

Performance of serial time-encoded amplified microscope

Kevin K. Tsia,^{1,2,*} Keisuke Goda,¹ Dale Capewell,¹ and Bahram Jalali¹

¹UCLA Photonics Laboratory, University of California, Los Angeles, CA, 90095 USA

²Current address, Department of Electrical and Electronic Engineering, University of Hong Kong, Pokfulam Road, Hong Kong

*tsia@hku.hk

Abstract: Serial time-encoded amplified microscopy (STEAM) is an entirely new imaging modality that enables ultrafast continuous real-time imaging with high sensitivity. By means of optical image amplification, STEAM overcomes the fundamental tradeoff between sensitivity and speed that affects virtually all optical imaging systems. Unlike the conventional microscope systems, the performance of STEAM depends not only on the lenses, but also on the properties of other components that are unique to STEAM, namely the spatial disperser, the group velocity dispersion element, and the back-end electronic digitizer. In this paper, we present an analysis that shows how these considerations affect the spatial resolution, and how they create a trade-off between the number of pixels and the frame rate of the STEAM imager. We also quantify how STEAM's optical image amplification feature improves the imaging sensitivity. These analyses not only provide valuable insight into the operation of STEAM technology but also serve as a blue print for implementation and optimization of this new imaging technology.

©2010 Optical Society of America

OCIS codes: (180.0180) Microscopy; (170.0180) Imaging systems

References and links

1. J. B. Pawley, ed., *Handbook of biological confocal microscopy*, 3rd ed., Springer (2006).
2. S. W. Hell, "Microscopy and its focal switch," *Nat. Methods* **6**(1), 24–32 (2009).
3. E. Betzig, G. H. Patterson, R. Sougrat, O. W. Lindwasser, S. Olenych, J. S. Bonifacino, M. W. Davidson, J. Lippincott-Schwartz, and H. F. Hess, "Imaging intracellular fluorescent proteins at nanometer resolution," *Science* **313**(5793), 1642–1645 (2006).
4. B. Huang, W. Wang, M. Bates, and X. Zhuang, "Three-dimensional super-resolution imaging by stochastic optical reconstruction microscopy," *Science* **319**(5864), 810–813 (2008).
5. H. R. Petty, "Spatiotemporal chemical dynamics in living cells: from information trafficking to cell physiology," *Biosystems* **83**(2–3), 217–224 (2004).
6. H. R. Petty, "High speed microscopy," *Opt. Photonics News*, 34–40 (2004).
7. J. V. Watson, *Introduction to flow cytometry*, (Cambridge Univ. Press, Oxford, U. K. 2004).
8. J. R. Janesick, *Scientific Charge-Coupled Devices*, (SPIE Publications, 2001).
9. G. C. Holst, and T. S. Lomheim, *CMOS/CCD Sensors and Camera Systems*, SPIE-International Society for Optical Engine (2007).
10. K. Goda, K. K. Tsia, and B. Jalali, "Amplified dispersive Fourier-transform imaging for ultrafast displacement sensing and barcode reading," *Appl. Phys. Lett.* **93**(13), 131109 (2008).
11. K. Goda, K. K. Tsia, and B. Jalali, "Serial time-encoded amplified imaging for real-time observation of fast dynamic phenomena," *Nature* **458**(7242), 1145–1149 (2009).
12. J. Chou, O. Boyraz, D. Solli, and B. Jalali, "Femtosecond real-time single-shot digitizer," *Appl. Phys. Lett.* **91**(16), 161105 (2007).
13. D. R. Solli, J. Chou, and B. Jalali, "Amplified wavelength-time transformation for real-time spectroscopy," *Nat. Photonics* **2**(1), 48–51 (2008).
14. J. Chou, D. R. Solli, and B. Jalali, "Real-time spectroscopy with subgigahertz resolution using amplified dispersive Fourier transformation," *Appl. Phys. Lett.* **92**(11), 111102 (2008).
15. K. Goda, D. R. Solli, K. K. Tsia, and B. Jalali, "Theory of amplified dispersive Fourier transformation," *Phys. Rev. A* **80**(4), 043821 (2009).

16. G. J. Tearney, M. Shishkov, and B. E. Bouma, "Spectrally encoded miniature endoscopy," *Opt. Lett.* **27**(6), 412–414 (2002).
17. C. Pitris, B. Bouma, M. Shishkov, and G. Tearney, "A GRISM-based probe for spectrally encoded confocal microscopy," *Opt. Express* **11**(2), 120–124 (2003).
18. Z. Yaqoob, and N. A. Riza, "Eye-safe passive-optics no-moving parts barcode scanners," *IEEE Photon. Technol. Lett.* **16**(3), 954–956 (2004).
19. S. Xiao, A. M. Weiner, and C. Lin, "A dispersion law for virtually-imaged phased-array spectral dispersers based on paraxial-wave theory," *IEEE J. Quantum Electron.* **40**(4), 420–426 (2004).
20. S. A. Diddams, L. Hollberg, and V. Mbele, "Molecular fingerprinting with the resolved modes of a femtosecond laser frequency comb," *Nature* **445**(7128), 627–630 (2007).
21. S. Xiao, and A. M. Weiner, "2-D wavelength demultiplexer with potential for ≥ 1000 channels in the C-band," *Opt. Express* **12**(13), 2895–2902 (2004).
22. K. K. Tsia, K. Goda, D. Capewell, and B. Jalali, "Simultaneous mechanical-scan-free confocal microscopy and laser microsurgery," *Opt. Lett.* **34**(14), 2099–2101 (2009).
23. Y. Han, and B. Jalali, "Continuous-time time-stretched analog-to-digital converter array implemented using virtual time gating," *IEEE Trans. Circ. Syst.* **52**(8), 1502–1507 (2005).
24. G. P. Agrawal, *Fiber Optic Communication Systems*, 3rd Ed., Wiley-Interscience (2002).
25. V. E. Perlin, and H. G. Winful, "On Distributed Raman Amplification for Ultrabroad-Band Long-Haul WDM Systems," *J. Lightwave Technol.* **20**(3), 409–416 (2002).
26. M. T. Tilli, M. C. Cabrera, A. R. Parrish, K. M. Torre, M. K. Sidawy, A. L. Gallagher, E. Makariou, S. A. Polin, M. C. Liu, and P. A. Furth, "Real-time imaging and characterization of human breast tissue by reflectance confocal microscopy," *J. Biomed. Opt.* **12**(5), 051901 (2007).
27. A. Vogel, J. Noack, G. Hüttman, and G. Paltauf, "Mechanisms of femtosecond laser nanosurgery of cells and tissues," *Appl. Phys. B* **81**(8), 1015–1047 (2005).
28. J. V. Watson, *Introduction to flow cytometry*, Cambridge Univ. Press, Oxford, U. K. (2004).
29. M. Fleisher, "Circulating tumor cells – a new opportunity for therapeutic management of cancer patients," *Clin. Lab. News* **34**, 10 (2008).

