrane effective area ( $\text{m}^2$ ). The conductivity values of feed solution and draw solution were  
121 obtained at intervals of 60 s, and the conductivity value is then used to calculate ion  
122 concentration. The osmotic pressure ( $\pi$ ) of the solution was calculated based on the empirical  
123 equation [\[42\]](#):

$$124 \quad \pi = 4.5032C^2 + 43.6426C \quad (2)$$

125 where  $C$  is the ion concentration (mol/L).

126 The osmotic pressure gap ( $\Delta\pi_{\text{drive}}$ ) between draw solution and feed solution provides a driving  
127 force for FO membrane filtration:

128 
$$\Delta\pi_{\text{drive}} = \pi_{\text{draw}} - \pi_{\text{feed}} \quad (3)$$



129

130 **Fig.1.** Schematic of the lab-scale external FOMBR.

131 **2.3. Novel non-invasive microscopic observation method of gel layer microstructure**

132 In this study, all alginate fouling layer samples used for subsequent experimental  
133 characterizations were formed by filtering 500 mL of foulants. Fresh gel layers are thin and  
134 transparent, and their microstructure cannot be directly observed by a conventional optical  
135 microscope [8]. Moreover, other observation methods such as SEM require freeze-drying the  
136 fouling layer for 24 hours and spray gold in vacuum before characterization. This process  
137 unavoidably destroy the original structure of gel layer, which necessitates the development of a  
138 novel method that can observe the microstructure of a transparent material without harming its  
139 original structure [43]. Phase-contrast microscope is a better alternative choice. It has been

140 extensively used to observe unstained cells owing to its capability of checking out the extremely  
141 fine structure in transparent materials through the diffraction and interference of light without  
142 damaging samples [44]. In this study, microscope images of fresh alginate gel layers were  
143 observed by an Olympus IX83 inverted microscope (Olympus America Inc., PA, USA) in phase  
144 (ph) mode.

145 **2.4. Analytical methods**

146 **2.4.1. Chemical analyses**

147 Fresh alginate layers were further dried in a vacuum freeze dryer (LC-10LCN, China) for  
148 24h before their functional groups were characterized by Nicolet iN10 Fourier Transform  
149 Infrared Spectrometer (FTIR). The spectra in the range of 4000-500  $\text{cm}^{-1}$  were collected by  
150 overhead and removable multi reflective attenuated total reflection accessory (ATR). The  
151 chemical composition of alginate layers was determined by an X-ray photoelectron spectrometer  
152 (XPS) (ESCALAB 250Xi, Thermo Fisher Scientific, USA) with Al-K X-ray as the excitation  
153 source.

154 **2.4.2. Thermal stability of alginate gel layers**

155 Thermal stability of alginate gel layers was measured by a thermogravimetric analyzer  
156 (Q200, TA Instruments, Germany). Fresh gel samples were placed in an  $\text{Al}_2\text{O}_3$  crucible and  
157 heated to 600°C at a heating rate of 10°C/min under the nitrogen atmosphere. Moreover, heating  
158 experiments were conducted to investigate the ability of gel layers to bind with water. Fresh gel  
159 layers were placed in a constant temperature oven (WRF-Q800, China) for 1 hour at 35°C, and  
160 their weight changes were recorded.

161 **2.4.3. Inductively coupled plasma optical emission spectrometer (ICP) analysis**

162 As the reverse diffused draw solute is trapped in gel layer, it is bound to produce a certain  
163 osmotic pressure. Gel layer samples were completely dissolved in deionized (DI) water by  
164 magnetically stirring for 2 hours. An inductively coupled plasma optical emission spectrometer  
165 (Nex ION 300D, PerkinElmer, USA) was utilized to detect the concentration of NaCl that was  
166 trapped in gel layers, followed by the calculation the osmotic pressure of the ions trapped in  
167 alginate gel layers according to [Eq.2](#).

168 **2.5. DFT calculations**

169 Density functional theory (DFT) is a classical quantum mechanical method used to study  
170 the properties of molecules, which is one of the most commonly used methods on computational  
171 chemistry [\[45, 46\]](#). The DFT computational of molecular structure and binding energy can  
172 provide accurate prediction for the interaction between reverse diffused solute (NaCl) and  
173 sodium alginate chains. Quantum chemical calculations were performed with the Gaussian 09  
174 program by using DFT/B3LYP hybrid method. In this study, a short alginate chain model  
175 containing 10 uronic units and a long alginate chain model containing 20 uronic units were  
176 established, and each alginate chain contained six uronic rings [\[47\]](#). The 6-31 G basic set was  
177 used to optimize the molecular geometry and yield energy data. Based on quantum chemistry  
178 calculation results, the effect of reverse diffusion of draw solution on the interactions between  
179 alginate chains was explored at the molecular level.

180 **2.6. Flory-Huggins lattice theory**

181 The lattice theory proposed by Paul Flory and Maurice Huggins is a mathematical model  
182 that describes the thermodynamics of polymer solutions [\[48, 49\]](#). In previous studies,

183 Flory-Huggins lattice theory was proposed to describe the water chemical potential change in  
184 the formation process of gel layer [47, 50]. According to Flory-Huggins theory, the change in  
185 Gibbs free energy during filtration can be described by [48, 51]:

186 
$$\Delta G = RT \left[ n_1 \ln \phi_1 + n_2 \ln \phi_2 + n_1 \phi_2 \chi \right] \quad (4)$$

187 where  $n$  and  $\phi$  represent mole number and volume fraction, respectively; the subscripts of 1  
188 and 2 mean solvent and solute, respectively;  $\chi$  is the Flory-Huggins interaction parameter. As  
189  $\phi_1 + \phi_2 = 1$ , the chemical potential change ( $\Delta\mu$ ) of this process is given as follows:

190 
$$\Delta\mu = RT \left[ \ln(1 - \phi_2) + (1 - N^{-1}) \phi_2 + \chi \phi_2^2 \right] \quad (5)$$