1. Introduction

As a bread-and-butter diagnostic tool in biomedicine, optical microscopy has fueled spectacular progress in unraveling the complexity of the physiological processes in biological tissues and cells [1–4]. In particular, high-speed optical microscopy has been of great importance to study dynamical processes, especially non-repetitive transient phenomena. Examples are (i) the spatiotemporal study of biochemical waves in cells/tissues, which requires imaging with a μs - to ns-response time [5,6], and (ii) flow cytometry where high-speed imagers are required to provide high-throughput cell characterization [7].

However, it has been challenging for conventional optical microscopy to capture fast dynamical processes with high sensitivity and temporal resolution. This is mainly due to the fundamental trade-off between sensitivity and frame rate that appears in standard charge-coupled devices (CCDs) and their complementary metal oxide semiconductor (CMOS) counterparts – the workhorse in microscopy [1, 8,9]. Other drawbacks of fast CCD/CMOS imagers are the requirement of cooling to reduce thermal noise, which adds complexity and cost of refrigeration; and the need for high-intensity illumination to ensure adequate signal-to-noise (SNR) ratio, which causes damage to, or modification of, the object being imaged.

We recently developed an entirely new imaging modality called *serial time-encoded amplified microscopy* (STEAM) [10, 11]. It overcomes the limitations of CCD/CMOS imagers and offers a few orders of magnitude higher frame rates (more than 5 MHz) than these imagers. It encodes the image onto the spectrum of a laser pulse and then converts the spectrum into an *optically-amplified* time-domain serial data. This allows us to capture images with a single-pixel detector, eliminating the need for the CCD/CMOS imagers and their associated trade-off between imaging sensitivity and speed by optical image amplification. Unlike the conventional microscopes in which the lens systems and CCD/CMOS imagers in general dictate the imaging quality, the performance of the STEAM system depends not only on the lens system, but also on the characteristics of the mapping processes which convert the spatial information to the serial temporal data. As a result, evaluating the final STEAM image quality requires careful considerations of the properties of the individual elements involved in these processes. In this paper, we present an analysis of STEAM performance to show how these considerations uniquely establish the limit to spatial

resolution, and create the trade-off between the number of pixels and the frame rate. More importantly, we will also describe the impact of optical image amplification on image sensitivity.

2. Working principle of STEAM

The key feature of STEAM is the mapping of an image into a serial time-domain waveform by a two-step approach (Fig. 1): (i) space-frequency mapping – the spatial information of an object is first encoded onto the spectrum of a broadband pulse by using a spatial disperser [10, 11]. (ii) Frequency-time mapping – A dispersive element (e.g., a dispersive fiber) is then used to perform a process called *amplified dispersive Fourier transform* (ADFT) which maps the spectrum of an optical pulse into a temporal waveform using group-velocity dispersion (GVD) [12–15]. The optical spectrum, which is encoded with the image, now appears as a serial sequence in time. To simultaneously amplify the image, the dispersive fiber is pumped with secondary light sources to implement optical amplification directly within fiber. This powerful approach compensates for the inherent loss associated with GVD [10–15] and brings the signal above the thermal noise of the photodetector – enabling high speed imaging at low light levels. Due to the optical image serialization, the image can be detected with a single-pixel photodiode and captured, not by a CCD/CMOS camera, but instead, with any real-time digitizer.

STEAM can either perform one-dimensional (1-D) or two-dimensional (2-D) imaging, called *1-D STEAM* or *2-D STEAM*, respectively. 1-D STEAM utilizes a 1-D spatial disperser such as a prism, or a diffraction grating to generate a 1-D spectral pattern in space for illumination, resembling a 1-D spectral shower [10, 16–18]. The 1-D spatial information is then encoded onto the back-reflected 1-D spectral shower which is subsequently converted into a serial temporal waveform by ADFT. In contrast, *2-D STEAM* uses a 2-D spatial disperser which comprises of two orthogonally oriented 1-D spatial dispersers to generate a 2-D spectral shower onto which the 2-D spatial information is encoded. The subsequent frequency-time mapping process is identical to 1-D STEAM. The analysis of the STEAM performance, in the context of both 1-D and 2-D STEAM, is of main focus of this paper.

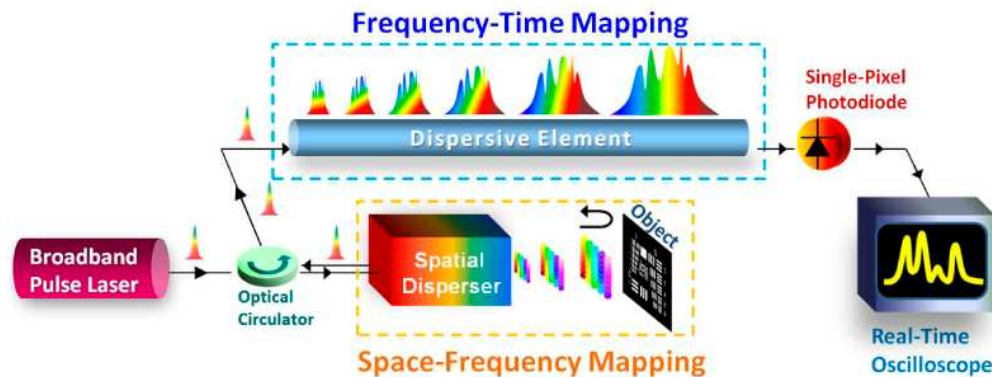


Fig. 1. Generic schematic of a STEAM system.

3. Spatial resolution of STEAM

The actual spatial resolution of STEAM is not solely determined by diffraction limit, as in the case of typical confocal microscopy [1]. It can also be affected during the space-frequency and frequency-time mapping processes. Specifically, it can be governed by (i) the spectral resolution of the spatial disperser (*spatial-dispersion-limited regime*), (ii) the spectral resolution imposed by ADFT through stationary-phase-approximation (SPA) (*SPA-limited regime*) and (iii) the temporal resolution of the digitizer (*digitizer-limited regime*). These limiting factors will be discussed in detail in the context of 1-D and 2-D STEAM below.