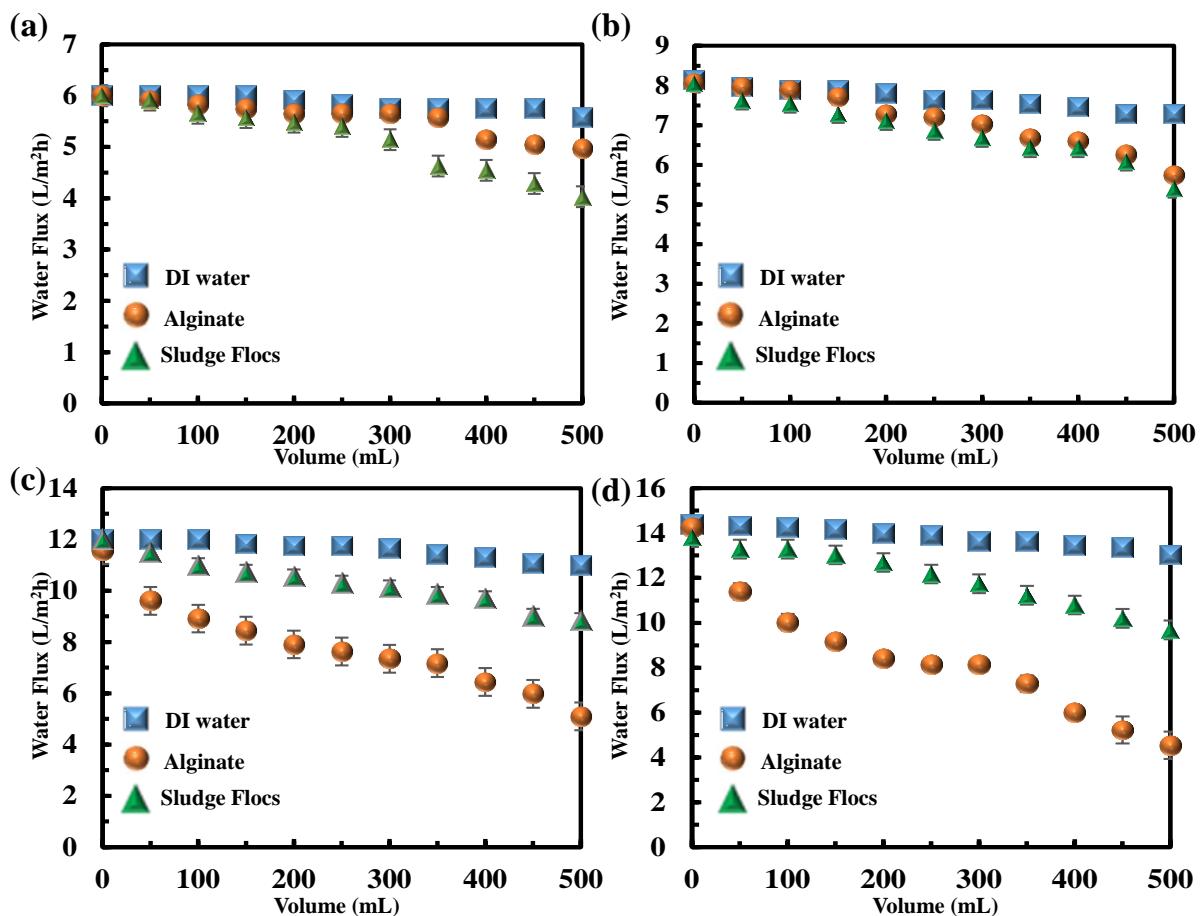
191 where  $N$  is the degree of polymerization of cross-linked polymer, which is usually assumed to  
192 infinity.

193 **3. Results and discussion**

194 **3.1. Effects of reverse diffusion of draw solution on filtration behavior**

195 [Fig. 2](#) shows the effects of reverse diffusion of draw solutions with different ionic strength  
196 (1M, 2M, 3M, 4M NaCl solutions) on the filtration behavior of different feed solution (DI water,  
197 sludge floc suspensions, and alginate solutions). In this study, same volume and same mass  
198 concentration of foulants (500mL, 1.0 g/L) were filtered in each filtration tests, which therefore  
199 the draw solution was diluted to same degree. As indicated by [Fig. 2](#), the DI water flux decline  
200 ratio approximated to 9.0% for all cases (8.3%, 9.0%, 8.5% and 9.5%, respectively), which was  
201 mainly ascribed to dilution of draw solution [52]. As for sludge floc suspensions, the water flux  
202 decline ratio was near 30% under all the circumstances (27.8%, 29.8%, 27.5% and 29.9%,  
203 respectively), suggesting that the membrane fouling caused by sludge floc suspensions are

204 moderate. Obviously, the reverse diffusion of draw solution has negligible effects on the  
 205 filtration behaviors of DI water and sludge floc suspensions. In contrast, the flux decline ratio of  
 206 alginate solutions increased dramatically (17.2%, 28.8%, 56.9% and 69.2%, respectively) with  
 207 the increase of the ionic strength of draw solutions. Thus, it is considered that that the filtration  
 208 behavior of alginate solutions was considerably affected by reverse diffusion of draw solution,  
 209 and extremely severe alginate fouling occurred at draw solutions with high ionic strength.