3.1 1-D STEAM

In 1-D STEAM, the space-frequency mapping is accomplished by using a 1-D spatial disperser to first generate a 1-D spectral shower [10, 16–18]. Consider the use of a diffraction grating as the 1-D spatial disperser in 1-D STEAM as shown in Fig. 2, its spectral resolution is well-known to be (assuming first-order diffraction)

$$\delta\lambda_g = \frac{\lambda \cdot d \cdot \cos\theta_g}{W}, \quad (1)$$

where θ_g is the diffracted angle under Littrow's condition [16], λ is the center wavelength, d is the grating period, and W is the input beam waist. When the spatial resolution is governed by the spatial dispersion of the grating, it is said to be *spatial-dispersion limited* and is given by,

$$\delta x_{1D}^{spatial} \approx \left(f \cdot \frac{d\theta_g}{d\lambda} \right) \cdot \delta\lambda_g = C_x \cdot \delta\lambda_g, \quad (2)$$

where f is the focal length of the objective lens, C_x represents the conversion factor between the space and wavelength, and $d\theta_g/d\lambda = 1/d \cos(\theta_g)$ is the angular dispersion of the diffraction grating. In practice, the 1-D spectral shower beam underfills the aperture of the objective lens in order for the objective lens to capture the whole spectrum. It implies that the spatial-dispersion-limited spatial resolution would be worse than the diffraction-limited resolution.

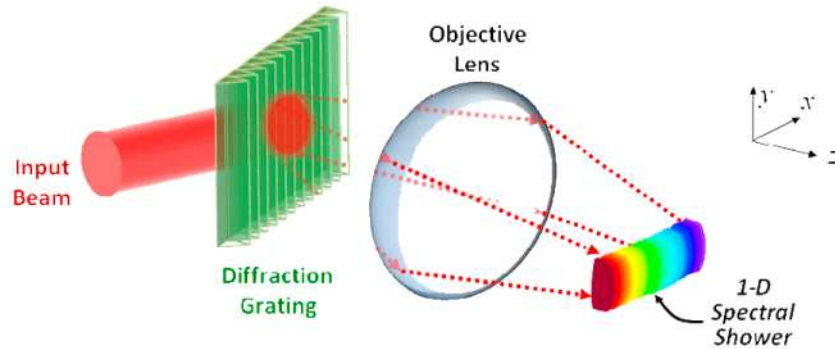


Fig. 2. Schematic of a 1-D spatial disperser used in a 1-D STEAM system.

On the other hand, the ADFT process should ideally perform one-to-one frequency-time mapping, i.e. only one wavelength contributes to the temporal waveform at any one time instant. However, there is always an ambiguity in this mapping process – a fundamental property of the dispersive Fourier transform [15]. Such an ambiguity, defined by SPA, can be used as a measure of the spectral resolution of the ADFT process ($\delta\lambda_{SPA}$) [15]:

$$\delta\lambda_{SPA} = \lambda \sqrt{\frac{2}{D \cdot c}}, \quad (3)$$

where D is the GVD and c is the speed of light. Note that in Eq. (3), we neglect the higher-order dispersions which result in nonlinear frequency-time mapping. Nevertheless, it is adequate here to illustrate the primary contribution of ADFT to the spatial resolution of STEAM. More detailed analysis including higher-order dispersion can be referred to ref [15].

Similar to Eq. (2), the spectral resolution $\delta\lambda_{SPA}$ can be translated to the *SPA-limited* spatial resolution because of the space-frequency mapping. It can be written as

$$\delta x_{1D}^{SPA} = C_x \cdot \delta\lambda_{SPA}. \quad (4)$$

Moreover, the temporal resolution (finite bandwidth) of the optical detection system in STEAM (i.e., photodetector, and electronic digitizer) can also be a limiting factor of the spatial resolution of STEAM. It can be understood by recognizing the temporal resolution of the digitizer imposes an equivalent spectral resolution via ADFT, which is given by [15],

$$\delta\lambda_{\text{det}} = \frac{0.35}{D \cdot f_{\text{det}}}, \quad (5)$$

where f_{det} is the bandwidth of the detection system (i.e., the photodetector and digitizer). Hence, the *digitizer-limited* spatial resolution can be expressed as

$$\delta x_{1D}^{\text{det}} = C_x \cdot \delta\lambda_{\text{det}}. \quad (6)$$

The contribution of each of the above three limiting factors [i.e. Equations (2), (4) and (6)] to the actual spatial resolution of 1-D STEAM (δx_{1D}) is exemplified in Fig. 3(a). The values of the parameters used in this example are detailed in the figure caption. We observe that the spatial resolution is digitizer-limited (i.e. $\delta x_{1D} \approx \delta x_{1D}^{\text{det}}$) if the GVD is small ($< \sim 0.1$ ns/nm). Increasing GVD can bring the system into *SPA-limited* (i.e. $\delta x_{1D} \approx \delta x_{1D}^{\text{SPA}}$). Further increase in GVD (> 1 ns/nm) results in a *spatial-dispersion-limited* operation (i.e. $\delta x_{1D} \approx \delta x_{1D}^{\text{spatial}}$) in which δx_{1D} is independent of GVD. The corresponding temporal resolution, which is given by $\delta t_{1D} = D \cdot \delta x_{1D} / C_x$, is shown in Fig. 3(b). It is ideal to have the STEAM system whose temporal resolution is limited by the available GVD, not the bandwidth of the digitizer, while keeping the best achievable spatial resolution [$\delta x_{1D} \sim 0.5 \mu\text{m}$ in Fig. 3(a)]. In order to meet this criterion, the favorable operating regime should be $\sim 0.1 - 1$ ns/nm in this example [see Fig. 3(b)].