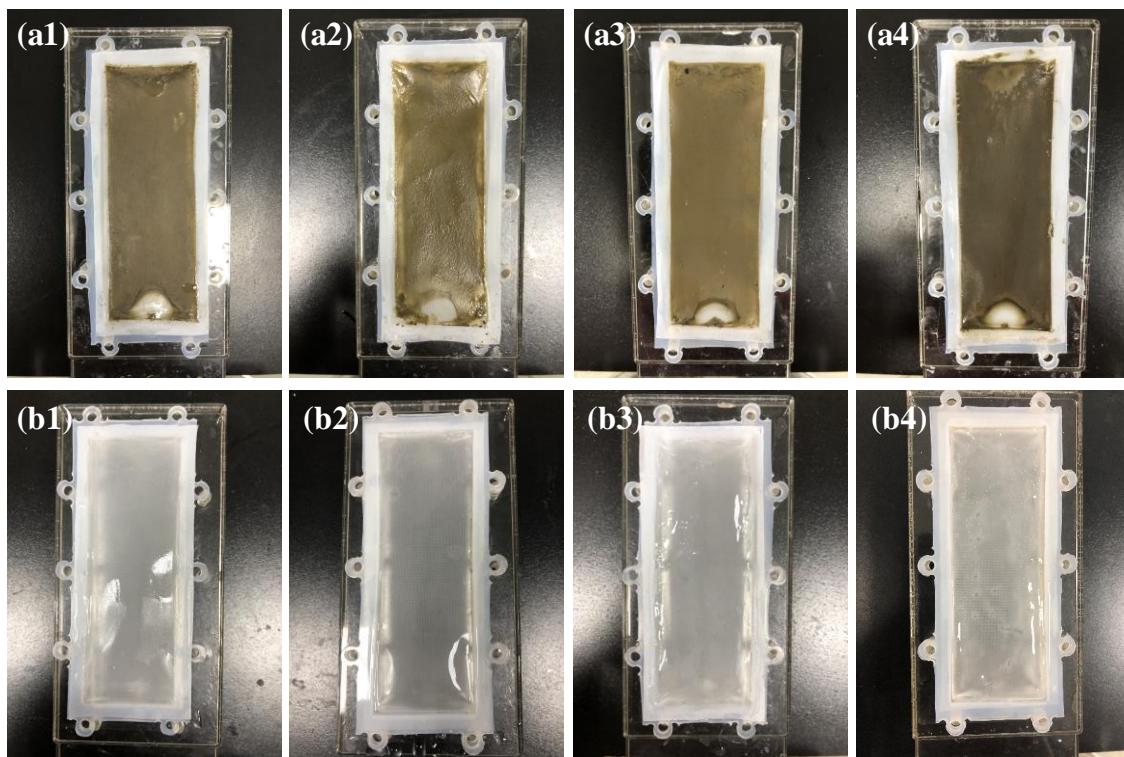


210

211 **Fig. 2.** Effects of reverse diffusion of draw solutions with different ionic strength ((a) 1 M,  
 212 (b) 2 M, (c) 3 M and (d) 4 M NaCl solutions) on the filtration behavior of DI water, sludge floc  
 213 suspensions and alginate solutions.

214 **Fig. 3** shows the optical images of fouling layers formed on FO membrane surface in draw  
 215 solutions with different ionic strength, the appearance of these layers is completely different. It

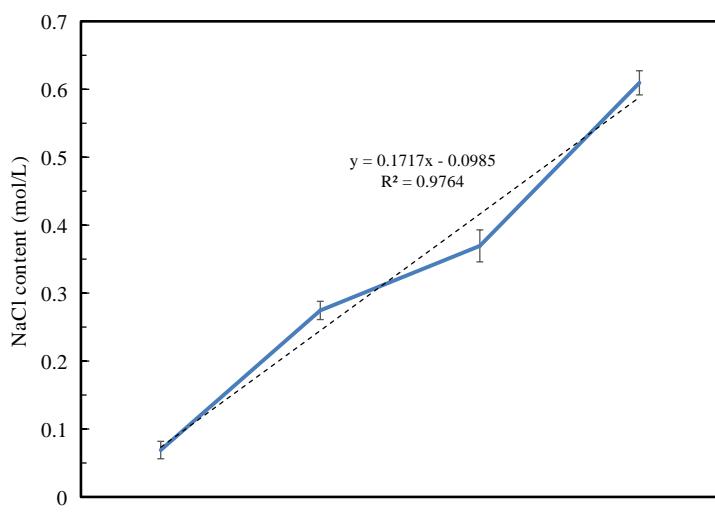
216 can be seen from [Fig. 3\(a1\)-\(a4\)](#) that the cake layers formed by sludge floc suspensions are  
217 loose and powdery. The weights of cake layers are  $0.316\pm0.015$  g,  $0.293\pm0.025$  g,  $0.299\pm0.023$  g,  
218 and  $0.306\pm0.011$  g, respectively, which are not significantly different. Contrary to sludge floc  
219 suspensions, the alginate retained on FO membrane surface shows typical gelling properties ([Fig.](#)  
220 [3\(b1\)-\(b4\)](#)). The weights of gel layers gradually increase ( $0.237\pm0.013$  g,  $0.666\pm0.020$  g,  
221  $1.139\pm0.170$  g, and  $1.518\pm0.110$  g, respectively) with the rising concentration of draw solutions.  
222 It should be noted that the filtration experiments were performed under the conditions of same  
223 volume (500mL) and same mass concentration (1.0 g/L) of foulants. Thus, it is considered that  
224 the reverse diffusion of draw solution significantly affects the FO membrane fouling behavior of  
225 alginate gel foulants, which directly causes severe FO membrane fouling. Overall, the main  
226 effect of reverse diffusion of draw solution on FO membrane fouling is that its significant  
227 impacts on gel layer fouling rather than cake layer fouling.



228  
229 **Fig. 3.** Optical images of foulant layers in draw solutions with different ionic strength. (a)

230 Sludge flocs layer, and (b) alginate layer. 1, 2, 3, and 4 refer to 1M, 2M, 3M, and 4M NaCl  
231 solutions, respectively.

232 **Fig. 4** shows the content of NaCl trapped in different SA layers formed by same volume of  
233 SA solutions filtered through draw solutions with different ionic strength. The content of NaCl  
234 trapped in SA gel layers increased almost linearly from  $0.0690 \pm 0.01283$  mol/L to  
235  $0.6090 \pm 0.01778$  mol/L, indicating that reverse diffusion of draw solution directly affected the  
236 NaCl content in gel layers. Due to the inevitable reverse diffusion of draw solution during FO  
237 process, the ion content in FOMBR gel layer was much higher than that in MBR gel layer. A  
238 recent study showed that, adding a small amount of NaCl (0-250mM) to the alginate solution  
239 significantly changed the fouling layer from a sparse structure to a dense structure and greatly  
240 improved the filtration resistance, which indicated that NaCl played a vital role in the structure  
241 change of the alginate fouling layer [28]. Thus, the critical effects of ion content in gel layers on  
242 FOMBR fouling behavior should be highlighted [28, 53, 54].

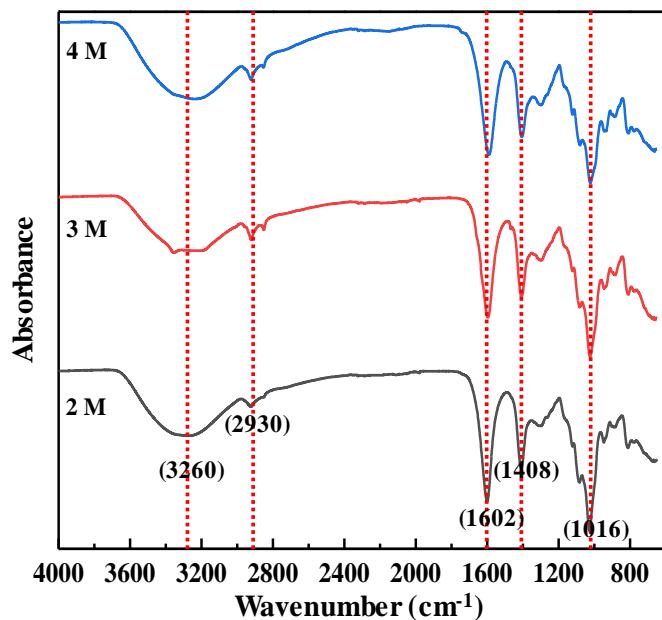


243  
244 **Fig. 4.** The content of NaCl trapped in different SA layers due to the reverse diffusion of draw  
245 solutions with different ionic strength.

246 **3.2. Characterizations of alginate layers formed in draw solutions with different ionic  
247 strength**

248 **3.2.1. Functional groups analyses**

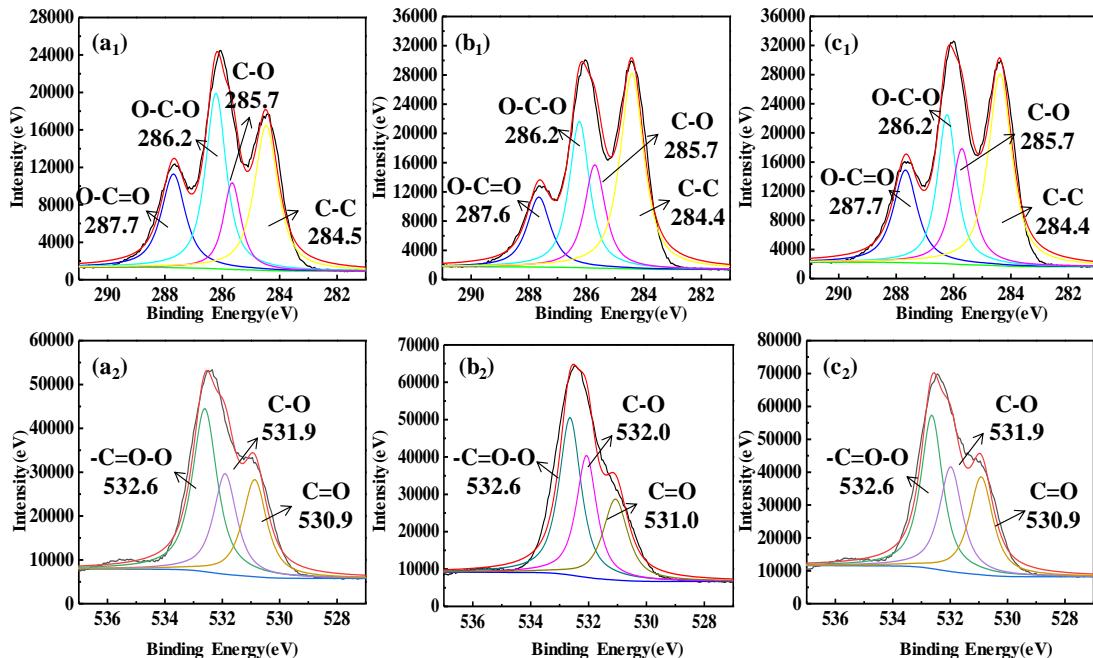
249 [Fig. 5](#) represents the ATI-FTIR spectra of alginate gel layers formed in three draw solutions  
250 with different ionic strength. Five typical alginate characteristic peaks and the corresponding  
251 functional groups are  $\sim 3260 \text{ cm}^{-1}$  ( $-\text{OH}$ ),  $\sim 2930 \text{ cm}^{-1}$  ( $-\text{C}-\text{H}$ ),  $\sim 1602 \text{ cm}^{-1}$  ( $>\text{C}=\text{O}$ ),  $\sim 1408$   
252  $\text{cm}^{-1}$  ( $-\text{COO}-$ ) and  $\sim 1016 \text{ cm}^{-1}$  ( $-\text{C}-\text{O}$ ), respectively [55]. The FTIR spectra of all alginate  
253 layers are similar to each other, which probably suggests that the FTIR technique may not detect  
254 difference in the functional group compositions of these samples.



255  
256 [Fig. 5](#). FTIR spectra of alginate layers formed in draw solutions with different ionic strength.

257 [Fig. 6](#) shows the XPS spectra of C1s and O1s of gel layers formed in draw solutions with  
258 different ionic strength. In [Fig.6\(a1\)-\(c1\)](#), C1s signals at 287.7, 286.2, 285.7, and 284.5 eV  
259 correspond to the O-C=O, O-C-O, C-O and C-C bonds, respectively [56]. The O1s signals at  
260 532.6, 531.9, and 530.9 eV represent the -C=O-O, C-O and C=O bonds, respectively, as

261 indicated by Fig. 6(a2)-(c2). It can be seen that the proportion of groups only changed slightly,  
 262 and the binding energy of the C and O chemical bonds of the alginate did not move. Some  
 263 studies suspected that different fouling behaviors were caused by the difference in the functional  
 264 groups of foulants [57]. These chemical characterization results clarified that the reverse  
 265 diffusion of draw solution or the ion content in gel layers could not significantly affect the  
 266 functional group of alginate gel layer. In other words, the tiny differences in functional groups  
 267 may not be the underlying cause for the dramatically different fouling behavior (Figs. 2 and 3).  
 268 Further research is required.

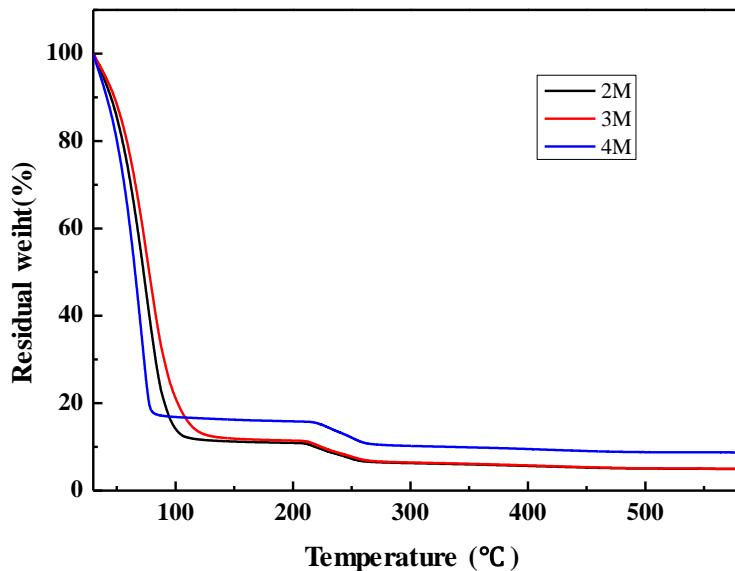


269  
 270 **Fig. 6.** XPS spectra of alginate layers formed in draw solutions with different ionic strength.  
 271 (a) 2 M, (b) 3M and (c) 4 M NaCl solutions, and the post-fixes of 1, 2 refer to C1s and O1s  
 272 spectra, respectively. Compared with FTIR spectra, XPS reveals more detailed information on  
 273 the functional groups and chemical bonds.