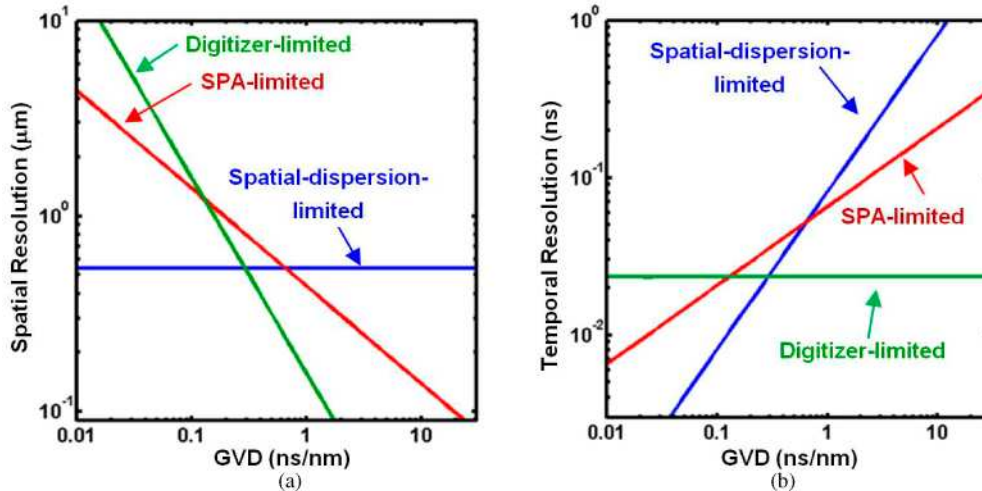


Fig. 3. (a) Spatial resolution and (b) temporal resolution of 1-D STEAM in various limiting cases: digitizer-limited (green), SPA-limited (red) and spatial-dispersion-limited (blue), as a function of GVD. The system parameters are: $\lambda = 800$ nm, $W = 3$ mm, an objective lens with $\text{NA} = 0.9$ ($f = 2$ mm), $1/d = 1800$ lines/mm, and $f_{\text{det}} = 15\text{GHz}$.

3.2 2-D STEAM

Here we require a 2-D spatial disperser to map the spectrum into a 2-D space, generating a 2-D spectral shower on which the spatial information is encoded. The 2-D spatial disperser consists of two spatial dispersers: a diffraction grating and a virtually-imaged phased array (VIPA) [19]. Their dispersion directions are orthogonal to each other. The key feature of the VIPA is that wavelengths differed by integer multiples of its free spectral range (FSR) are

spatially overlapped with each other in one dimension. A diffraction grating is used to remove this spatial degeneracy in the orthogonal dimension, resulting in a 2-D spectral shower (Fig. 4). Such a grating-VIPA arrangement has previously been used for spectroscopy [20] and for wavelength de-multiplexing in telecommunication applications [21]. In contrast, here we employ this 2D spatial disperser for the purpose of imaging [11, 22].

Following the 1-D analysis, we note that the spatial resolution of 2-D STEAM is also governed by the three aforementioned limiting regimes (i.e., the *spatial-dispersion-limited*, *SPA-limited* and *digitizer-limited* regimes.) Moreover, the spatial-dispersion-limited spatial resolutions in the two orthogonal dimensions are different because of the different dispersive properties of the VIPA and the diffraction grating. To estimate these spatial resolutions, we need to recognize different effects which limit the spatial resolution in the x - and y -directions.

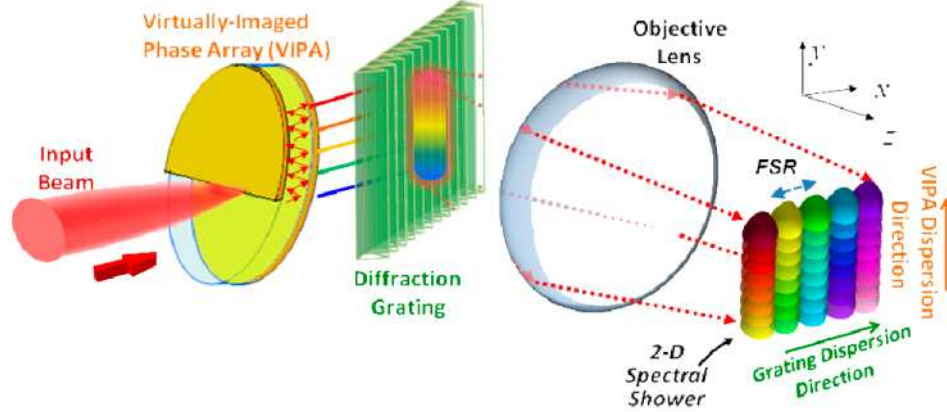


Fig. 4. Schematic of a 2-D spatial disperser (a virtually-imaged phase array (VIPA) and a diffraction grating) which generates the 2-D spectral shower.

3.2.1 Spatial-dispersion-limited spatial resolution in the y -direction

The spectral resolution of the VIPA, namely its finite spectral linewidth, is the key factor determining the spatial-dispersion-limited spatial resolution in the y -direction in 2-D STEAM. This linewidth ($\delta\lambda_{VIPA}$) is defined by the full-width half-maximum (FWHM) of the transmission resonance spectrum of the VIPA, which is given by Ref [19].

$$I_t(k) \propto \frac{[1 - (R_1 R_2)^N]^2 + 4(R_1 R_2)^N \sin^2 [Nkt \cos(\theta_{in})]}{(1 - R_1 R_2)^2 + 4R_1 R_2 \sin^2 [kt \cos(\theta_{in})]}, \quad (7)$$

where R_1 and R_2 are the reflectivities of the VIPA's front and back mirrors, respectively. θ_{in} is the angle related to the VIPA's tilt angle θ_{VIPA} by $\sin(\theta_{VIPA}) = n \cdot \sin(\theta_{in})$, where n is the refractive index of the VIPA. t is the thickness of the VIPA. k is the wavenumber, $k = 2\pi n/\lambda$.

In Eq. (7), N is the number of VIPA's "virtual sources" contributed to the actual spectral shower and it is an important parameter determining the spectral resolution of the VIPA. It can be approximated as $N \approx L/2t \cdot \tan(\theta_{in})$, where L is the aperture size of the VIPA. However, if the output beam from the VIPA overfills the back aperture of the objective lens (with a size of D), the objective lens is essentially unable captures all the "virtual sources". In this case, the N is rather limited by the aperture size of the objective lens, i.e. $N \approx D/2t \cdot \tan(\theta_{in})$. For example, with $L = 1$ cm, $\theta_{in} = 3^\circ$, $t = 1.5$ mm, and $D = 5$ mm, we can only obtain $N \approx 25$. The consequence of such finite N is linewidth broadening [see Eq. (7)], which in turns limits the spatial resolution in the y -direction ($\delta y_{2D}^{spatial}$) through space-frequency mapping. This finite- N effect is analogous to the diffraction grating in which the number of illuminated groove lines defines the spectral resolution. Similar to Eq. (2), $\delta y_{2D}^{spatial}$ is given by,

$$\delta y_{2D}^{spatial} \approx f \cdot \frac{d\theta_{VIPA}}{d\lambda} \cdot \delta\lambda_{VIPA} = C_y \cdot \delta\lambda_{VIPA}, \quad (8)$$

where C_y is the conversion factor between space in the y -direction and the wavelength, and $d\theta_{VIPA}/d\lambda \approx -2[n^2 - \sin^2\theta_{VIPA}]/[\lambda\sin(2\theta_{VIPA})]$ is the angular dispersion of the VIPA [19]. Here, we ignore the higher-order dispersion effect of the VIPA which is only significant when large diffracted angle from the VIPA is considered [19].