274 **3.2.2. Gel layer properties analyses**

275 Fig. 7 shows the thermogravimetric analysis (TGA) results of different gels. It can be seen

276 that the TGA curve of these samples can be divided into two stages. When the temperature  
277  $<100$  °C, the weights of all samples decreased sharply, which was ascribed to the evaporation of  
278 water. When the temperature rose above 200 °C, a slight decrease of the weights was observed  
279 due to the evaporation of the alginate. [Fig. 7](#) shows that there is difference in the thermal  
280 stability of alginate gel layers formed in three draw solutions with different ionic strength.  
281 According to reports, the thermal stability of alginate is affected by ions, which may be caused  
282 by the changes in the ability of gel layers to bind with water resulted from the cross-linking  
283 effect [[58-60](#)].

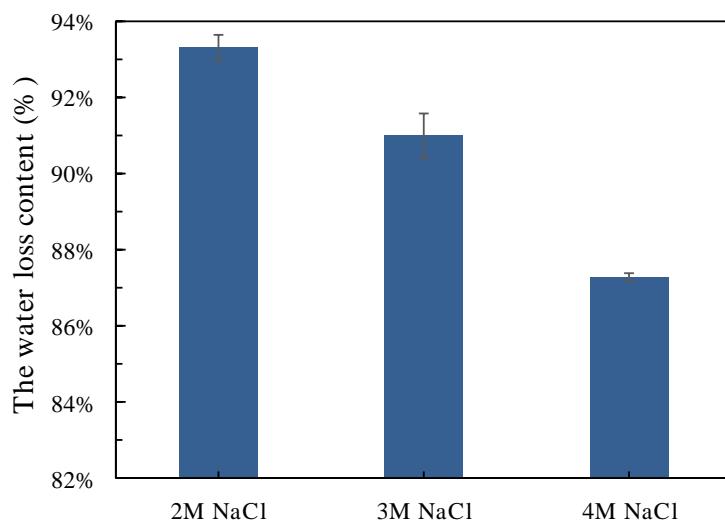


284

285 **Fig. 7.** TGA analysis of alginate gels formed in draw solutions with different ionic strength.

286 The ability of gel layers to bind with water can be verified by heating experiments [[61](#)].  
287 The chemical potential of bound water in gel layers is much lower than that of free water, and  
288 this chemical potential gap needs to be overcome in order to drag the bound water in gel layers  
289 to the free water side [[27](#)]. [Fig. 8](#) shows the amount of water loss in different gel layers under  
290 the same heating conditions, and the water loss decreased linearly with the NaCl content in gel  
291 layers. This result indicated that the bound water was more difficult to drag out from dense gel  
292 layers that contained a large amount of NaCl. In other words, considerable energy was

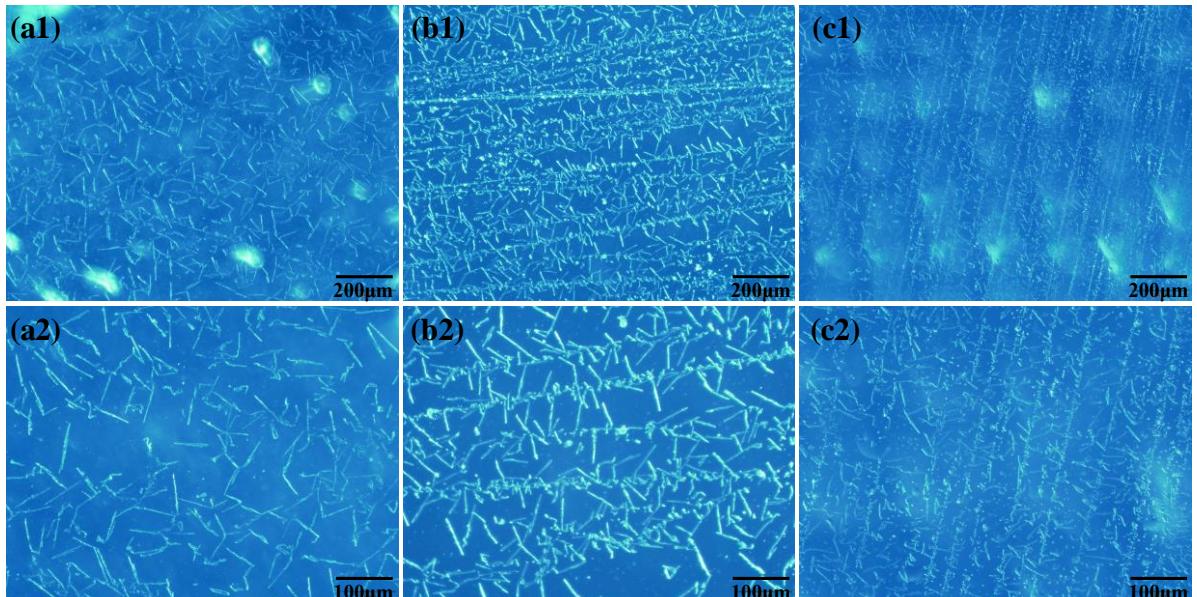
293 demanded in order to overcome the huge chemical potential gap to drag out bound water from  
294 dense gel layer with high NaCl content during the filtration, resulting in **significant** decline in  
295 FO membrane filtration efficiency [28]. Therefore, the effect of reverse diffusion of draw  
296 solution on the filtration behavior of alginate gel layer could be attributed to the ability of gel  
297 layer to bind with water. Previous studies reported that the ability of gel layer to bind with water  
298 was directly determined by the gel layer structure [27, 28, 62].



299  
300 **Fig. 8.** Water loss of alginate gel layers formed in draw solutions with different ionic strength  
301 under heating conditions (heating time: 1 h, oven temperature: 35 °C).