The finite- N effect on $\delta y_{2D}^{spatial}$ is experimentally verified as shown in Fig. 5. Figure 5(a) shows the spectral shower generated using a broadband pulsed beam. Another continuous-wave (CW) laser beam is also coupled together and appeared as a single spot on top of the spectral shower. $\delta y_{2D}^{spatial}$ can then be extracted from the lineshape measured across the spot in the y -direction, that is $\delta y_{2D}^{spatial} \sim 3.2 \mu\text{m}$ [Fig. 5(b)–5(c)]. It is noted that the lineshape predicted by Eq. (7) [blue line in Fig. 5(c)] fits reasonably well with the measured data. The result is also verified by employing an optical system design simulation tool (ZEMAX), which gives $\delta y_{2D}^{spatial} \sim 2.8 \mu\text{m}$ [22]. We note that the linewidth broadening effect due to the finite N becomes clear when comparing with the FWHM of the infinite- N case.

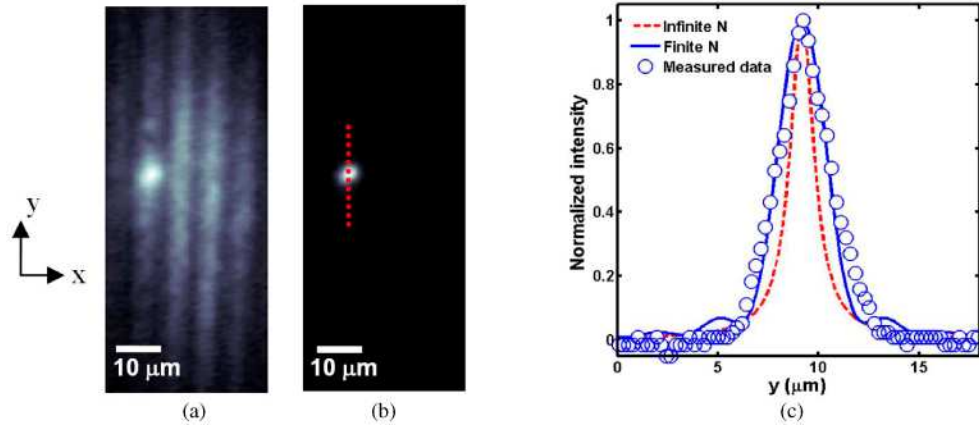


Fig. 5. (a) A 2-D spectral shower, which is generated using a broadband source (a center wavelength at 1570 nm), overlaid with the CW spot (a wavelength at 1565 nm). (b) The same CW spot but with the spectral shower turned off. (c) (blue circles) The measured lineshape of CW spot (along the red dotted line in (b)). The lineshape predicted by the cases of finite N (blue solid line, $N \sim 25$ in this case) and infinite N (red dashed line) are also shown for comparison. In this example, we employ a glass VIPA ($n = 1.48$) with $t = 1.5$ mm and $\theta_{VIPA} = 3^\circ$; a diffraction grating with a groove density ($1/d$) is 1200 lines/mm and $\theta_g = 75^\circ$; an objective lens with $f = 5$ mm, $D = 5$ mm.

3.2.2 Spatial-dispersion-limited spatial resolution in the x -direction

From Fig. 5(a), it is conceivable that the spatial-dispersion-limited spatial resolution in the x -direction ($\delta x_{2D}^{spatial}$) can be defined as one column separation of the spectral shower (δx_{2D}^{FSR}), which corresponds to one FSR of the VIPA in wavelength ($\Delta\lambda_{FSR}$). This relation can be written as

$$\delta x_{2D}^{spatial} = \delta x_{2D}^{FSR} = C_x \cdot \Delta\lambda_{FSR}, \quad (9)$$

where $\Delta\lambda_{FSR} \sim \lambda^2/2nt\cos(\theta_m)$. Consider the example shown in Fig. 5(a), then we can estimate $\delta x_{2D}^{spatial} \sim 8 \mu\text{m}$ using Eq. (9). It matches reasonably well the column separation shown in Fig. 5(a). It is also in good agreement with the recent demonstration of simultaneous mechanical-scan-free microscopy and laser surgery using similar 2-D spectral shower [22].

However, such definition is only valid when δx_{2D}^{FSR} is smaller than the spatial resolution imposed by both the spatial dispersion of the diffraction grating ($\delta x_{2D}^{grating}$) and the finite VIPA linewidth (δx_{2D}^{VIPA}). They are respectively given by $\delta x_{2D}^{grating} = C_x \cdot \delta \lambda_g$ and $\delta x_{2D}^{VIPA} = C_x \cdot \delta \lambda_{VIPA}$. In other words, these two conditions can be expressed as

$$\Delta \lambda_{FSR} \geq \delta \lambda_{VIPA}, \quad (10)$$

$$\Delta \lambda_{FSR} \geq \delta \lambda_g. \quad (11)$$

Clearly, the condition in Eq. (10) can readily be satisfied for a wide range of reasonable parameter values. The condition in Eq. (11) is, however, not straightforward and requires careful design of the diffraction grating, VIPA, and input beam profile. This defines a minimum input beam size for the VIPA-grating configuration. From Eq. (11), we can obtain

$$W \geq W_{min} = \frac{2n \cdot t \cdot d \cdot \cos(\theta_{in}) \cdot \cos(\theta_i^{grat})}{\lambda}. \quad (12)$$

Figure 6(a) and 6(b) show the required W_{min} in order to satisfy Eq. (12) and the corresponding $\delta x_{2D}^{spatial}$ as a function of both the grating density ($1/d$) and the VIPA thickness (t), respectively. Note that there is a region in which the spatial resolution is governed by the diffraction limit, rather than spatial-dispersion [shaded regions in Fig. 6 (a) and 6(b)]. These two figures provide a general guideline for optimizing $\delta x_{2D}^{spatial}$ in 2-D STEAM. As an example, if we employ a diffraction grating with $1/d = 250$ lines/mm and a VIPA with $t = 0.25$ mm and $\theta_{VIPA} = 7^\circ$, the minimum input beam size should be ~ 3.5 mm in order to achieve $\delta x_{2D}^{spatial} = 0.7 \mu\text{m}$ at $\lambda = 800$.