302 **Fig. 9** presents the phase-contrast microscope images of molecular chain structures of fresh  
303 alginate gel layers formed in draw solutions **with** different ionic strength. To the best of our  
304 knowledge, this paper is the first direct microscopic observation study on the molecular chain  
305 structures of fresh fouling layers [8]. It is clear that the NaCl trapped in alginate gel layer had an  
306 appreciable effect on the arrangement of alginate chains in gel layer (Fig. 9). A lot of short  
307 alginate chains were sparsely distributed in alginate gel layer with low NaCl content (Fig. 9(a)),  
308 making the water in gel layer unable to be firmly bound. This was because short alginate chains  
309 moved away from each other due to the mutual exclusion of the polar carboxylic acid groups in  
310 alginate chains with low NaCl content [27, 28]. Short alginate chains cross-linked and produced

parallel and compact long alginate chains in [Figs. 9\(b\) and \(c\)](#), indicating that the large amount of NaCl trapped in alginate gel layers promoted the growth and symmetrical distribution of alginate chains. Long and dense alginate chains contributed to the formation of a compact molecular chain structure, which acted as the skeleton of gel layer and bound with the free water in feed solution to form [a gel layer](#) [\[47\]](#). Hence, the results of [Fig.8](#) and [Fig.9](#) suggest that due to change of the molecular chain structure, the bound water is more difficult to drag out from the dense gel layer containing a large amount of NaCl. In view of this, fouling behavior of alginate solution in draw solution with different ionic strength ([Figs. 2 and 3](#)) could be reasonably attributed to the change in the structure of alginate chains in gel layer [\[38, 50, 63\]](#).



**Fig. 9.** Phase-contrast microscopy images of the molecular structure in alginate fouling layers formed in draw solutions with different ionic strength. (a) 2M, (b) 3M and (c) 4M NaCl solutions, and the 1, 2 represent 200 times and 400 magnifications, respectively.

Accordingly, exploring the forces that control this molecular structure transformation might provide essential fouling mechanisms for FO processes. In this paper, DFT analysis was used to [simulate the interactions of alginate chains with varied content of NaCl](#). DFT simulation was performed to further determine the possible molecular structure changes ([Fig. 9](#)) of alginate gel

328 layers [46].

329 **3.3. DFT simulation of alginate chains cross-linking behaviors**

330 A computational model of two short alginate chains was constructed to explicate the  
331 intermolecular interaction [20, 47]. Fig. 10 shows that the energy of two alginate chains in  
332 vertical (-341.93 kcal/mol) and linear (-308.51 kcal/mol) states was very close and low, which  
333 illustrates when NaCl was absent in gel layers, the alginate chains can be arranged freely due to  
334 the mutual exclusion of the polar carboxylic acid groups in alginate chains. DFT simulation  
335 results could well explain the distribution of alginate chains in Fig. 9(a).

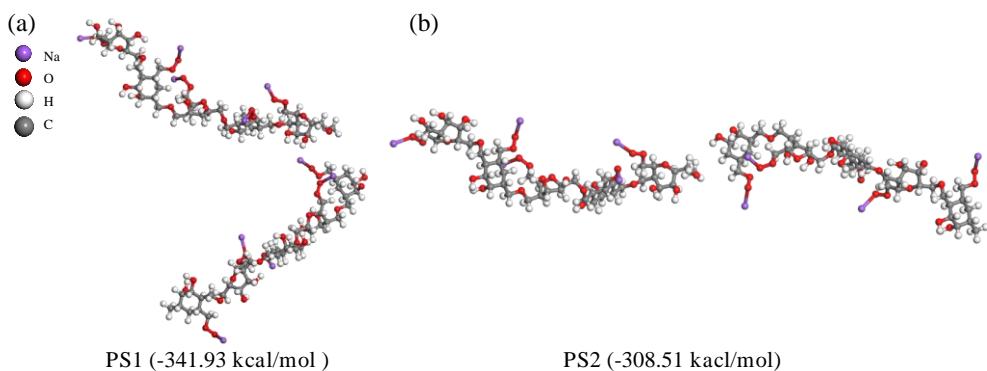
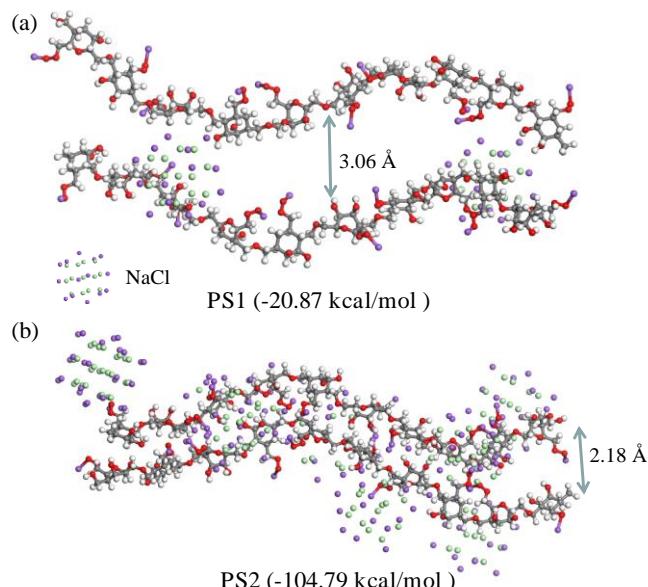


Fig. 10. Two optimized geometries for two single alginate chains that contained 10 uronic units but no NaCl.

Fig. 11 shows the states between two long alginate chains at low and high NaCl content.

DFT calculations revealed the relative low energy of two parallel long alginate chains, indicating that the parallel state was energetically stable. The reason was that when NaCl present, the intermolecular interactions of two hydrogen bonds formed by the hydroxyl and carboxyl groups of the aldaldehyde rings at terminals of alginate chains ensured the aldaldehyde rings tightly connected but parallel to each other [47]. At low NaCl content, the distance between the two hydrogen bonds of two alginate chains was 3.06 Å and the energy was -20.87 kcal/mol (Fig. 11(a)). However, the distance between the two alginate chains was 2.18 Å, with lower energy

347 (-104.79 kcal/mol) at high ionic strength (Fig. 11(b)). To summarize, the simulation results  
348 demonstrated that the NaCl trapped in gel layers could shorten the distance between hydrogen  
349 bonds and lower the energy of alginate chain terminals.