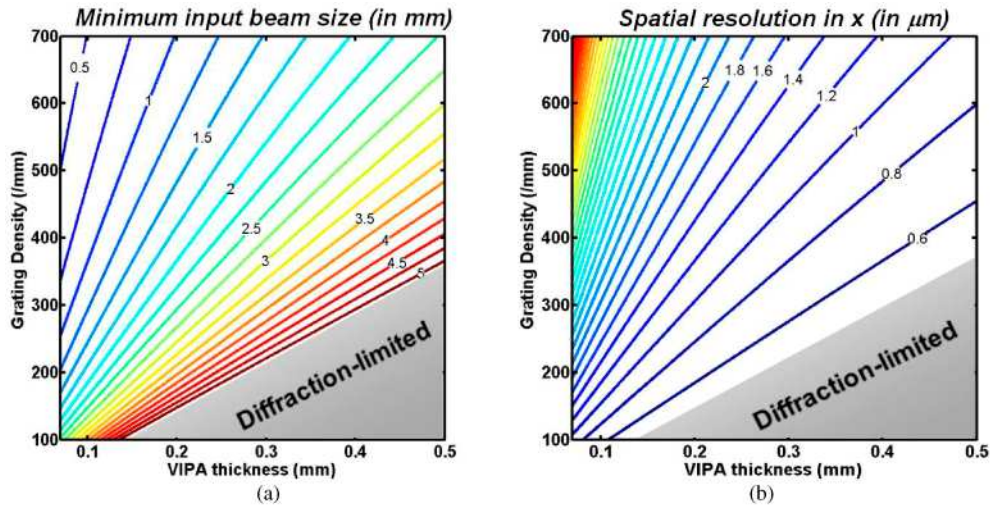


Fig. 6. A contour plot of (a) minimum input beam size in order to satisfy Eq. (12), and (b) the corresponding spatial-dispersion-limited resolution in the x -direction. The contour label unit is in mm in (a) and μm in (b). The shaded region represents the regime of diffraction-limited spatial resolution by the objective lens ($NA = 0.9, f = 2\text{mm}$). The center wavelength is 800 nm. The grating is assumed to satisfy Littrow's condition and the tilt angle of the glass VIPA ($n = 1.48$) is $\theta_{VIPA} = 7^\circ$.

3.2.3 SPA-limited and digitizer-limited spatial resolution

As a result of the serialization of the 2-D image by ADFT, only one dimension's spatial resolution, i.e. the y -direction, is subject to the effects of the SPA-limited and the digitizer-limited operation regimes. Similar to Eqs. (3) and (5), we can respectively write the SPA-limited (δy_{2D}^{SPA}) and digitizer-limited spatial resolution (δy_{2D}^{det}) as,

$$\delta y_{2D}^{SPA} = C_y \cdot \delta \lambda_{SPA} \quad (13)$$

$$\delta y_{2D}^{det} = C_y \cdot \delta \lambda_{det}. \quad (14)$$

In contrast, the actual spatial resolution in the x -direction (δx_{2D}) does not depend on the properties of ADFT and the digitizer, and hence is only affected by the spatial-dispersion properties by the 2-D spatial disperser. Therefore,

$$\delta x_{2D} = \delta x_{2D}^{spatial}. \quad (15)$$

The dependence of actual spatial resolution in the y -direction (δy_{2D}) on GVD follows the similar trend shown in the 1-D case [compare Fig. 7(a) and 2(a)]. The system transits from digitizer-limited, SPA-limited to spatial-dispersion limited operation with increasing GVD. Same trend can also be observed in temporal resolution, which is given by $\delta t_{2D} = D \cdot \delta y_{2D} / C_y$ [Fig. 7(b)]. In this example, the 2-D STEAM system is preferable to operate at $\sim 5 - 6$ ns/nm in order to make the temporal resolution of the system to be limited only by the available GVD while keeping the best achievable spatial resolution, i.e. $\delta y_{2D} \sim 0.7 \mu\text{m}$. It should be reminded such a large GVD can readily be made possible by the mean of optical amplification in STEAM, which overcomes the inherent loss associated with GVD. An extraordinarily large dispersion of > 10 ns/nm has been demonstrated in ADFT-based spectroscopy with ultra-high spectral resolution [13].

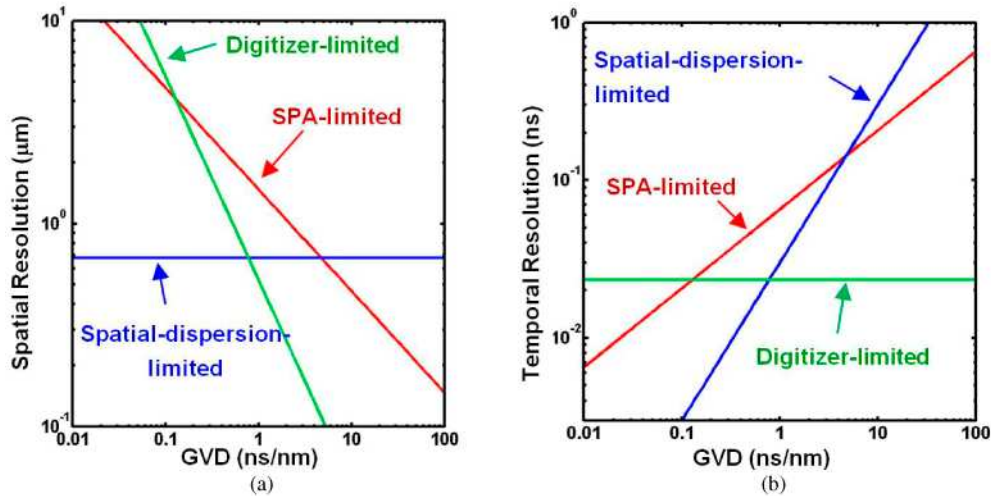


Fig. 7. (a) Spatial resolution of 2-D STEAM in the y -direction and (b) the corresponding temporal resolution in various limiting cases: digitizer-limited (green), SPA-limited (red) and spatial-dispersion-limited (blue), as a function of total GVD. The relevant system parameters are: a center wavelength of 800 nm, an objective lens with NA = 0.9 ($f = 2$ mm), a glass VIPA ($n = 1.48$) with tilt angle of 7° , thickness of 0.25 mm, $f_{det} = 15$ GHz.