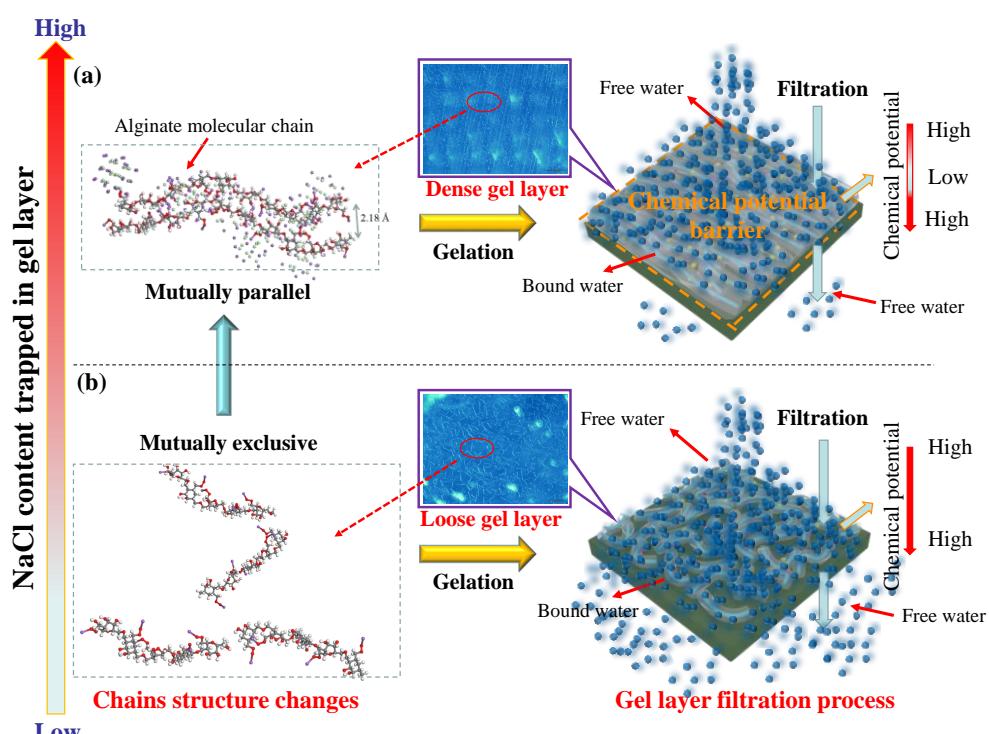
350  
351 Fig. 11. Two optimized geometries of long alginate chains containing 20 uronic units: (a)  
352 low NaCl content and (b) high NaCl content.

353 Figs. 10 and 11 provide structural and energy information of alginate molecular chains **with**  
354 **varied content of NaCl**. It is clear that alginate chains were mutually repulsive at extremely low  
355 NaCl content or in the absence of NaCl, while the presence of NaCl ensured that the terminal  
356 aldehyde-aldehyde rings were tightly connected and parallel to each other due to the hydrogen  
357 bonds existing between the carboxyl groups at the terminals of alginate chains [28]. As the NaCl  
358 content in gel layer increased, the distance and the energy between hydrogen bonds formed by  
359 two carboxyl groups decreased. Consequently, two long alginate chains stayed in closer  
360 proximity to each other and thus formed a regular and dense alginate layer. According to the  
361 formation mechanism of alginate chain structures with varied content of NaCl revealed by DFT  
362 simulation, the reverse diffusion of draw solutions or the ion content in gel layers had a critical

363 influence on the molecular chain structure and interaction of alginate gel layers (Fig. 9).  
 364 According to the formation mechanism of alginate chain structures with varied NaCl content  
 365 revealed by DFT simulation, the reverse diffusion of draw solutions or the ion content in gel  
 366 layers had a critical influence on the molecular chain structure and interaction of alginate gel  
 367 layer (Fig. 9).

368 **3.4. Mechanistic insights into FO membrane fouling behaviors**

369 The above-mentioned advanced techniques including phase-contrast microscope and DFT  
 370 simulation provided detailed structure information of alginate gel layers formed in draw  
 371 solutions with different ionic strength. Based on these results, Fig. 12 introduces a schematic  
 372 diagram of the effect mechanisms of reverse diffusion of draw solution on the fouling behavior  
 373 of alginate layer.



374  
 375 **Fig. 12.** Schematic diagram of the cross-linking process of alginate chains affected by reverse  
 376 NaCl and the filtration process of alginate layers governed by Flory-Huggins mechanism.

377 It can be seen from Fig.12(b) that the gel layers (shown in Figs. 3(b1) and (b2)) formed on  
 378 the FO membrane surface were very loose at low content of reverse diffused NaCl due to the  
 379 repulsion between carboxylic acid groups on the alginate chains. These loose gel structures  
 380 could not effectively bind with water, so making it impossible to substantially lower the water  
 381 flux (Figs. 2(a) and (b)) [25, 64]. However, a large amount of NaCl trapped in gel layers reduced  
 382 the repulsion between the alginate chains and shortened the distance between hydrogen bonds at  
 383 the two terminals of alginate chains, thereby providing a highly cross-linked polymeric network  
 384 (Fig. 12(a)) [28]. According to the Flory-Huggins lattice theory expressed by Eq. (5), the  
 385 chemical potential change accompany by the formation of gel layer is dependent on  
 386 polymerization degree, and highly cross-linked network corresponds to higher chemical  
 387 potential change. From thermodynamic viewpoint, filtration through a gel layer can be regarded  
 388 as the inverse process of gel formation. Therefore, filtration through a denser gel layer (Fig.  
 389 12(a)) is considered as dragging water from gel layer (much low chemical potential) to permeate  
 390 side (high chemical potential) [24, 65]. In other words, dense gel layer functioned as a  
 391 “chemical potential barrier”, which offset a part of chemical potential gap between draw  
 392 solution and feed solution, resulting in water penetration becomes difficult. During the FO  
 393 membrane filtration process of gel, the osmotic pressure gap ( $\Delta\pi_{\text{gel}}$ ) was used to overcome the  
 394 water chemical potential gap ( $\Delta\mu_{\text{gel}}$ ) between draw solution and gel layer [17]:

$$395 \Delta\mu_{\text{gel}} = -V_B \Delta\pi_{\text{gel}} \quad (6)$$

396 where  $V_B$  represents the molar volume of solvent and is generally considered to be equal to the  
 397 molar volume of the dilute solution ( $V$ ),  $\Delta\pi_{\text{gel}}$  is the part of osmotic pressure gap ( $\Delta\pi_{\text{drive}}$ ) that  
 398 is used to overcome the chemical potential gap between gel layer and combination of free water  
 399 and pure polymers ( $\Delta\mu_{\text{gel}}$ ). Substituting Eq. (5) into Eq. (6), According to Flory-Huggins theory,

400  $\Delta\pi_{\text{gel}}$  can be calculated by:

401 
$$\Delta\pi_{\text{gel}} = -\Delta\mu_{\text{gel}}/V_B = -RT \left[ \ln(1-\phi_2) + (1-N^{-1})\phi_2 + \chi\phi_2^2 \right]/V_B \quad (7)$$

402 For FO membrane filtration processes, the trapped NaCl in gel layer also contributes  
403 osmotic pressure ( $\pi_{\text{NaCl}}$ ), which can be calculated by Eq. 2. Therefore, during gel layer filtration,  
404 the net osmotic pressure gap ( $\Delta\pi_{\text{net}}$ ) can be described by:

405 
$$\Delta\pi_{\text{net}} = \Delta\pi_{\text{drive}} - \Delta\pi_{\text{gel}} - \pi_{\text{NaCl}} \quad (8)$$

406 The above Eqs were perfect for the description of the chemical potential since previous  
407 studies have calculated the chemical potential of agar gels and based on these Eqs [17, 19]. The  
408 calculation results suggested that the chemical potential gap between gel layer and permeate was  
409 huge, so the  $\Delta\pi_{\text{gel}}$  used to overcome this chemical potential gap may be extremely high [17,  
410 61].  $\Delta\pi_{\text{gel}}$  can be quantitatively assessed by Eq. (7) provided that **several thermodynamic**  
411 **parameters are determined**. Due to the  $\chi$  value of alginate has not been reported and is difficult  
412 to be estimated, it is hard to conduct quantitative simulation of variation of the Flory-Huggins  
413 induced chemical potential gap with the degree of polymerization [27]. **The degree of**  
414 **polymerization of the polymer chains increased with the rising content of NaCl**. Although it is  
415 **hard to conduct a quantitative simulation of the variation of the Flory-Huggins induced**  
416  **$\Delta\mu_{\text{gel}}$  with the degree of polymerization**. The general profile of relation between Flory-Huggins  
417 **induced the  $\Delta\mu_{\text{gel}}$  and polymerization degree could be depicted by Eq. (5)**. According to Eq. (5)  
418 **and Eq. (7)**,  $\Delta\mu_{\text{gel}}$  decrease with the degree of polymerization, which corresponded to  $\Delta\pi_{\text{gel}}$   
419 **increase, resulting in more flux decline**. The real existence of the  $\Delta\pi_{\text{gel}}$  induced by  
420 Flory-Huggins mechanism was confirmed by quantifying the changes in osmotic pressure  
421 during FO membrane filtration.

422 In the filtration experiment of alginate solutions in 4 mol/L NaCl draw solution ( $\Delta\pi_{\text{drive}}$  is

423 24662.16 kPa). After filtration of 500 mL foulants, the  $\Delta\pi_{\text{drive}}$  was diluted to 22705.49 kPa, as  
424 can be calculated by converting the conductivity gap between draw solution and feed solution,  
425 and its driving force decreased by 7.9% compared with that in the initial stage. However, water  
426 flux decreased by about 69.2%, suggesting that the  $\Delta\pi_{\text{net}}$  at this time was only 7595.95 kPa  
427 (30.8% $\times$ 24662.16 kPa). This meant that the osmotic pressure gap offset by gel layer should be  
428 15109.54 kPa (22705.49 kPa-7595.95 kPa). It is important to note that the osmotic pressure of  
429 the gel layer includes the  $\pi_{\text{NaCl}}$  caused by reverse diffused NaCl and the  $\Delta\pi_{\text{gel}}$ . The  
430 concentration of NaCl trapped in the gel layer was then measured to be 0.60 mol/L by the  
431 inductively coupled plasma, and the  $\pi_{\text{NaCl}}$  was 2780.67 kPa. Thus, the  $\Delta\pi_{\text{gel}}$  to overcome the  
432 chemical potential gap of this gel layer was calculated to be 12328.87 kPa (15109.54  
433 kPa-2780.67 kPa) by Eq. (8), accounting for 54.30% of the  $\Delta\pi_{\text{drive}}$  of 4 mol/L NaCl draw  
434 solution. Seen from the calculation results, the osmotic pressure gap required to overcome the  
435 chemical potential gap between gel layer and draw solution was extremely high, which offsets a  
436 lot of driving force. Filtration experiments show that the  $\Delta\pi_{\text{gel}}$  increased with the degree of  
437 polymerization of the polymer chains rising, resulting in decrease of the net osmotic pressure  
438 gap, corresponding to flux decrease. The increase trend of the  $\Delta\pi_{\text{gel}}$  depicted by Eq. (7) was  
439 consistent with filtration experiment results. Therefore, the Flory-Huggins theory not only  
440 provided a description of mechanism of chemical potential gap during the FO filtration process,  
441 but also explained the fouling tendency of alginate solution with reverse diffusion of solution.

#### 442 4. Conclusions

443 Filtration tests showed that the reverse diffusion of draw solutions had critical effects on  
444 FOMBR filtration behavior of alginate solutions. Phase-contrast microscope images and DFT  
445 calculation results proved at the molecular level that the content of ions trapped in gel layers

446 directly determined the molecular chain structure of gel layers. Moreover, heating experiments  
447 confirmed that gel layers with high NaCl content required more energy to drag out bound water,  
448 indicating that gel layer acted as a "chemical potential barrier" that prevented bound water from  
449 turning into free water. The above results established for the first time that the gel layer (the  
450 "chemical potential barrier") was primarily affected by changes of the layer's molecular chain  
451 structure. Subsequently, the "chemical potential barrier" described by Flory-Huggins theory was  
452 proposed to elucidate the FOMBR fouling behavior affected by reverse diffusion of draw  
453 solutions **from the perspective of chemical potential changes. This hand-in-hand method of**  
454 **combining experimental characterizations and theoretical calculations provided a**  
455 **comprehensive understanding of the effect of reverse diffusion of draw solution on alginate**  
456 **fouling properties.**

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**Conflict of Interest**

The authors declared that they have no conflicts of interest to this work.

We declare that we do not have any commercial or associative interest that represents a conflict of interest in connection with the work submitted.

## Author Statement

**Jiaheng Teng:** Investigation, Methodology, Writing - original draft.

**Hanmin Zhang:** Conceptualization, Funding acquisition, Project administration, Writing - review & editing.

**Chuyang Tang:** Investigation, Data curation, Formal analysis.

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