4. Field-of-view, number of pixels and imaging frame rate

The field-of-view (FOV) of 1-D STEAM scales with the bandwidth of the spectral shower ($\Delta \lambda_{SS}$). Based on Eq. (2), the 1-D FOV is given by $\Delta x_{1D} = C_x \cdot \Delta \lambda_{SS}$. On the other hand, the

FOV of 2-D STEAM is set by spectral shower bandwidth and the FSR of the VIPA in the x - and y -direction, respectively. It can thus be estimated as $\Delta x_{2D} = C_x \cdot \Delta \lambda_{SS}$, and $\Delta y_{2D} = C_y \cdot \Delta \lambda_{FSR}$. Note that with the other parameters intact, the aspect ratio of the FOV can be tuned by varying $\Delta \lambda_{SS}$ and $\Delta \lambda_{FSR}$. Using the parameters shown in Fig. 5, we find $\Delta y_{2D} \sim 80 \mu\text{m}$ which agrees well with the measurement [Fig. 5(a)], whereas $\Delta x_{2D} \sim 200 \mu\text{m}$ with $\Delta \lambda_{SS} = 15 \text{ nm}$.

Another important parameter is the total number of pixels. It is equivalent to the number of data points sampled within each image-encoded temporal pulse after ADFT (i.e. single image frame). In 1-D STEAM, the temporal width of each pulse is set by spectral shower bandwidth ($\Delta \lambda_{SS}$) through ADFT, which is given by $\Delta \lambda_{SS} \cdot D$, the total number of pixels in 1-D STEAM (N_{1D}) can be written as

$$N_{1D} = \Delta \lambda_{SS} \cdot D \cdot f_{dig}, \quad (16)$$

where f_{dig} is the sampling rate of the digitizer.

In 2-D STEAM, the numbers of pixels along the two orthogonal dimensions are defined differently. In the x -direction, the number of pixels (N_{2D-x}) is essentially the number of columns in the spectral shower. In contrast, the number of pixels in the y -direction (N_{2D-y}) is the number of sampled point by the digitizer along each column of the spectral shower. Hence,

$$N_{2D-x} = \frac{\Delta \lambda_{SS}}{\Delta \lambda_{FSR}}, \quad (17)$$

$$N_{2D-y} = \Delta \lambda_{FSR} \cdot D \cdot f_{dig}. \quad (18)$$

Note that because of the serialization, only N_{2D-y} depends on the GVD, and the sampling rate. The total number of pixels N_{2D} is thus given by,

$$N_{2D} = N_x \cdot N_y = \Delta \lambda_{SS} \cdot D \cdot f_{dig} \leq \frac{f_{dig}}{f_{rep}}, \quad (19)$$

where f_{rep} is the repetition rate of the pulse laser, and hence is the frame rate of STEAM. From Eq. (19), we note that increasing GVD or the optical bandwidth can increase the frame length in time and hence the number of pixels. However, it comes at an expense of the laser repetition rate (i.e., the frame rate) in order to avoid the overlap between consecutive frames (i.e., $\Delta \lambda_{SS} \cdot D < f_{rep}^{-1}$). Fortunately, this limitation can be overcome by using a technique called virtual time gating, based on wavelength division multiplexing, to carve the single frame into multiple bands [23]. In such a parallel architecture, the number of pixels can be increased by the number of parallel channels (M) without sacrificing the frame rate (see Fig. 8),

$$N_{1D} = N_{2D} \leq M \cdot f_{dig} / f_{rep}. \quad (20)$$

As shown in Eqs. (19) and (20), the number of pixels can be scaled up by increasing the optical bandwidth, GVD, the digitizer sampling rate and the number of parallel channels for implementing virtual time gating. For example, it is practical with today's technology to achieve $\sim 300,000$ pixels for a frame rate of 1 MHz using an optical bandwidth of $\sim 150 \text{ nm}$, a dispersion of $\sim 5 \text{ ns/nm}$, a digitizer sampling rate of 50 GS/s, using eight parallel virtual time gating channels ($M = 8$) (Fig. 8).

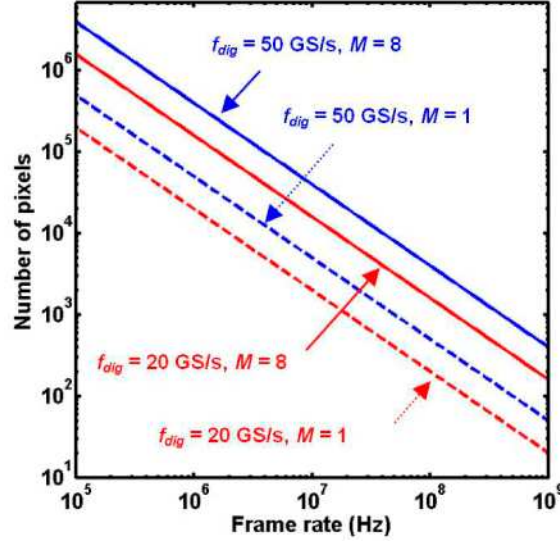


Fig. 8. Relationship between the number of pixels in STEAM and the frame rate based on Eq. (20). Note that it can be applied to both 1-D and 2-D STEAM. It is clear that the trade-off between the number of pixels and the frame rate can be overcome by employing virtual time gating with multiple channels ($M = 8$, solid lines).

5. Detection sensitivity

The detection sensitivity of STEAM is typically limited by a number of noise sources, namely the inherent shot noise of the input light (N_{shot}), the dark current noise (N_{dark}) and the thermal noise ($N_{thermal}$) of the photodetector. The shot noise is given by $N_{shot} = S_{in}^{-1/2}$, where S_{in} is the number of signal photoelectrons collected by the photodetector. Consider an optical amplification process with a gain of G and a noise figure of F , the resultant shot noise at the photodetector becomes $G \cdot F \cdot N_{shot}$ [24]. Hence, the total noise of the system is

$$N_{total} = \sqrt{(G \cdot F \cdot N_{shot})^2 + N_{dark}^2 + N_{thermal}^2}. \quad (21)$$

Figure 9(a) shows the individual noise component in the system as a function of the number of signal photon per pixel, which is defined as Φ / f_{dig} , where Φ is the signal photon flux. For a system without gain (i.e. $G = 1$), it is shot-noise-limited when the signal photon number $> \sim 1000$. Decrease in signal photon number ($< \sim 1000$) however would turn the system into thermal-noise-limited ($N_{dark} \ll N_{thermal}$ at room temperature). In contrast, the system becomes shot-noise-limited in the presence of optical amplification. The merit of optical amplification becomes more apparent by investigating the SNR of the STEAM system. With an optically amplified signal, i.e. $G \cdot S_{in}$, the SNR is given by

$$SNR = \frac{G \cdot S_{in}}{N_{total}} = \frac{S_{in}}{\sqrt{(F \cdot \sqrt{S_{in}})^2 + \left(\frac{N_{dark}}{G}\right)^2 + \left(\frac{N_{thermal}}{G}\right)^2}}, \quad (22)$$

where $S_{in} = \eta \cdot \Phi / f_{dig}$, where η is the quantum efficiency of the photodetector. Equation (22) shows that optical amplification effectively reduces both the dark current noise and the thermal noise by a factor of G , albeit the increase in shot noise by a factor of F . Hence, the SNR can be significantly enhanced if we implement the optical amplification with high gain and low noise-figure. Here, we employ distributed Raman amplification (DRA) within the dispersive fiber because DRA is well-known to have widely tunable and broadband gain spectrum, low noise figure, and the ability to maximize the SNR-to-distortion ratio by

keeping the signal power away from low power (noisy) and high power (nonlinear) regimes because of its distributed amplification nature [10–14].

The improvement in SNR as a result of optical amplification is depicted in Fig. 9(b). The optical amplification considerably enhances the SNR especially for the low light regime (i.e. $< \sim 1000$ photons per pixel) in which the system is originally thermal-noise-limited.

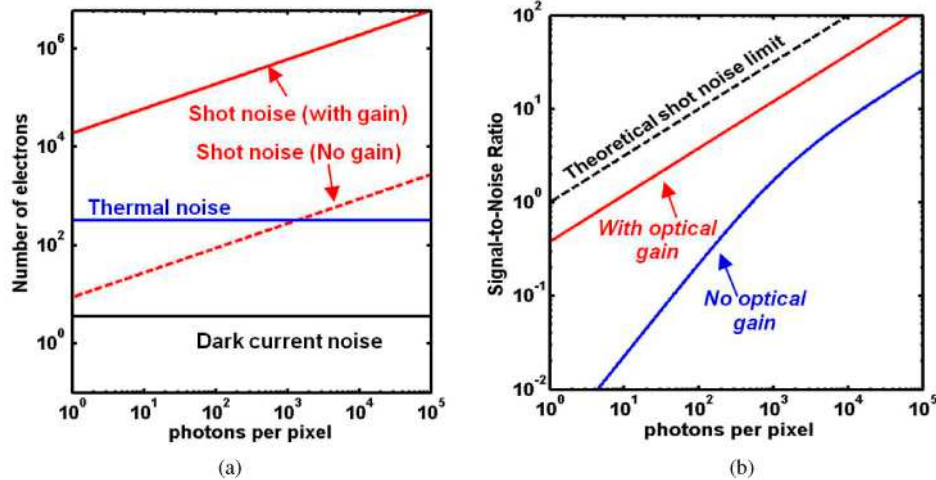


Fig. 9. (a) Noise components of STEAM: Dark current noise (black), thermal noise (blue), shot noise with gain, $G = 30$ dB (red solid line) and shot noise without gain (red dashed line). (b) SNR of the system with gain, $G = 30$ dB (red), and without gain (blue). The system parameters are: a photodetector with a bandwidth of 40 GHz, dark current of 100 nA, noise equivalent noise of $50 \text{ pW/Hz}^{1/2}$ and $\eta = 0.8$. The $f_{dig} = 50 \text{ GS/s}$, the wavelength is 800 nm. Detailed calculation of N_{dark} and $N_{thermal}$ can be referred to Ref [24]. We assume DRA is employed within the dispersive fiber and a noise figure of ~ 3.5 dB [25]. The dispersive fiber loss is assumed to be 20 dB. The black dashed line in (b) represents the theoretical shot noise limited SNR.

Furthermore, the system shows the minimum detectable number of signal photons is as low as ~ 7 for SNR = 1. It corresponds to an input referred noise of ~ -150 dBm/Hz at $\lambda = 800$ nm. It should be emphasized again such detection sensitivity improvement enables the ultrafast imaging, on the order of MHz, in STEAM.

6. Concluding remarks

In conclusion, we have presented a model that quantifies the spatial and temporal resolution of the recently demonstrated STEAM imaging technology. We have also quantified the imager's detection sensitivity. We have shown that the spatial resolution is not simply governed by the diffraction limit, but it also depends on the spectral resolutions imposed by (1) the spatial disperser, (2) the amplified dispersive Fourier Transform (ADFT), and the digitizer's sampling speed and input bandwidth. We have also shown that there is a trade-off between the number of pixels and the frame rate as a consequence of the image serialization. However, this can be remedied by a technique known as virtual time gating; an all-optical time demultiplexer that enables parallel detection. This analysis not only provides valuable insight into this new imaging system, it also serves as a tool for design and optimization of STEAM imaging systems.

Real-time, continuous and ultrafast operation of STEAM naturally meets the demands of many high-speed imaging applications which involve detecting fast events that are very rare, rogue events. Both the 1-D and 2-D STEAM configurations can find a myriad of applications in these areas. 2-D STEAM in its native form could play a significant role in non-invasive high-speed reflectance confocal imaging, which has been employed in clinical applications, e.g. to monitor fast response to therapeutic treatment such as laser surgery [26, 27].

In contrast, having a simpler spatial disperser implementation and less stringent requirement on temporal dispersion (see Fig. 3 and 7), 1-D STEAM could find a compelling application in high-speed and sensitive imaging flow-cytometry [28]. In this case, the 1-D STEAM imager, operating in the line scan mode, is able to reconstruct the 2-D cross sectional images of the cells as they flow in the microfluidic channel at a high flow rate. Current flow cytometers have no means to offer real-time and high-speed imaging that matches the throughput of state-of-the-art flow cytometers (i.e. up to 100,000 cells per second [28]). This is mostly because of the lack of a camera technology with sufficient combination of speed and sensitivity. Combined with flow cytometry, 1-D STEAM provides an attractive means of delivering real-time imaging of cells and potentially performing screening of rare cancer cells such as circulating tumor cells [29].

Acknowledgements

This work was partially supported by the Defense Advanced Research Projects Agency (DARPA).